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(71) Applicants (for all designated States except US): **MOR-
PHOSYS AG** [DE/DE]; Lena-Christ-Str. 48, 82152
Martinsried/Munich (DE). **PROCHON BIOTECH LTD.**
[IL/IL]; Kiryat Weizmann Science Park, P.O. Box 1482,
76114 Rehovot (IL).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **THOMASSEN-**

WOLF, Elisabeth [DE/DE]; Einsteinstr. 10, 82152 Mar-
tinsried (DE). **BORGES, Eric** [DE/DE]; Pentenriederstr.
32, 82152 Krailling (DE). **YAYON, Avner** [IL/IL];
Moshav Sitria # 104, 76834 (IL). **ROM, Eran** [IL/IL];
Isenberg Street 8b, Rehovot (IL).

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(54) Title: ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREEN-
ING FOR AND USES THEREOF

(57) Abstract: Molecules containing the antigen-binding portion of antibodies that block constitutive and/or ligand-dependent ac-
tivation of a receptor protein tyrosine kinase, such as fibroblast growth factor receptor 3 (FGFR3), are found through screening
methods, where a soluble dimeric form of a receptor protein tyrosine kinase is used as target for screening a library of antibody
fragments displayed on the surface of bacteriophage. The molecules of the present invention which block constitutive activation can
be administered to treat or inhibit skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor
progression associated with the constitutive activation of a receptor protein tyrosine kinase.



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ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREENING FOR AND USES THEREOF

Field of the Invention

5 The present invention relates to: immunoglobulins (and functional fragments thereof) useful for blocking activation of receptor protein tyrosine kinases, methods for screening for such immunoglobulins, compositions comprising said immunoglobulins and methods of using the same for treating or inhibiting disease, such as skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression.

10 Background of the Invention

A wide variety of biological processes involve complex cellular communication mechanisms. One of the primary means of continual exchange of information between cells and their internal and external environments is via the secretion and specific binding of
15 peptide growth factors. Growth factors exert pleiotropic effects and play important roles in oncogenesis and the development of multicellular organisms regulating cell growth, differentiation and migration. Many of these factors mediate their effects by binding to specific cell surface receptors. The ligand-activated receptors start an enzymatic signal transduction cascade from the cell membrane to the cell nucleus, resulting in specific gene
20 regulation and diverse cellular responses.

Protein Kinases

One of the key biochemical mechanisms of signal transduction involves the reversible phosphorylation of proteins, which enables regulation of the activity of mature proteins by altering their structure and function.

25 Protein kinases ("PKs") are enzymes that catalyze the phosphorylation of hydroxy groups on tyrosine, serine and threonine residues of proteins. The consequences of this seemingly simple activity are staggering; cell growth, differentiation and proliferation; eg., virtually all aspects of cell life in one way or another depend on PK activity. Furthermore, abnormal PK activity has been related to a host of disorders, ranging from relatively non-life
30 threatening diseases such as psoriasis to extremely virulent diseases such as glioblastoma.

The kinases fall largely into two groups, those specific for phosphorylating serine and threonine, and those specific for phosphorylating tyrosine. Some kinases, referred to as “dual specificity” kinases, are able to phosphorylate tyrosine as well as serine/threonine residues.

Protein kinases can also be characterized by their location within the cell. Some kinases are transmembrane receptor proteins capable of binding ligands external to the cell membrane. Binding the ligands alters the receptor protein kinase’s catalytic activity. Others are non-receptor proteins lacking a transmembrane domain and yet others are ecto-kinases that have a catalytic domain on the extracellular (ecto) portion of a transmembrane protein or which are secreted as soluble extracellular proteins.

Many kinases are involved in regulatory cascades where their substrates may include other kinases whose activities are regulated by their phosphorylation state. Thus, activity of a downstream effector is modulated by phosphorylation resulting from activation of the pathway.

Receptor protein tyrosine kinases (RPTKs) are a subclass of transmembrane-spanning receptors endowed with intrinsic, ligand-stimulatable tyrosine kinase activity. RPTK activity is tightly controlled. When mutated or altered structurally, RPTKs can become potent oncoproteins, causing cellular transformation. In principle, for all RPTKs involved in cancer, oncogenic deregulation results from relief or perturbation of one or several of the auto-control mechanisms that ensure the normal repression of catalytic domains. More than half of the known RPTKs have been repeatedly found in either mutated or overexpressed forms associated with human malignancies (including sporadic cases; Blume-Jensen et al., 2001). RPTK overexpression leads to constitutive kinase activation by increasing the concentration of dimers. Examples are Neu/ErbB2 and epidermal growth factor receptor (EGFR), which are often amplified in breast and lung carcinomas and the fibroblast growth factors (FGFR) associated with skeletal and proliferative disorders (Blume-Jensen et al., 2001).

Fibroblast Growth Factors

Normal growth, as well as tissue repair and remodeling, require specific and delicate control of activating growth factors and their receptors. Fibroblast Growth Factors (FGFs) constitute a family of over twenty structurally related polypeptides that are developmentally regulated and expressed in a wide variety of tissues. FGFs stimulate proliferation, cell migration and differentiation and play a major role in skeletal and limb development, wound

healing, tissue repair, hematopoiesis, angiogenesis, and tumorigenesis (reviewed in Ornitz and Itoh, 2001).

The biological action of FGFs is mediated by specific cell surface receptors belonging to the RPTK family of protein kinases. These proteins consist of an extracellular ligand binding domain, a single transmembrane domain and an intracellular tyrosine kinase domain which undergoes phosphorylation upon binding of FGF. The FGF receptor (FGFR) extracellular region contains three immunoglobulin-like (Ig-like) loops or domains (D1, D2 and D3), an acidic box, and a heparin binding domain. Five FGFR genes that encode for multiple receptor protein variants have been identified to date.

Another major class of cell surface binding sites includes binding sites for heparan sulfate proteoglycans (HSPG) that are required for high affinity interaction and activation of all members of the FGF family. Tissue-specific expression of heparan sulfate structural variants confer ligand-receptor specificity and activity of FGFs.

FGFR-Related Disease

Recent discoveries show that a growing number of skeletal abnormalities, including achondroplasia, the most common form of human dwarfism, result from mutations in FGFRs. Specific point mutations in different domains of FGFR3 are associated with autosomal dominant human skeletal disorders including hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) and thanatophoric dysplasia (TD) (Cappellen et al., 1999; Webster et al., 1997; Tavormina et al., 1999). FGFR3 mutations have also been described in two craniosynostosis phenotypes: Muenke coronal craniosynostosis (Bellus et al., 1996; Muenke et al., 1997) and Crouzon syndrome with acanthosis nigricans (Meyers et al., 1995). Crouzon syndrome is associated with specific point mutations in FGFR2 and both familial and sporadic forms of Pfeiffer syndrome are associated with mutations in FGFR1 and FGFR2 (Galvin et al., 1996; Schell et al., 1995). Mutations in FGFRs result in constitutive activation of the mutated receptors and increased receptor protein tyrosine kinase activity, rendering cells and tissue unable to differentiate. Specifically, the achondroplasia mutation results in enhanced stability of the mutated receptor, dissociating receptor activation from down-regulation, leading to restrained chondrocyte maturation and bone growth inhibition (reviewed in Vajo et al., 2000).

There is accumulating evidence for FGFR3 activating mutations in various types of cancer. Constitutively activated FGFR3 in a large proportion of two common epithelial

cancers, bladder and cervix, as well as in multiple myeloma, is the first evidence of an oncogenic role for FGFR3 in carcinomas. FGFR3 currently appears to be the most frequently mutated oncogene in bladder cancer where it is mutated in almost 50% of the cases and in about 70% of cases having recurrent superficial bladder tumors (Cappellen, et al, 1999; van Rhijn, et al, 2001; Billerey, et al, 2001). FGFR3 mutations are seen in 15-20% of multiple myeloma cases where point mutations that cause constitutive activation directly contribute to tumor development and progression (Chesi, et al, 1997; Jang, et al, 2000; Plowright, et al, 2000, Ronchetti, et al, 2001).

In this context, the consequences of FGFR3 signaling appear to be cell type-specific. In chondrocytes, FGFR3 hyperactivation results in growth inhibition (reviewed in Ornitz, 2001), whereas in the myeloma cell it contributes to tumor progression (Chesi et al., 2001).

In view of the link between RPTK-related cellular activities and a number of human disorders various strategies have been employed to target the receptors and/or their variants for therapy. Some of these have involved biomimetic approaches using large molecules patterned on those involved in the cellular processes, e.g., mutant ligands (US Patent 4,966,849); soluble receptors and antibodies (WO 94/10202, US 6,342,219); RNA ligands (US Patent 5,459,015) and tyrosine kinase inhibitors (WO 94/14808; US Patent 5,330,992).

Antibody therapy

The search for new therapies to treat cancer and other diseases associated with growth factors and their corresponding cell surface receptors has resulted in the development of humanized antibodies capable of inhibiting receptor function. For example, US patents 5,942,602 and 6,365,157 disclose monoclonal antibodies specific for the HER2/neu and VEGF receptors, respectively. US patent 5,840,301 discloses a chimeric, humanized monoclonal antibody that specifically binds to the extracellular domain of VEGF and neutralizes ligand-dependent activation.

There is an unmet need for highly selective molecules capable of blocking aberrant constitutive receptor protein tyrosine kinase activity, in particular FGFR activity, thereby addressing the clinical manifestations associated with the above-mentioned mutations, and modulating various biological functions.

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information available to applicant at the time of filing and does not constitute an admission as to the correctness of such a statement.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide molecules which are able to block receptor protein tyrosine kinase (RPTK) activity, preferably fibroblast growth factor receptor (FGFR) activity, and more preferably fibroblast growth factor receptor 3 (FGFR3) activity.

5 It is another object of the present invention to provide a method to screen for molecules which are able to block said receptor activity.

It is yet another object of the present invention to provide a pharmaceutical composition comprising as an active ingredient a molecule of the invention useful in treating or preventing skeletal and proliferative diseases and disorders.

10 It is a further object of the present invention to provide a method for inhibiting growth of tumor cells associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

15 It is yet a further object of the present invention to provide a method for treating skeletal disorders associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

It is yet a further object of the present invention to provide a method for blocking receptor protein tyrosine kinase activation in the cells of patients in need thereof by treatment with molecules capable of inhibiting receptor protein tyrosine kinase function.

20 It is still a further object of the invention to provide a kit containing molecules of the invention.

These and other objects are met by the invention disclosed herein.

25 The present invention provides a molecule that contains the antigen-binding portion of an antibody which blocks constitutive activation of a receptor protein tyrosine kinase. The present invention further provides a molecule that contains the antigen-binding portion of an antibody which blocks ligand-dependent activation of a fibroblast growth factor receptor (FGFR), including FGFR1 and FGFR3. Certain molecules of the present invention were found to inhibit or block constitutive, or ligand independent, activation of the FGFR3. Generation of inhibitory molecules would be useful for developing medicaments for use in

treating or preventing skeletal and proliferative diseases and disorders associated with constitutive activation of receptor protein tyrosine kinases.

Certain mutations in the genes of receptor protein tyrosine kinases result in activation of the receptor in a manner that is independent of the presence of a ligand. Such ligand-
5 independent or constitutive receptor protein tyrosine kinase activation results in increased receptor activity. The clinical manifestations of certain mutations are skeletal and proliferative disorders and diseases, including achondroplasia and various cancers.

Furthermore, the present invention is directed to novel molecules comprising an antigen
10 binding domain which binds to a receptor protein tyrosine kinase and blocks constitutive activation of said receptor protein tyrosine kinase. The molecules of the invention may be antibodies or antigen binding fragments thereof.

A currently preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of a receptor protein tyrosine kinase and blocks constitutive and ligand-dependent activation of the receptor.

15 A currently more preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of an FGF receptor and blocks constitutive and ligand-dependent activation of the receptor.

The invention provides multiple antibodies have characteristics as described herein, *e.g.*, the ability to bind FGFR3 and block constitutive and/or ligand-dependent activation of the
20 receptor. Such antibodies include, but are not limited to, the following: MSPRO 59 having a VH sequence of SEQ ID NO: 60 and a VL sequence of SEQ ID NO: 42, and the corresponding polynucleotide sequences of SEQ ID NO: 91 and SEQ ID NO: 76, respectively; MSPRO 21 having a VH sequence of SEQ ID NO: 47 and a VL sequence of SEQ ID NO: 36, and the corresponding polynucleotide sequences of SEQ ID NO: 78 and
25 SEQ ID NO: 67, respectively; MSPRO 24 having a VH sequence of SEQ ID NO: 48 and a VL sequence of SEQ ID NO: 33, and the corresponding polynucleotide sequences of SEQ ID NO: 79 and SEQ ID NO: 64, respectively; MSPRO 28 having a VH sequence of SEQ ID NO: 49 and a VL sequence of SEQ ID NO: 31, and the corresponding polynucleotide sequences of SEQ ID NO: 80 and SEQ ID NO: 62, respectively; MSPRO 02 having a VH
30 sequence of SEQ ID NO: 51 and a VL sequence of SEQ ID NO: 43, and the corresponding polynucleotide sequences of SEQ ID NO: 84 and SEQ ID NO: 74, respectively; MSPRO 11 having a VH sequence of SEQ ID NO: 52 and a VL sequence of SEQ ID NO: 38, and the

corresponding polynucleotide sequences of SEQ ID NO: 85 and SEQ ID NO: 70, respectively; MSPRO 26 having a VH sequence of SEQ ID NO: 53 and a VL sequence of SEQ ID NO: 39, and the corresponding polynucleotide sequences of SEQ ID NO: 86 and SEQ ID NO: 71, respectively; MSPRO 29 having a VH sequence of SEQ ID NO: 54 and a
5 VL sequence of SEQ ID NO: 34, and the corresponding polynucleotide sequences of SEQ ID NO: 87 and SEQ ID NO: 65, respectively; MSPRO 54 having a VH sequence of SEQ ID NO: 55 and a VL sequence of SEQ ID NO: 45, and the corresponding polynucleotide sequences of SEQ ID NO: 82 and SEQ ID NO: 73, respectively; MSPRO 55 having a VH sequence of SEQ ID NO: 56 and a VL sequence of SEQ ID NO: 40, and the corresponding
10 polynucleotide sequences of SEQ ID NO: 83 and SEQ ID NO: 69, respectively; and MSPRO 12 having a VH sequence of SEQ ID NO: 58 and a VL sequence of SEQ ID NO: 44, and the corresponding polynucleotide sequences of SEQ ID NO: 89 and SEQ ID NO: 75, respectively.

According to the principles of the present invention, molecules which bind FGFR, more
15 preferably FGFR3, and block ligand-dependent receptor activation are provided. These molecules are useful in treating hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, proliferative retinopathy including proliferative diabetic retinopathy, macular degeneration, hemangiomas, angiofibromas, and psoriasis.

20 In addition, the present invention also relates to methods for screening for the molecules according to the present invention by using a soluble dimeric form of a receptor protein tyrosine kinase as a target for screening a library of antibody fragments displayed on the surface of a bacteriophage

A further aspect of the present invention provides a pharmaceutical composition
25 comprising as an active ingredient a molecule of the present invention useful for preventing or treating skeletal or cartilage diseases or disorders and craniosynostosis disorders associated with constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

In a currently preferred embodiment the pharmaceutical compositions of the present
30 invention may be used for treating or preventing skeletal disorders associated with aberrant FGFR signaling, including achondroplasia, thanatophoric dysplasia, Apert or Pfeiffer syndrome and related craniosynostosis disorders.

A further aspect of the present invention provides a pharmaceutical composition comprising as an active ingredient a molecule of the present invention useful for preventing or treating cell proliferative diseases or disorders or tumor progression, associated with the constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

5 In a currently preferred embodiment the pharmaceutical compositions of the present invention may be used for treating or preventing proliferative diseases associated with aberrant FGFR signaling, including multiple myeloma, transitional cell carcinoma of the bladder, mammary and cervical carcinoma, chronic myeloid leukemia and osteo- or chondrosarcoma.

10 A still further aspect of the present invention provides methods for treating or inhibiting the aforementioned diseases and disorders by administering a therapeutically effective amount of a pharmaceutical composition comprising a molecule of the present invention to a subject in need thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows a Western blot of the purification of hFR3²³⁻³⁷⁴TDhis.

Figure 2 shows hFR3²³⁻³⁷⁴TDhis binding to heparin and FGF9.

Figure 3 shows the purification of FR3exFc and FR1exFc on SDS-PAGE.

- 5 Figure 4 shows the neutralization effect of the hFR3²³⁻³⁷⁴TDhis and FR3exFc soluble receptors in a ligand-dependent proliferation assay.

Figure 5 shows the effect of several MSPRO Fabs on proliferation of FGFR1 and FGFR3 expressing cells.

- 10 Figure 6 shows the effect of several MSPRO Fabs on proliferation of FGFR3 expressing cells.

Figures 7A and 7B show the neutralizing activity of several MSPRO Fabs in a proliferation assay using the FDCP-FR3 (C10; Fig. 7A) or the FDCP-FR1 cells (Fig. 7B).

- 15 Figure 8 shows the receptor specificity of MSPRO Fabs on RCJ-FR3 cells by Western blot using an anti-P-ERK antibody. Figure 8A shows different MSPRO Fabs while Figure 8B shows a dose response of MSPRO 12, 29 and 13 on RCJ-FR3 cells.

Figures 9A-9D demonstrates the specificity and potency of MS-PRO Fabs by Western blot with anti-P-ERK antibody.

- 20 Figure 10 shows a diagrammatic representation of FGFR3 and of FGFR3 truncations (D2-3, D2) and isoforms (IIIb, IIIc). The isoform IIIb differs from IIIc at the carboxy terminus of the IgIII domain as indicated with a dotted line.

Figure 11 shows that the FGFR3 neutralizing Fabs recognize IgII or IgII and III in the extracellular region of FGFR3.

Figure 12 shows that MSPRO29 specifically recognizes the IIIc isoform of FGFR3.

- 25 Figure 13 shows the results of a proliferation assay for FDCP-FR3IIIb or FDCP-FR3IIIc cells incubated with increasing dose of the indicated Fabs.

Figure 14 shows iodinated MSPRO29 binding to FGFR3.

Figure 15 shows results of a proliferation assay is a graph wherein iodinated MSPRO29 retained its activity against FGFR3.

- 30 Figures 16A-16F show the selective binding of radiolabelled MS-PRO29 to histological sections of growth plate.

Figure 17 shows a proliferation assay of FDCP-FR3 (C10) and FDCP-FR3ach cells incubated with FGF9 and with increasing doses of the indicated Fabs.

Figure 18A shows analysis of the ligand-dependent FDCP-FR3wt cells.

Figure 18B shows that MSPRO12 and MSPRO59 inhibit the ligand independent proliferation of FDCP-FR3ach cells.

Figure 19 shows the restoration of cell growth in RCS cells by MS-PRO54 and MSPRO59..

Figure 20 represents the growth rate of treated bone with MS-PRO 59.

Figure 21 is a flow chart of the experimental protocol for assessing receptor activation and signaling.

10 Figure 22 shows I¹²⁵MSPRO59 localization to the FDCP-FR3ach derived tumor.

Figure 23 shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth after 4 and 24 hours.

Figure 24 shows Binding of Fab Miniantibodies to FGFR3-Fc and FGFR1-Fc (ELISA).

15 Figure 25A is an example of a Fab expression vector for use in accordance with the present invention.

Figure 25B is the DNA sequence of the vector according to Figure 25A

Figure 26A is an example of a phage display vector for use in accordance with the present invention.

Figure 26B is the DNA and amino acid sequence of the vector according to Figure 29A.

20 Figure 27A is a description of the DNA sequence for V_H and V_L regions of certain antibodies of the invention.

Figure 27B is a description of the amino acid sequence for V_H and V_L regions of certain antibodies of the invention.

25

DETAILED DESCRIPTION OF THE INVENTION

The present invention is based on the discovery that neutralizing antibodies that block ligand-dependent and ligand-independent activation of fibroblast growth factor 3 (FGFR3), a receptor protein tyrosine kinase (RPTK), can be obtained by screening a synthetic antibody library against a soluble dimeric form of the extracellular portion of FGFR3. Until the present invention, the present inventors are unaware of any success in obtaining neutralizing antibodies that block FGFR3 activation.

RPTKs comprise a large family of proteins, expressed in a spatially and temporally restricted manner, that regulate many different aspects of growth and development. When the restraint is removed, as in certain mutations, and the receptors are over activated, a diseased state may arise. Molecules, including antibodies and fragments thereof, comprising an antigen binding domain to a receptor protein tyrosine kinase are highly necessary for the treatment of various pathological conditions.

In the past, the laboratory of the present inventors found it very difficult to raise neutralizing antibodies against FGFR3. When mice were immunized with the soluble monomeric FGFR3 receptor, by the time the antibody titers begins to increase, the mice died. The experiments performed in the laboratory of the present inventors that failed to obtain anti-FGFR3 neutralizing antibodies in mice are presented in the Examples.

By using a soluble dimeric form of the extracellular domain of the FGFR3 receptor to screen for antibodies, e.g., Fabs, that bind from a phage display antibody library, the present inventors were able to overcome a problem in the prior art for which there was yet no solution and to obtain numerous high affinity ($K_D < 50$ nM; preferably < 20 nM) antibodies (Fabs) that bind FGFR3 and interfere with ligand binding, thereby blocking ligand-dependent activation of FGFR3 signaling. Very surprisingly, from among the group of Fabs that block ligand-dependent activation, Fab antibodies which also block ligand-independent (constitutive) activation of FGFR3 by blocking signaling caused by constitutive dimerization of FGFR3 were identified. To the best of the present inventors' knowledge, the Fab antibodies obtained which block constitutive activation of FGFR3 are the first antibodies against any receptor protein tyrosine kinase that blocks constitutive, ligand-independent activation/signaling.

Trastuzumab, an anti-human epidermal growth factor receptor 2 (HER2) antibody, was the first humanized monoclonal antibody approved for therapeutic use. Its mode of action

appears to be manifold, including HER2 down regulation, prevention of heterodimer formation, prevention of HER2 cleavage and others (Baselga and Albanell, 2001). US patents 5, 677171; 5772997; 6165464 and 6,399,063 disclose the anti-HER2 invention but neither teach nor suggest that the antibody blocks ligand-independent receptor activation.

5 One aspect of the present invention is directed to neutralizing antibodies and more generally to a molecule that includes the antigen binding portion of an antibody which blocks ligand-dependent activation and constitutive, ligand-independent activation of a receptor protein tyrosine kinase, preferably an FGFR and more preferably FGFR3.

10 Another aspect of the present invention is directed to molecules comprising an antigen binding domain which blocks ligand-dependent activation of an FGFR, more preferably FGFR3.

15 The molecule having the antigen-binding portion of an antibody according to the present invention can be used in a method for blocking the ligand-dependent activation and/or ligand independent (constitutive) activation of FGFR3. Preferred embodiments of such antibodies/molecules, designated as HuCAL[®] (Human Combinatorial Antibody Library) clone, is presented in Table 1 with the unique VH-CDR3 and VL-CDR3 sequences given.

TABLE 1A

HuCAL[®] - Clone	VH-CDR3 Sequence	VL-CDR3 equence	Framework
MSPRO- 2	DFLGYEFDY (SEQ ID NO: 8)	QSYDYSADY (SEQ IDNO: 9)	VH1B_L3
MSPRO- 11	YYGSSLYHYVFGGFIDY (SEQ ID NO: 10)	QSHHFYE (SEQ ID NO: 11)	VH1B_L2
MSPRO- 12	YHSWYEMGYYGSTVGYMFD (SEQ ID NO: 12)	QSYDFDFA (SEQ ID NO: 13)	VH2_L3
MSPRO- 21	DNWFKPFSDV (SEQ ID NO: 14)	QQYDSIPY (SEQ ID NO: 15)	VH1A_k4
MSPRO- 24	VNHWTYTFDY (SEQ ID NO: 16)	QQMSNYPD (SEQ ID NO: 17)	VH1A_k3
MSPRO- 26	GYWYAYFTYINYGYFDN (SEQ ID NO: 18)	QSYDNNSDV (SEQ ID NO: 19)	VH1B_L2
MS- PRO-28	GGGWVSHGYYYLFDL (SEQ ID NO: 26)	FQYGSIPP (SEQ ID NO: 27)	VH1A_k1
MSPRO- 29	TWQYSYFYLDGGYYFDI (SEQ ID NO: 20)	QQTNNAPV (SEQ ID NO:21)	VH1B_k3
MSPRO- 54	NMAYTNYQYVNMPHFDY (SEQ ID NO: 22)	QSYDYFKL (SEQ ID NO:23)	VH1B_L3
MSPRO- 55	SMNSTMYWYLRRVLFDH (SEQ ID NO: 28)	QSYDMYMYI (SEQ ID NO: 29)	VH1B_L2
MSPRO- 59	SYYPDFDY (SEQ ID NO:24)	QSYDGPDW (SEQ ID NO:25)	VH6_L3

Table 1B

Clone	Affinity to FGFR3 (BIAcore)	Affini ty to FGFR3 (FACS)	Affini ty to FGFR1	K _{off} (s ⁻¹)	IC ₅₀ FR3 (FGF9)	Domain Specifici ty	Ligand independent inhibition of FGFR3	Available formats
MS-PRO59	1.5nM	<1nM	-	7.1x10e-4	19 nM	2	+	Fab, Fab-dHLX IgG1, IgG4, mIgG3, scFv
MS-PRO-2	37nM	43 nM	-	2x10e-2	360 nM	2	~	Fab (+/- tags), Fab- dHLX, IgG1, IgG4,
MS-PRO- 12	14nM	6.5 nM	-	2.3x10e-3	58 nM	2	+	Fab (+/- tag), Fab- dHLX, IgG1, IgG4, scFv
MS-PRO- 11	4	4 nM	108	4 x 10e-4	220 nM	3		Fab. Fab-dHLX
MS-PRO- 21	9 nM		-	3.6x10e-3	50 nM	3c		Fab, IgG1, Fab-dHLX
MS-PRO- 24	10 nM		-	5.4x10e-3	70 nM	3c		Fab, IgG1
MS-PRO- 26	4 nM	1.4		5 x 10e-4	70 nM	3		Fab, Fab-dHLX
MS-PRO- 28	9 nM	0.3 nM	160 nM	4 x10e-3	50 nM	3		Fab
MS-PRO 29	6 nM	<1nM	29 nM	1.4x10e-3	20nM	3c	-	Fab (+/- tag), IgG1, IgG4, Fab-dHLX,

								scFv
MS-PRO-54	3.7nM		2.5nM	2x10e-3	45nM	3c		Fab, IgG1
MS-PRO55	2.9nM		-	7.4x10e-4	34nM	3c		Fab

Table 1C

(BIAcor results for certain molecules)

IC50S:

- 5 (Under the "IC50" column, the numbers on the left indicate the dimeric dHLX format of the respective binder in the FDCP1-FGFR3 proliferation assay performed with FGF9. The numbers in parentheses are the IC50s of the monomeric Fabs in the same assay.)

binder	IC50 (FR3)
MS-Pro-2	61nM (360)
MS-Pro-12	26nM (58)
MS-Pro-21	20nM (50)
MS-Pro-26	8nM (70)

Table 1D**K_D determination for certain molecules**

Clone	BIAcore K _D [nM]	Number of measurements
MS-Pro-2-dHLX-MH	4.3 (37)	1
MS-Pro-11-dHLX-MH	0.7 (4)	1
MS-Pro-12-dHLX-MH	1.2 (14)	1
MS-Pro-21-dHLX-MH	2.2 (4.1)	1
MS-Pro-24-dHLX-MH	2 (10)	1
MS-Pro-26-dHLX-MH	2 (4)	1
MS-Pro-28-dHLX-MH	1.6 (9)	1

(Under the “BIAcore” column, the numbers on the left indicate the dimeric dHLX format of the respective binder in the FDCP1-FGFR3 proliferation assay performed with FGF9. The numbers in parentheses are the BIAcore measurements of the monomeric Fabs in the same assay).

The preferred, but non-limiting, embodiments of molecules according to the present invention that block constitutive (ligand-independent) activation of FGFR3 are referred to herein MS-PRO-2, MS-PRO-12 and MS-PRO-59 comprising VH-CDR3 and VL-CDR3 domains having SEQ ID Nos: 8 and 9; 12 and 13; and 24 and 25, respectively. The preferred, but non-limiting, embodiments of molecules according to the present invention that block ligand-dependent activation of FGFR3 are referred to herein MS-PRO-11, MS-PRO-21, MS-PRO-24, MS-PRO-26, MS-PRO-29, and MS-PRO-54 comprising VH-CDR3 and VL-CDR3 domains having SEQ ID Nos: 10 and 11; 14 and 15; 16 and 17, 18 and 19; 21 and 22; 23 and 24, respectively. Preferably, an antibody or a molecule of the present invention has an affinity (K_D) for binding a soluble dimeric form of FGFR3 of less than about 50 nM, preferably less than about 30 nM and more preferably less than about 10 nM, as determined by the BIAcore chip assay for affinity, by a FACS-Scatchard analysis or other methods known in the art.

While the specific discovery of an antibody/molecule that blocks constitutive activation was made with respect to the FGFR3 receptor using a soluble dimeric form of FGFR3 to screen a phage display antibody library, it is believed that for all, or almost all receptor protein tyrosine kinases, antibodies/molecules that block constitutive activation can be

similarly obtained using a soluble dimeric form of a corresponding extracellular domain of a receptor protein tyrosine kinase. Non-limiting examples of receptor protein tyrosine kinases disclosed in Blume-Jensen et al. (2001) include EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR- α , PDGFR- β , CSF-1R, kit/SCFR, Flk2/FH3, 5 Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROSA1k, Ryk, DDR, LTK and MUSK.

Furthermore, antibodies/molecules that block ligand-dependent or ligand independent activation of heterodimer receptor protein tyrosine kinases are intended to be included in the 10 scope of the invention. Heterodimerization is well documented for members of the EGFR subfamily of receptor protein tyrosine kinases and others. Non-limiting examples include EGFR/PDGFR β , Flt1/KDR and EGFR/ErbB2 heterodimers. EGFR and PDGFR β heterodimers have been identified as a mechanism for PDGF signal transduction in rat vascular smooth muscle cells (Saito et al., 2001) and Flt-1/KDR heterodimers are required 15 for vinculin assembly in focal adhesions in response to VEGF signaling (Sato et al., 2000).

The present invention is also directed to a molecule having the antigen-binding portion of an antibody which binds to a dimeric form of an extracellular portion of a receptor protein tyrosine kinase (RPTK), such as a FGFR, and blocks the ligand-independent (constitutive) activation and/or ligand-dependent activation of the RPTK.

20 Antibodies

Antibodies, or immunoglobulins, comprise two heavy chains linked together by disulfide bonds and two light chains, each light chain being linked to a respective heavy chain by disulfide bonds in a "Y" shaped configuration. Proteolytic digestion of an antibody yields three domains, Fv (Fragment variable), Fab (fragment antigen binding) and Fc 25 (fragment crystalline). The antigen binding domains, Fab', include regions where the polypeptide sequence varies. The term F(ab')₂ represents two Fab' arms linked together by disulfide bonds. The central axis of the antibody is termed the Fc fragment, while Fv comprises the variable heavy (VH) and variable light (VL) domains. Each heavy chain has at one end a variable domain (VH) followed by a number of constant domains (CH). Each light 30 chain has a variable domain (VL) at one end and a constant domain (CL) at its other end, the light chain variable domain being aligned with the variable domain of the heavy chain and the light chain constant domain being aligned with the first constant domain of the heavy chain (CH1).

The variable domains of each pair of light and heavy chains form the antigen binding site. The domains on the light and heavy chains have the same general structure and each domain comprises four framework regions, whose sequences are relatively conserved, joined by three hypervariable domains known as complementarity determining regions (CDR1-3).

5 These domains contribute specificity and affinity of the antigen binding site.

The isotype of the heavy chain (gamma, alpha, delta, epsilon or mu) determines immunoglobulin class (IgG, IgA, IgD, IgE or IgM, respectively). The light chain is either of two isotypes (kappa, κ or lambda, λ) found in all antibody classes.

It should be understood that when the terms "antibody" or "antibodies" are used, this is
10 intended to include intact antibodies, such as polyclonal antibodies or monoclonal antibodies (mAbs), as well as proteolytic fragments thereof such as the Fab or F(ab')₂ fragments. Further included within the scope of the invention are chimeric antibodies; human and humanized antibodies; recombinant and engineered antibodies, and fragments thereof. Furthermore, the DNA encoding the variable region of the antibody can be inserted into the DNA encoding
15 other antibodies to produce chimeric antibodies (see, for example, U.S. Patent 4,816,567). Single chain antibodies fall within the scope of the present invention. Single chain antibodies can be single chain composite polypeptides having antigen binding capabilities and comprising amino acid sequences homologous or analogous to the variable regions of an immunoglobulin light and heavy chain (linked VH-VL or single chain Fv (ScFv)). Both V_H
20 and V_L may copy natural monoclonal antibody sequences or one or both of the chains may comprise a CDR-FR construct of the type described in U.S. Patent 5,091,513, the entire contents of which are hereby incorporated herein by reference. The separate polypeptides analogous to the variable regions of the light and heavy chains are held together by a polypeptide linker. Methods of production of such single chain antibodies, particularly
25 where the DNA encoding the polypeptide structures of the V_H and V_L chains are known, may be accomplished in accordance with the methods described, for example, in U.S. Patents 4,946,778, 5,091,513 and 5,096,815, the entire contents of each of which are hereby incorporated herein by reference.

Additionally, CDR grafting may be performed to alter certain properties of the antibody molecule including affinity or specificity. A non-limiting example of CDR grafting is disclosed in US patent 5,225,539.

5 A "molecule having the antigen-binding portion of an antibody" as used herein is intended to include not only intact immunoglobulin molecules of any isotype and generated by any animal cell line or microorganism, but also the antigen-binding reactive fraction thereof, including, but not limited to, the Fab fragment, the Fab' fragment, the F(ab')₂ fragment, the variable portion of the heavy and/or light chains thereof, Fab miniantibodies (see WO 93/15210, U.S. Patent application 08/256,790, WO 96/13583, U.S. Patent
10 application 08/817,788, WO 96/37621, U.S. Patent application 08/999,554, the entire contents of which are incorporated herein by reference) and chimeric or single-chain antibodies incorporating such reactive fraction, as well as any other type of molecule or cell in which such antibody reactive fraction has been physically inserted, such as a chimeric T-cell receptor or a T-cell having such a receptor, or molecules developed to deliver therapeutic
15 moieties by means of a portion of the molecule containing such a reactive fraction. Such molecules may be provided by any known technique, including, but not limited to, enzymatic cleavage, peptide synthesis or recombinant techniques.

The term "Fc" as used herein is meant as that portion of an immunoglobulin molecule (Fragment crystallizable) that mediates phagocytosis, triggers inflammation and targets Ig to
20 particular tissues; the Fc portion is also important in complement activation.

In one embodiment of the invention, a chimera comprising a fusion of the extracellular domain of the RPTK and an immunoglobulin constant domain can be constructed useful for assaying for ligands for the receptor and for screening for antibodies and fragments thereof.

The "extracellular domain" when used herein refers the polypeptide sequence of the
25 RPTKs disclosed herein which are normally positioned to the outside of the cell. The extracellular domain encompasses polypeptide sequences in which part of or all of the adjacent (C-terminal) hydrophobic transmembrane and intracellular sequences of the mature RPTK have been deleted. Thus, the extracellular domain-containing polypeptide can comprise the extracellular domain and a part of the transmembrane domain. Alternatively, in
30 the preferred embodiment, the polypeptide comprises only the extracellular domain of the RPTK. The truncated extracellular domain is generally soluble. The skilled practitioner can readily determine the extracellular and transmembrane domains of a RPTK by aligning the

RPTK of interest with known RPTK amino acid sequences for which these domains have been delineated. Alternatively, the hydrophobic transmembrane domain can be readily delineated based on a hydrophobicity plot of the polypeptide sequence. The extracellular domain is N-terminal to the transmembrane domain.

5 The term "epitope" is meant to refer to that portion of any molecule capable of being bound by an antibody or a fragment thereof which can also be recognized by that antibody. Epitopes or antigenic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and have specific three-dimensional structural characteristics as well as specific charge characteristics.

10 An "antigen" is a molecule or a portion of a molecule capable of being bound by an antibody which is additionally capable of inducing an animal to produce antibody capable of binding to an epitope of that antigen. An antigen may have one or more than one epitope. The specific reaction referred to above is meant to indicate that the antigen will react, in a highly selective manner, with its corresponding antibody and not with the multitude of other
15 antibodies which may be evoked by other antigens.

 A monoclonal antibody (mAb) is a substantially homogeneous population of antibodies to a specific antigen. MAbs may be obtained by methods known to those skilled in the art. See, for example Kohler et al (1975); USP 4,376,110; Ausubel et al (1987-1999); Harlow et al (1988); and Colligan et al (1993), the contents of which references are incorporated
20 entirely herein by reference. The mAbs of the present invention may be of any immunoglobulin class including IgG, IgM, IgE, IgA, and any subclass thereof. The hybridoma producing the mAbs of this invention may be cultivated *in vitro* or *in vivo*. High titers of mAbs can be obtained in *in vivo* production where cells from the individual hybridomas are injected intraperitoneally into pristane-primed Balb/c mice to produce ascites
25 fluid containing high concentrations of the desired mAbs. MAbs of isotype IgM or IgG may be purified from such ascites fluids, or from culture supernatants, using column chromatography methods well known to those of skill in the art.

 Chimeric antibodies are molecules, the different portions of which are derived from different animal species, such as those having a variable region derived from a murine mAb
30 and a human immunoglobulin constant region. Antibodies which have variable region framework residues substantially from human antibody (termed an acceptor antibody) and complementarity determining regions substantially from a mouse antibody (termed a donor

antibody) are also referred to as humanized antibodies. Chimeric antibodies are primarily used to reduce immunogenicity in application and to increase yields in production, for example, where murine mAbs have higher yields from hybridomas but higher immunogenicity in humans, such that human/murine chimeric mAbs are used. Chimeric antibodies and methods for their production are known in the art (Better et al, 1988; Cabilly et al, 1984; Harlow et al, 1988; Liu et al, 1987; Morrison et al, 1984; Boulianne et al, 1984; Neuberger et al, 1985; Sahagan et al (1986); Sun et al, 1987; Cabilly et al, European Patent Application 125023 (published November 14, 1984); Taniguchi et al, European Patent Application 171496 (published February 19, 1985); Morrison et al, European Patent Application 173494 (published March 5, 1986); Neuberger et al, PCT Application WO 8601533, (published March 13, 1986); Kudo et al, European Patent Application 184187 (published June 11, 1986); Morrison et al., European Patent Application 173494 (published March 5, 1986); and Robinson et al., International Patent Publication WO 9702671 (published May 7, 1987) Queen et al., (1989) and WO 90/07861, U.S. Patent 5,693,762, U.S. Patent 5,693,761, U.S. Patent 5,585,089, U.S. Patent 5,530,101 and Winter, U.S. Patent 5,225,539, and WO 92/22653. These references are hereby incorporated by reference.

Besides the conventional method of raising antibodies *in vivo*, antibodies can be produced in vitro using phage display technology. Such a production of recombinant antibodies is much faster compared to conventional antibody production and they can be generated against an enormous number of antigens. In contrast, in the conventional method, many antigens prove to be non-immunogenic or extremely toxic, and therefore cannot be used to generate antibodies in animals. Moreover, affinity maturation (i.e., increasing the affinity and specificity) of recombinant antibodies is very simple and relatively fast. Finally, large numbers of different antibodies against a specific antigen can be generated in one selection procedure. To generate recombinant monoclonal antibodies one can use various methods all based on phage display libraries to generate a large pool of antibodies with different antigen recognition sites. Such a library can be made in several ways: One can generate a synthetic repertoire by cloning synthetic CDR3 regions in a pool of heavy chain germline genes and thus generating a large antibody repertoire, from which recombinant antibody fragments with various specificities can be selected. One can use the lymphocyte pool of humans as starting material for the construction of an antibody library. It is possible to construct naive repertoires of human IgM antibodies and thus create a human library of large diversity. This method has been widely used successfully to select a large number of

antibodies against different antigens. Protocols for bacteriophage library construction and selection of recombinant antibodies are provided in the well-known reference text Current Protocols in Immunology, Colligan et al (Eds.), John Wiley & Sons, Inc. (1992-2000), Chapter 17, Section 17.1.

5 Another aspect of the present invention is directed to a method for screening for the antibody or molecule of the present invention by screening a library of antibody fragments displayed on the surface of bacteriophage, such as disclosed in the Example herein and described in WO 97/08320, U.S. Patent 6,300,064 , and Knappik et al. (2000), for binding to a soluble dimeric form of a receptor protein tyrosine kinase. An antibody fragment which
10 binds to the soluble dimeric form of the RPTK used for screening is identified as a candidate molecule for blocking ligand-dependent activation and/or constitutive activation of the RPTK in a cell. Preferably the RPTK of which a soluble dimeric form is used in the screening method is a fibroblast growth factor receptor (FGFR), and most preferably FGFR3.

As a first screening method, the soluble dimeric form of a receptor tyrosine kinase can
15 be constructed and prepared in a number of different ways. For instance the extracellular domain of a RPTK joined to Fc and expressed as a fusion polypeptide that can dimerize naturally by means of the Fc portion of the RPTK-Fc fusion. Other suitable types of constructs of FGFR3, serving as guidance for other RPTKs, are disclosed in the Examples presented herein.

20 The assays for determining binding of antibody fragments to FGFR3, binding affinities, inhibition of cell proliferation, etc., are also described in the Example herein below.

The term "cell proliferation" refers to the rate at which a group of cells divides. The number of cells growing in a vessel can be quantified by a person skilled in the art when that person visually counts the number of cells in a defined volume using a common light
25 microscope. Alternatively, cell proliferation rates can be quantified by laboratory apparatus that optically or conductively measure the density of cells in an appropriate medium.

A second screen for antibody fragments as candidate molecules can be done using cells which have very high overexpression of the RPTK, such as for instance RCJ-M15 cells overexpressing mutant (achondroplasia) FGFR3. In cells expressing very high levels of
30 receptor some ligand-independent activation occurs even without the presence of a mutation, such as a constitutive activation mutation. It is believed that RPTK overexpression forces RPTKs to dimerize and signal even in the absence of ligand. These cells have monomeric

receptors as well as dimeric receptors present on their cell surface. Using this type of cell, one of skill in the art would be able to identify all different kinds of antibodies, i.e., blocking ligand-dependent activation, blocking constitutive activation, blocking activation and binding only to monomeric form, etc.

5 A third screen can be carried out on a cell line expressing a RPTK carrying a mutation, such as the FDCP-FR3ach line expressing the achondroplasia mutation in FGFR3. The FDCP-FR3ach cells become constitutively active, e.g. are able to signal in the absence of a ligand as determined by ERK phosphorylation.

10 A further aspect of the present invention relates to a method for treating or inhibiting a skeletal dysplasia or craniosynostosis disorder associated with constitutive activation of a RPTK which involves administering the molecule of the present invention to a subject need thereof. Non-limiting examples of skeletal dysplasias include achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, and severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia. Non-limiting examples of craniosynostosis
15 disorder are Muenke coronal craniosynostosis and Crouzon syndrome with acanthosis nigricans.

20 The present invention also provides for a method for treating or inhibiting a cell proliferative disease or disorder associated with the action of an abnormal constitutively activated RPTK, *e.g.*, tumor progression where the tumor progression includes progression of transitional cell carcinoma, osteo or chondrosarcoma, multiple myeloma, and mammary carcinoma (one of the known RPTKs involved in mammary carcinoma is ErbB3). A molecule containing the antigen binding portion of an antibody that blocks constitutive activation of a RPTK is administered to a subject in need thereof to treat or inhibit such a cell proliferative disease or disorder.

25 In another aspect of the present invention, molecules which bind FGFR, more preferably FGFR3, and block ligand-dependent receptor activation are provided. These molecules are useful in treating hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, proliferative retinopathy including proliferative diabetic retinopathy, macular degeneration, hemangiomas, angiofibromas, and
30 psoriasis. The role of FGFs and their receptors in neo- and hypervascularization has been well documented (Frank, 1997; Gerwins et al, 2000).

In another aspect of the present invention, the pharmaceutical compositions according to the present invention is similar to those used for passive immunization of humans with other antibodies. Typically, the molecules of the present invention comprising the antigen binding portion of an antibody will be suspended in a sterile saline solution for therapeutic uses. The pharmaceutical compositions may alternatively be formulated to control release of active ingredient (molecule comprising the antigen binding portion of an antibody) or to prolong its presence in a patient's system. Numerous suitable drug delivery systems are known and include, e.g., implantable drug release systems, hydrogels, hydroxymethylcellulose, microcapsules, liposomes, microemulsions, microspheres, and the like. Controlled release preparations can be prepared through the use of polymers to complex or adsorb the molecule according to the present invention. For example, biocompatible polymers include matrices of poly(ethylene-co-vinyl acetate) and matrices of a polyanhydride copolymer of a stearic acid dimer and sebacic acid (Sherwood et al, 1992). The rate of release molecule according to the present invention, i.e., of an antibody or antibody fragment, from such a matrix depends upon the molecular weight of the molecule, the amount of the molecule within the matrix, and the size of dispersed particles (Saltzman et al., 1989 and Sherwood et al., 1992). Other solid dosage forms are described in (Ansel et al., 1990 and Gennaro, 1990).

The pharmaceutical composition of this invention may be administered by any suitable means, such as orally, intranasally, subcutaneously, intramuscularly, intravenously, intra-arterially, or parenterally. Ordinarily, intravenous (i.v.) or parenteral administration will be preferred.

It will be apparent to those of ordinary skill in the art that the therapeutically effective amount of the molecule according to the present invention will depend, *inter alia* upon the administration schedule, the unit dose of molecule administered, whether the molecule is administered in combination with other therapeutic agents, the immune status and health of the patient, the therapeutic activity of the molecule administered and the judgment of the treating physician. As used herein, a "therapeutically effective amount" refers to the amount of a molecule required to alleviate one or more symptoms associated with a disorder being treated over a period of time.

Although an appropriate dosage of a molecule of the invention varies depending on the administration route, age, body weight, sex, or conditions of the patient, and should be determined by the physician in the end, in the case of oral administration, the daily dosage can generally be between about 0.01-200 mg, preferably about 0.01-10 mg, more preferably

about 0.1-10 mg, per kg body weight. In the case of parenteral administration, the daily dosage can generally be between about 0.001-100 mg, preferably about 0.001-1 mg, more preferably about 0.01-1 mg, per kg body weight. The daily dosage can be administered, e.g., in 1-4 regimens. Various considerations in arriving at an effective amount are described, e.g.,
5 in Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th ed., Pergamon Press, 1990; and Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, Pa., 1990.

The molecule of the present invention as an active ingredient is dissolved, dispersed or admixed in an excipient that is pharmaceutically acceptable and compatible with the active
10 ingredient as is well known. Suitable excipients are, for example, water, saline, phosphate buffered saline (PBS), dextrose, glycerol, ethanol, or the like and combinations thereof. Other suitable carriers are well-known to those in the art. (See, for example, Ansel et al., 1990 and Gennaro, 1990). In addition, if desired, the composition can contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents.

15 Combination therapy

The combined treatment of one or more of the molecules of the invention with an anti-neoplastic or anti-chemotherapeutic drug such as doxorubicin, cisplatin or taxol provides a more efficient treatment for inhibiting the growth of tumor cells than the use of the molecule by itself. In one embodiment, the pharmaceutical composition comprises the antibody and
20 carrier with an anti-chemotherapeutic drug.

The present invention also provides for a nucleic acid molecule, which contains a nucleotide sequence encoding the molecule having the antigen binding portion of an antibody that blocks ligand-dependent activation and/or constitutive activation of a receptor protein tyrosine kinase such as FGFR3, and a host cell transformed with this nucleic acid
25 molecule. Furthermore, also within the scope of the present invention is a nucleic acid molecule containing a nucleotide sequence having at least 90% sequence identity, preferably about 95%, and more preferably about 97% identity to the above encoding nucleotide sequence as would well understood by those of skill in the art.

The invention also provides nucleic acids that hybridize under high stringency
30 conditions to the VH and/or VL sequences of antibodies selected from the group consisting of MSPRO 21, MSPRO 24, MSPRO28, MSPRO 02, MSPRO 11, MSPRO 26, MSPRO 29, MSPRO 54, MSPRO 55, and MSPRO 12; or the complement; or a fragment thereof. As

used herein, highly stringent conditions are those which are tolerant of up to about 5-20% sequence divergence, preferably about 5-10%. Without limitation, examples of highly stringent (-10°C below the calculated T_m of the hybrid) conditions use a wash solution of 0.1 X SSC (standard saline citrate) and 0.5% SDS at the appropriate T_i below the calculated T_m of the hybrid. The ultimate stringency of the conditions is primarily due to the washing conditions, particularly if the hybridization conditions used are those which allow less stable hybrids to form along with stable hybrids. The wash conditions at higher stringency then remove the less stable hybrids. A common hybridization condition that can be used with the highly stringent to moderately stringent wash conditions described above is hybridization in a solution of 6 X SSC (or 6 X SSPE), 5 X Denhardt's reagent, 0.5% SDS, 100 µg/ml denatured, fragmented salmon sperm DNA at an appropriate incubation temperature T_i. See generally Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, 2d edition, Cold Spring Harbor Press (1989)) for suitable high stringency conditions.

Stringency conditions are a function of the temperature used in the hybridization experiment and washes, the molarity of the monovalent cations in the hybridization solution and in the wash solution(s) and the percentage of formamide in the hybridization solution. In general, sensitivity by hybridization with a probe is affected by the amount and specific activity of the probe, the amount of the target nucleic acid, the detectability of the label, the rate of hybridization, and the duration of the hybridization. The hybridization rate is maximized at a T_i (incubation temperature) of 20-25°C below T_m for DNA:DNA hybrids and 10-15°C below T_m for DNA:RNA hybrids. It is also maximized by an ionic strength of about 1.5M Na⁺. The rate is directly proportional to duplex length and inversely proportional to the degree of mismatching.

Specificity in hybridization, however, is a function of the difference in stability between the desired hybrid and "background" hybrids. Hybrid stability is a function of duplex length, base composition, ionic strength, mismatching, and destabilizing agents (if any).

The T_m of a perfect hybrid may be estimated for DNA:DNA hybrids using the equation of Meinkoth *et al* (1984), as

$$T_m = 81.5^{\circ}\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$$

and for DNA:RNA hybrids, as

$$T_m = 79.8^{\circ}\text{C} + 18.5 (\log M) + 0.58 (\%GC) - 11.8 (\%GC)^2 - 0.56(\% \text{ form}) - 820/L$$

where M, molarity of monovalent cations, 0.01-0.4 M NaCl,

%GC, percentage of G and C nucleotides in DNA, 30%-75%,
% form, percentage formamide in hybridization solution, and
L, length hybrid in base pairs.

T_m is reduced by 0.5-1.5°C (an average of 1°C can be used for ease of calculation) for
5 each 1% mismatching.

The T_m may also be determined experimentally. As increasing length of the hybrid (L) in the above equations increases the T_m and enhances stability, the full-length rat gene sequence can be used as the probe.

Filter hybridization is typically carried out at 68°C, and at high ionic strength (e.g., 5 - 6
10 X SSC), which is non-stringent, and followed by one or more washes of increasing stringency, the last one being of the ultimately desired high stringency. The equations for T_m can be used to estimate the appropriate T_i for the final wash, or the T_m of the perfect duplex can be determined experimentally and T_i then adjusted accordingly.

The present invention also relates to a vector comprising the nucleic acid molecule of the present
15 invention. The vector of the present invention may be, e.g., a plasmid, cosmid, virus, bacteriophage or another vector used e.g. conventionally in genetic engineering, and may comprise further genes such as marker genes which allow for the selection of said vector in a suitable host cell and under suitable conditions.

Furthermore, the vector of the present invention may, in addition to the nucleic acid sequences
20 of the invention, comprise expression control elements, allowing proper expression of the coding regions in suitable hosts. Such control elements are known to the artisan and may include a promoter, a splice cassette, translation initiation codon, translation and insertion site for introducing an insert into the vector.

Preferably, the nucleic acid molecule of the invention is operatively linked to said expression
25 control sequences allowing expression in eukaryotic or prokaryotic cells.

Control elements ensuring expression in eukaryotic or prokaryotic cells are well known to those skilled in the art. As mentioned herein above, they usually comprise regulatory sequences ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript.

30 Methods for construction of nucleic acid molecules according to the present invention, for construction of vectors comprising said nucleic acid molecules, for introduction of said

vectors into appropriately chosen host cells, for causing or achieving the expression are well-known in the art (see, e.g., Sambrook et al., 1989; Ausubel et al., 1994).

The invention also provides for conservative amino acid variants of the molecules of the invention. Variants according to the invention also may be made that conserve the overall molecular structure of the encoded proteins. Given the properties of the individual amino acids comprising the disclosed protein products, some rational substitutions will be recognized by the skilled worker. Amino acid substitutions, *i.e.* "conservative substitutions," may be made, for instance, on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

For example: (a) nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; (b) polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; (c) positively charged (basic) amino acids include arginine, lysine, and histidine; and (d) negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

Substitutions typically may be made within groups (a)-(d). In addition, glycine and proline may be substituted for one another based on their ability to disrupt α -helices. Similarly, certain amino acids, such as alanine, cysteine, leucine, methionine, glutamic acid, glutamine, histidine and lysine are more commonly found in α -helices, while valine, isoleucine, phenylalanine, tyrosine, tryptophan and threonine are more commonly found in β -pleated sheets. Glycine, serine, aspartic acid, asparagine, and proline are commonly found in turns. Some preferred substitutions may be made among the following groups: (i) S and T; (ii) P and G; and (iii) A, V, L and I. Given the known genetic code, and recombinant and synthetic DNA techniques, the skilled scientist readily can construct DNAs encoding the conservative amino acid variants.

As used herein, "sequence identity" between two polypeptide sequences indicates the percentage of amino acids that are identical between the sequences. "Sequence similarity" indicates the percentage of amino acids that either are identical or that represent conservative amino acid substitutions.

Conjugates

One embodiment of the present invention provides molecules of the present invention conjugated to cytotoxins. The cytotoxic moiety of the antibody may be a cytotoxic drug or an enzymatically active toxin of bacterial or plant origin, or an enzymatically active fragment of such a toxin including, but not limited to, diphtheria A chain, nonbinding active fragments of

diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, *Aleurites fordii* proteins, dianthin proteins, curcin, crotin, saponin, gelonin, mitogellin, restrictocin, phenomycin, and enomycin. In another embodiment, the molecules of the present invention are conjugate to small molecule anti-
5 cancer drugs. Conjugates of the antibody and such cytotoxic moieties are made using a variety of bifunctional protein coupling agents. Examples of such reagents include SPDP, IT, bifunctional derivatives of imidoesters such as dimethyl adipimidate HCl, active esters such as disuccinimidyl suberate, aldehydes such as glutaraldehyde, bis-azido compounds such as bis-(p-azidobenzoyl) hexanediamine, bis-diazonium derivatives, dissocyanates and bis-active
10 fluorine compounds. The lysing portion of a toxin may be joined to the Fab fragment of the antibodies.

The methods and compositions described herein may be performed, for example, by utilizing pre-packaged diagnostic test kits comprising in one or more containers (i) at least one immunoglobulin of the invention and (ii) a reagent suitable for detecting the presence of
15 said immunoglobulin when bound to its target. A kit may be conveniently used, *e.g.*, in clinical settings or in home settings, to diagnose patients exhibiting a disease (*e.g.*, skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression), and to screen and identify those individuals exhibiting a predisposition to such disorders. A composition of the invention also may be used in conjunction with a reagent
20 suitable for detecting the presence of said immunoglobulin when bound to its target, as well as instructions for use, to carry out one or more methods of the invention.

Having now generally described the invention, the same will be more readily understood through reference to the following examples, which are provided by way of illustration and are not intended to be limiting of the present invention.

25

EXAMPLES

An important approach to control FGFR3 activity is the generation of reagents that block receptor signaling. Without wishing to be bound by theory, molecules which bind the extracellular domain of the receptor may inhibit the receptor by competing with FGF or heparin binding or, alternatively, by preventing receptor dimerization. Additionally, binding to the extracellular domain may accelerate receptor internalization and turnover. Humanized antibodies are expected to have inhibitory/neutralizing action and are of particular interest since they are considered to be valuable for therapeutic applications, avoiding the human anti-mouse antibody response frequently observed with rodent antibodies. The experiments in which the neutralizing antibodies are screened, identified and obtained using fully synthetic human antibody libraries (for discovering highly specific binders against a wide variety of antigens) and FGFR3 extracellular domain are described below.

Example 1: Attempt to generate anti-FGFR3 antibodies

One hundred micrograms of soluble FGFR3 in complete Freund's Adjuvant were injected into Balb/c 3T3 naive mice (9 animals). Two repeated injections of 20 micrograms were performed at week intervals. 10 days after the second booster injection, blood was drawn from animals and serum was tested for the presence of polyclonal antibodies both by monitoring for binding to the receptor as well as for neutralizing activity at a dilution of 1:50. No significant neutralizing activity was observed in the tested serum (20% at most in some animals). A perfusion injection of 20 micrograms of soluble receptor was administered 1-2 days later but all the mice harboring some activity of neutralizing Ab died. The experiment was repeated twice with the same results.

Example 2: Generation of the FGFR3 Antigens

Two dimeric forms of the extracellular domain of the human FGFR3 were prepared for use as antigen. One was a histidine-tagged domain with a Serine 371 to Cysteine (S371C) substitution (thanatophoric dysplasia (TD) mutation) to facilitate dimerization and the second one an Fc fusion. The S371C variant was shown to bind heparin and FGF9 coated plates and to inhibit FGF9-dependent FDCP-FR3 proliferation. The Fc fusion was similarly effective in binding assays demonstrating its potential as an inhibitor of FGFR function and as a target for selecting FGFR3 inhibitory molecules. Both soluble receptors were employed to select neutralizing human recombinant antibodies.

The two variants of the FGFR3 extracellular domain were prepared as follows:

1. A construct containing the extracellular portion of FGFR3 with a thanatophoric dysplasia (TD) mutation to facilitate dimer formation conjugated to a His-tag (histidine tag) was generated. A bluescript plasmid comprising the human FGFR3 gene (pBS-hFGFR3) was used as template for PCR with the following primers:

- 5 5'-ACGTGCTAGC TGAGTCCTTG GGGACGGAGC AG (SEQ ID NO: 2).
 5'-ACGTCTCGAG TTAATGGTGA TGGTGATGGT GTGCATACAC **ACAG**CCCGCC TCGTC
 (SEQ ID NO: 3),

wherein the Ser 371 Cys (S371C) substitution is bold and underlined.

The nucleotide sequence encoding the extracellular domain of FGFR3 with the TD substitution is denoted herein SEQ ID NO: 7:

10 TGAGTCCTTG GGGACGGAGC AGCGCGTCGT GGGGCGAGCG GCAGAAGTCC CGGGCCCAGA 60
 GCGCGGCCAG CAGGAGCAGT TGGTCTTCGG CAGCGGGGAT GCTGTGGAGC TGAGCTGTCC 120
 CCCGCCCGGG GGTGGTCCCA TGGGGCCCAC TGTCTGGGTC AAGGATGGCA CAGGGCTGGT 180
 GCCCTCGGAG CGTGTCCTGG TGGGGCCCCA GCGGCTGCAG GTGCTGAATG CCTCCCACGA 240
 15 GGACTCCGGG GCCTACAGCT GCCGGCAGCG GCTCACGCAG CGCGTACTGT GCCACTTCAG 300
 TGTGCGGGTG ACAGACGCTC CATCCTCGGG AGATGACGAA GACGGGGAGG ACAGAGGCTGA 360
 GGACACAGGT GTGGACACAG GGGCCCCTTA CTGGACACGG CCCGAGCGGA TGGACAAGAA 420
 GCTGCTGGCC GTGCCGGCCG CCAACACCGT CCGCTTCCGC TGCCCAGCCG CTGGCAACCC 480
 CACTCCCTCC ATCTCCTGGC TGAAGAACGG CAGGGAGTTC CGCGGCGAGC ACCGCATTGG 540
 20 AGGCATCAAG CTGCGGCATC AGCAGTGGAG CCTGGTTCATG GAAAGCGTGG TGCCCTCGGA 600
 CCGCGGCAAC TACACCTGCG TCGTGGAGAA CAAGTTTGGC AGCATCCGGC AGACGTACAC 660
 GCTGGACGTG CTGGAGCGCT CCCCACCG GCCCATCCTG CAGGCGGGGC TGCCGGCCAA 720
 CCAGACGGCG GTGCTGGGCA GCGACGTGGA GTTCCACTGC AAGGTGTACA GTGACGCACA 780
 GCCCCACATC CAGTGGCTCA AGCACGTGGA GGTGAACGGC AGCAAGGTGG GCGCGACCG 840
 25 CACACCCTAC GTTACCGTGC TCAAGACGGC GGGCGCTAAC ACCACCGACA AGGAGCTAGA 900
 GGTCTCTCTC TTGCACAACG TCACCTTTGA GGACGCCGGG GAGTACACCT GCCTGGCGGG 960
 CAATTCTATT GGGTTTTCTC ATCACTCTGC GTGGCTGGTG GTGCTGCCAG CCGAGGAGGA 1020
 GCTGGTGGAG GCTGACGAGG CGGGCTGTGT GTATGCACAC CATCACCATC ACCATTAA 1078

The PCR fragment was digested with XhoI and ligated into pBlueScript digested with
 30 EcoRV and XhoI. The resulting plasmid, pBsFR3²³⁻³⁷⁴TDhis, was cleaved with NdeI and
 XhoI and the DNA fragment encoding the extracellular domain of FGFR3 was ligated into
 the same restriction sites in pCEP-Pu/Ac7 (Yamaguchi et al., 1999; Kohfeldt et al., 1997),
 generating the pCEP-hFR3²³⁻³⁷⁴TDhis plasmid construct.

To express this FGFR3 variant, 293E cells (EBNA virus transfected 293 cells) were
 35 transfected with the aforementioned plasmid, pCEP-hFR3²³⁻³⁷⁴TDhis, clones were identified

and grown. Cell supernatants analyzed by Western blot with anti-His antibody demonstrated high expression of the soluble receptor. Supernatants from large scale preparations were then subjected to batch affinity purification with Ni-NTA beads and the tagged soluble receptor was eluted by a step gradient ranging from 20 mM to 500 mM imidazol. A sample from each eluate was loaded onto a 7.5% SDS-PAGE and stained with GelCode (Pierce). In parallel Western blot analysis was performed and analyzed with anti-His antibodies. SDS-PAGE (Fig. 1) and immunoblot (not shown) analyses demonstrated peak amounts of purified extracellular FGFR3 in the 2nd (2) 50 mM imidazol fraction. About 0.5 mg of pure protein was obtained following this single step purification. In Figure 1, T=total protein, D= dialysed protein, U= unbound fraction.

To assess whether hFR3²³⁻³⁷⁴TDhis (also identified as hFR3-TDhis) retained the ability to associate with heparin and heparin-FGF complex, heparin coated wells were incubated with purified (2, 4 or 10 µg, labeled as FR3 2, FR3 4 or FR3 10, respectively) or unpurified (FR3 sup) hFR3²³⁻³⁷⁴TDhis with or without FGF9 (200ng/well). The binding of hFR3²³⁻³⁷⁴TDhis to each well was determined with anti-His antibody. Mock supernatant (M sup), PBS and unpurified mouse FR3AP (FGFR3-alkaline phosphatase, labeled as FRAP sup) were included as controls. This demonstrated that, like what was reported for the wild-type receptor, hFR3²³⁻³⁷⁴TDhis binds to heparin and that this interaction is augmented by the presence of FGF9 (Fig. 2). Finally, it was demonstrated that hFR3²³⁻³⁷⁴TDhis inhibits FDCP-FR3 FGF-dependent proliferation in a dose dependent manner. hFR3²³⁻³⁷⁴TDhis had no inhibitory effect when FDCP-FR3 cells were grown in the presence of IL-3. Taken together, hFR3²³⁻³⁷⁴TDhis proved to be a good candidate as a target antigen for screening for FGFR3 neutralizing antibodies.

2. The extracellular domain of FGFR3 and FGFR1 were prepared as Fc fusions (FR3exFc and FR1exFc). The amino acid sequence of FGFR3 (NCBI access no: NP_000133) is denoted herein SEQ ID NO:1.

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1  MGAPACALAL  CVAVAIVAGA  SSES LGTEQR  VVGRAAEVPG  PEPGQQEQLV  FGSGDAVELS
61  CPPPGGGPMG  PTVWVKDGTG  LVPSEVLVG  PQLQVLNAS  HEDSGAYSCR  QRLTQRVLCH
121 FSVRVTDAPS  SGDDDEGEDE  AEDTGVDTGA  PYWTRPERMD  KLLAVPAAN  TVRFRCPAAG
181 NPTPSISWLK  NGREFRGEHR  IGGIKLRHQQ  WSLVMESVVP  SDRGNYTCVV  ENKFGSIRQT
241 YTLDLVLERSP  HRPILQAGLP  ANQTAVLGSD  VEFHCKVYSD  AQPHIQWLKH  VEVNGSKVGP
301 DGTPYVTVLK  TAGANTTDKE  LEVL SLHNVT  FEDAGEYTCL  AGNSIGFSHH  SAWLVVLP AE
361 EELVEADEAG  SVYAGILSYG  VGFFLFILVV  AAVTLCRLRS  PPKKGLGSPT  VHKISRFP LK
421 RQVSLESNAS  MSSNTPLVRI  ARLSSGEGPT  LANVSELELP  ADPKWELSRA  RLTLGKPLGE
481 GCFGQVMAE  AIGIDKDRAA  KPVTVAVKML  KDDATDKDLS  DLVSEMEMMK  MIQKHKN IIN

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541 LLGACTQGGP LYVLVEYAAK GNLREFLRAR RPPGLDYSFD TCKPPEEQLT FKDLVSCAYQ
 601 VARGMEYLAS QKCIHRDLAA RNVLVTEDNV MKIADFGLAR DVHNLDDYYKK TTNGRLPVKW
 661 MAPEALFDRV YTHQSDVWSF GVLLWEIFTL GGSPYPGIPV EELFKLLKEG HRMDKPANCT
 721 HDLYMIMREC WHAAPSQRPT FKQLVEDLDR VLTVTSTDEY LDLSAPFEQY SPGGQDTPSS
 5 781 SSSGDDSVFA HDLLPPAPPS SSGSRT

To construct the FR3exFc fusion, a nucleotide sequence (SEQ ID NO:4) encoding the extracellular domain of FGFR3 was PCR amplified to contain terminal KpnI and BamHI restriction sites for insertion into the KpnI and BamHI sites of pCXFc (SEQ ID NO:5). This insertion positions the extracellular domain of FGFR3 to be expressed as a fusion with the Fc amino acid sequence (SEQ ID NO:6).

SEQ ID NO:4:

	GCGCGCTGCC	TGAGGACGCC	GCGGCCCCCG	CCCCGCCCAT	GGGCGCCCCT	GCCTGCGCCC	60
	TCGCGCTCTG	CGTGGCCGTG	GCCATCGTGG	CCGGCGCCTC	CTCGGAGTCC	TTGGGGACGG	120
	AGCAGCGCGT	CGTGGGGCGA	GCGGCAGAAG	TCCCGGGCCC	AGAGCCCGGC	CAGCAGGAGC	180
15	AGTTGGTCTT	CGGCAGCGGG	GATGCTGTGG	AGCTGAGCTG	TCCCCCGCCC	GGGGGTGGTC	240
	CCATGGGGCC	CACTGTCTGG	GTCAAGGATG	GCACAGGGCT	GGTGCCCTCG	GAGCGTGTCC	300
	TGGTGGGGCC	CCAGCGGCTG	CAGGTGCTGA	ATGCCTCCCA	CGAGGACTCC	GGGGCCTACA	360
	GCTGCCGGCA	GCGGCTCACG	CAGCGCGTAC	TGTGCCACTT	CAGTGTGCGG	GTGACAGACG	420
	CTCCATCCTC	GGGAGATGAC	GAAGACGGGG	AGGACGAGGC	TGAGGACACA	GGTGTGGACA	480
20	CAGGGGCCCC	TTACTGGACA	CGGCCCCGAG	GGATGGACAA	GAAGCTGCTG	GCCGTGCCGG	540
	CCGCCAACAC	CGTCCGCTTC	CGCTGCCCAG	CCGCTGGCAA	CCCCACTCCC	TCCATCTCCT	600
	GGCTGAAGAA	CGGCAGGGAG	TTCCGCGGCG	AGCACC GCAT	TGGAGGCATC	AAGCTGCGGC	660
	ATCAGCAGTG	GAGCCTGGTC	ATGGAAAGCG	TGGTGCCCTC	GGACCGCGGC	AACTACACCT	720
	GCGTCGTGGA	GAACAAGTTT	GGCAGCATCC	GGCAGACGTA	CACGCTGGAC	GTGCTGGAGC	780
25	GCTCCCCGCA	CCGGCCCATC	CTGCAGGCGG	GGCTGCCGGC	CAACCAGACG	GCGGTGCTGG	840
	GCAGCGACGT	GGAGTTCCAC	TGCAAGGTGT	ACAGTGACGC	ACAGCCCCAC	ATCCAGTGGC	900
	TCAAGCACGT	GGAGGTGAAC	GGCAGCAAGG	TGGGCCCGGA	CGGCACACCC	TACGTTACCG	960
	TGCTCAAGAC	GGCGGGCGCT	AACACCACCG	ACAAGGAGCT	AGAGGTTCCT	TCCTTGACACA	1020
	ACGTCACCTT	TGAGGACGCC	GGGGAGTACA	CCTGCCCTGGC	GGGCAATTCT	ATTGGGTTTT	1080
30	CTCATCACTC	TGCGTGGCTG	GTGGTGCTGC	CAGCCGAGGA	GGAGCTGGTG	GAGGCTGACG	1140
	AGGCGGG						1147

SEQ ID NO:5:

35	GACGGATCGG	GAGATCTCCC	GATCCCCTAT	GGTCGACTCT	CAGTACAATC	TGCTCTGATG	60
	CCGCATAGTT	AAGCCAGTAT	CTGCTCCCTG	CTTGTGTGTT	GGAGGTCGCT	GAGTAGTGCG	120
	CGAGCAAAAT	TTAAGCTACA	ACAAGGCAAG	GCTTGACCGA	CAATTGCATG	AAGAATCTGC	180
	TTAGGGTTAG	GCGTTTTGCG	CTGCTTCGCG	ATGTACGGGC	CAGATATACG	CGTTGACATT	240

	GATTATTGAC	TAGTTATTAA	TAGTAATCAA	TTACGGGGTC	ATTAGTTCAT	AGCCCATATA	300
	TGGAGTTCGG	CGTTACATAA	CTTACGGTAA	ATGGCCCCGC	TGGCTGACCG	CCCAACGACC	360
	CCCGCCCAT	GACGTCAATA	ATGACGTATG	TTCCCATAGT	AACGCCAATA	GGGACTTTCC	420
	ATTGACGTCA	ATGGGTGGAC	TATTTACGGT	AAACTGCCCA	CTTGGCAGTA	CATCAAGTGT	480
5	ATCATATGCC	AAGTACGCCC	CCTATTGACG	TCAATGACGG	TAAATGGCCC	GCCTGGCATT	540
	ATGCCCAGTA	CATGACCTTA	TGGGACTTTC	CTACTTGGCA	GTACATCTAC	GTATTAGTCA	600
	TCGCTATTAC	CATGGTGATG	CGGTTTTGGC	AGTACATCAA	TGGGCGTGGA	TAGCGGTTTG	660
	ACTCACGGGG	ATTTCCAAGT	CTCCACCCCA	TTGACGTCAA	TGGGAGTTTG	TTTTGGCACC	720
	AAAATCAACG	GGACTTTCCA	AAATGTCGTA	ACAACCTCCGC	CCCATTGACG	CAAATGGGCG	780
10	GTAGGCGTGT	ACGGTGGGAG	GTCTATATAA	GCAGAGCTCT	CTGGCTAACT	AGAGAACCCA	840
	CTGCTTACTG	GCTTATCGAA	ATTAATACGA	CTCACTATAG	GGAGACCCAA	GCTGGCTAGC	900
	GTTTAAACTT	AAGCTTGGTA	CCGAGCTCGG	ATCCCCGTCG	TGCATCTATC	GAAGGTCGTG	960
	GA GAT CCC	GAG GAG CCC	AAA TCT	TGT GAC AAA	ACT CAC ACA	TGC CCA	1007
15	ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO						
	<u>SEQ ID NO. 97</u>						
	5 10 15						
	CCG TGC CCA	GCA CCT	GAA CTC	CTG GGG	GGA CCG	TCA GTC	TTC CTC TTC 1055
	PRO CYS PRO	ALA PRO	GLU LEU	LEU GLY	GLY PRO	SER VAL	PHE LEU PHE
	20 25 30						
20	CCC CCA AAA	CCC AAG	GAC ACC	CTC ATG	ATC TCC	CGG ACC	CCT GAG GTC 1103
	PRO PRO LYS	PRO LYS	ASP THR	LEU MET	ILE SER	ARG THR	PRO GLU VAL
	35 40 45						
	ACA TGC GTG	GTG GTG	GAC GTG	AGC CAC	GAA GAC	CCT GAG	GTC AAG TTC 1151
	THR CYS VAL	VAL VAL	ASP VAL	SER HIS	GLU ASP	PRO GLU	VAL LYS PHE
25	50 55 60						
	AAC TGG TAC	GTG GAC	GGC GTG	GAG GTG	CAT AAT	GCC AAG	ACA AAG CCG 1199
	ASN TRP TYR	VAL ASP	GLY VAL	GLU VAL	HIS ASN	ALA LYS	THR LYS PRO
	65 70 75						
	CGG GAG GAG	CAG TAC	AAC AGC	ACG TAC	CGG GTG	GTC AGC	GTC CTC ACC 1247
30	ARG GLU GLU	GLN TYR	ASN SER	THR TYR	ARG VAL	VAL SER	VAL LEU THR
	80 85 90 95						
	GTC CTG CAC	CAG GAC	TGG CTG	AAT GGC	AAG GAG	TAC AAG	TGC AAG GTC 1295
	VAL LEU HIS	GLN ASP	TRP LEU	ASN GLY	LYS GLU	TYR LYS	CYS LYS VAL
	100 105 110						
35	TCC AAC AAA	GCC CTC	CCA GCC	CCC ATC	GAG AAA	ACC ATC	TCC AAA GCC 1343
	SER ASN LYS	ALA LEU	PRO ALA	PRO ILE	GLU LYS	THR ILE	SER LYS ALA
	115 120 125						
	AAA GGG CAG	CCC CGA	GAA CCA	CAG GTG	TAC ACC	CTG CCC	CCA TCC CGG 1391
	LYS GLY GLN	PRO ARG	GLU PRO	GLN VAL	TYR THR	LEU PRO	PRO SER ARG
40	130 135 140						

	GAT GAG CTG ACC AAG AAC CAG GTC AGC CTG ACC TGC CTG GTC AAA GGC	1439
	ASP GLU LEU THR LYS ASN GLN VAL SER LEU THR CYS LEU VAL LYS GLY	
	145 150 155	
	TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG GAG AGC AAT GGG CAG CCG	1487
5	PHE TYR PRO SER ASP ILE ALA VAL GLU TRP GLU SER ASN GLY GLN PRO	
	160 165 170 175	
	GAG AAC AAC TAC AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GGC TCC	1535
	GLU ASN ASN TYR LYS THR THR PRO PRO VAL LEU ASP SER ASP GLY SER	
	180 185 190	
10	TTC TTC CTC TAC AGC AAG CTC ACC GTG GAC AAG AGC AGG TGG CAG CAG	1583
	PHE PHE LEU TYR SER LYS LEU THR VAL ASP LYS SER ARG TRP GLN GLN	
	195 200 205	
	GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT GAG GCT CTG CAC AAC CAC	1631
	GLY ASN VAL PHE SER CYS SER VAL MET HIS GLU ALA LEU HIS ASN HIS	
15	210 215 220	
	TAC ACG CAG AAG AGC CTC TCC CTG TCT CCG GGT AAA TGATCTAGAG	1677
	TYR THR GLN LYS SER LEU SER LEU SER PRO GLY LYS	
	225 230 235	
	GGCCCGTTTA AACCCGCTGA TCAGCCTCGA CTGTGCCTTC TAGTTGCCAG CCATCTGTTG	1737
20	TTTGCCCTC CCCCCTGCCT TCCTTGACCC TGGAAGGTGC CACTCCCACT GTCCTTTCCT	1797
	AATAAAATGA GGAAATTGCA TCGCATTGTC TGAGTAGGTG TCATTCTATT CTGGGGGGTG	1857
	GGGTGGGGCA GGACAGCAAG GGGGAGGATT GGGAAGACAA TAGCAGGCAT GCTGGGGATG	1917
	CGGTGGGCTC TATGGCTTCT GAGGCGGAAA GAACCAGCTG GGGCTCTAGG GGGTATCCCC	1977
	ACGCGCCCTG TAGCGGCGCA TTAAGCGCGG CGGGTGTGGT GGTACGCGC AGCGTGACCG	2037
25	CTACACTTGC CAGCGCCCTA GCGCCCGCTC CTTTCGCTTT CTTCCCTTCC TTTCTCGCCA	2097
	CGTTCGCCCG CTTTCCCCGT CAAGCTCTAA ATCGGGGCAT CCCTTTAGGG TTCCGATTTA	2157
	GTGCTTTACG GCACCTCGAC CCCAAAAAAC TTGATTAGGG TGATGGTTCA CGTAGTGGGC	2217
	CATCGCCCTG ATAGACGGTT TTTGCGCCTT TGACGTTGGA GTCCACGTTT TTTAATAGTG	2277
	GACTCTTGTT CCAAACCTGGA ACAACACTCA ACCCTATCTC GGTCTATTCT TTTGATTTAT	2337
30	AAGGGATTTT GGGGATTTTC GCCTATTGGT TAAAAAATGA GCTGATTTAA CAAAAATTTA	2397
	ACGCGAATTA ATTCTGTGGA ATGTGTGTCA GTTAGGGTGT GGAAAGTCCC CAGGCTCCCC	2457
	AGGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCAGG TGTGGAAAGT	2517
	CCCCAGGCTC CCCAGCAGGC AGAAGTATGC AAAGCATGCA TCTCAATTAG TCAGCAACCA	2577
	TAGTCCCGCC CCTAACTCCG CCCATCCCGC CCCTAACTCC GCCCAGTTCC GCCCATTTCTC	2637
35	CGCCCCATGG CTGACTAATT TTTTTTATTT ATGCAGAGGC CGAGGCCGCC TCTGCCTCTG	2697
	AGCTATTCCA GAAGTAGTGA GGAGGCTTTT TTGGAGGCCT AGGCTTTTGC AAAAAGCTCC	2757
	CGGGAGCTTG TATATCCATT TTCGGATCTG ATCAGCACGT GTTGACAATT AATCATCGGC	2817
	ATAGTATATC GGCATAGTAT AATACGACAA GGTGAGGAAC TAAACCATGG CCAAGTTGAC	2877
	CAGTGCCGTT CCGGTGCTCA CCGCGCGCGA CGTCGCCGGA GCGGTCGAGT TCTGGACCGA	2937
40	CCGGCTCGGG TTCTCCCGGG ACTTCGTGGA GGACGACTTC GCCGGTGTGG TCCGGGACGA	2997

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	CTTCCGGGAC	GCCTCCGGGC	CGGCCATGAC	CGAGATCGGC	GAGCAGCCGT	GGGGCCGGGA	3177
	GTTCGCCCTG	CGCGACCCGG	CCGGCAACTG	CGTGCAC TTC	GTGGCCGAGG	AGCAGGACTG	3237
5	ACACGTGCTA	CGAGATTTCG	ATTCCACCGC	CGCCTTCTAT	GAAAGGTTGG	GCTTCGGAAT	3297
	CGTTTTCCGG	GACGCCGGCT	GGATGATCCT	CCAGCGCGGG	GATCTCATGC	TGGAGTTCTT	3357
	CGCCCACCCC	AACTTGTTTA	TTGCAGCTTA	TAATGGTTAC	AAATAAAGCA	ATAGCATCAC	3417
	AAATTTTACA	AATAAAGCAT	TTTTTTTCACT	GCATTCTAGT	TGTGGTTTTGT	CCAAACTCAT	3477
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10	GTCATAGCTG	TTTCCTGTGT	GAAATTGTTA	TCCGCTCACA	ATTCCACACA	ACATACGAGC	3597
	CGGAAGCATA	AAGTGTAAG	CCTGGGGTGC	CTAATGAGTG	AGCTAACTCA	CATTAATTGC	3657
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	GCCAGTTAAT	AGTTTGCGCA	ACGTTGTTGC	CATTGCTACA	GGCATCGTGG	TGTCACGCTC	5037
35	GTCGTTTGGT	ATGGCTTCAT	TCAGCTCCGG	TTCCCAACGA	TCAAGGCGAG	TTACATGATC	5097
	CCCCATGTTG	TGCAAAAAAG	CGGTTAGCTC	CTTCGGTCCT	CCGATCGTTG	TCAGAAGTAA	5157
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	GCCATCCGTA	AGATGCTTTT	CTGTGACTGG	TGAGTACTCA	ACCAAGTCAT	TCTGAGAATA	5277
	GTGTATGCGG	CGACCGAGTT	GCTCTTGCCC	GGCGTCAATA	CGGGATAATA	CCGCGCCACA	5337
40	TAGCAGAACT	TTAAAAGTGC	TCATCATTGG	AAAACGTTCT	TCGGGGCGAA	AACTCTCAAG	5397
	GATCTTACCG	CTGTTGAGAT	CCAGTTCGAT	GTAACCCACT	CGTGCACCCA	ACTGATCTTC	5457

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AGCATCTTTT ACTTTCACCA GCGTTTCTGG GTGAGCAAAA ACAGGAAGGC AAAATGCCGC 5517
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TTATTGAAGC ATTTATCAGG GTTATTGTCT CATGAGCGGA TACATATTTG AATGTATTTA 5637
GAAAAATAAA CAAATAGGGG TTCCGCGCAC ATTTCCCCGA AAAGTGCCAC CTGACGTC 5695

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SEQ ID NO:6

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ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO PRO
1          5          10          15
10 CYS PRO ALA PRO GLU LEU LEU GLY GLY PRO SER VAL PHE LEU PHE PRO
          20          25          30
PRO LYS PRO LYS ASP THR LEU MET ILE SER ARG THR PRO GLU VAL THR
          35          40          45
CYS VAL VAL VAL ASP VAL SER HIS GLU ASP PRO GLU VAL LYS PHE ASN
15          50          55          60
TRP TYR VAL ASP GLY VAL GLU VAL HIS ASN ALA LYS THR LYS PRO ARG
65          70          75          80
GLU GLU GLN TYR ASN SER THR TYR ARG VAL VAL SER VAL LEU THR VAL
          85          90          95
20 LEU HIS GLN ASP TRP LEU ASN GLY LYS GLU TYR LYS CYS LYS VAL SER
          100          105          110
ASN LYS ALA LEU PRO ALA PRO ILE GLU LYS THR ILE SER LYS ALA LYS
          115          120          125
GLY GLN PRO ARG GLU PRO GLN VAL TYR THR LEU PRO PRO SER ARG ASP
25          130          135          140
GLU LEU THR LYS ASN GLN VAL SER LEU THR CYS LEU VAL LYS GLY PHE
145          150          155          160
TYR PRO SER ASP ILE ALA VAL GLU TRP GLU SER ASN GLY GLN PRO GLU
          165          170          175
30 ASN ASN TYR LYS THR THR PRO PRO VAL LEU ASP SER ASP GLY SER PHE
          180          185          190
PHE LEU TYR SER LYS LEU THR VAL ASP LYS SER ARG TRP GLN GLN GLY
          195          200          205
ASN VAL PHE SER CYS SER VAL MET HIS GLU ALA LEU HIS ASN HIS TYR
35          210          215          220
THR GLN LYS SER LEU SER LEU SER PRO GLY LYS
225          230          235

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Both FR3exFc and FR1exFc soluble receptors were demonstrated to be expressed to a
40 high level in transiently transfected 293T cells (T-cell antigen infected human embryonic

kidney 293 cells). The observation that both soluble receptors remain bound to heparin-coated wells even following extensive washes led the laboratory of the present inventors to try to purify the proteins with the commercial heparin-Sepharose™ resin (Pharmacia). One hundred ml volume supernatants, harvested 48 hours post transfection with either FR3exFc or FR1exFc coding plasmids, were incubated overnight at 4°C with 1 ml heparin-Sepharose™ resin. The resin was washed and then subjected to PBS supplemented with increasing concentration of NaCl. Aliquots of each fraction were analyzed by 7.5% SDS-PAGE stained with GelCode (Pierce) demonstrating a purification profile of more than 90% homogeneity and a peak elution at 400 mM NaCl for FR3exFc (Fig. 3; T=total protein, U=unbound fraction, W=wash). In contrast, FR1exFc was hardly retained on the resin. This result was confirmed by Western analysis of the same fractions with anti-FGFR1ex antibodies demonstrating that most of FR1exFc is in the unbound fraction (not shown).

Functional analysis of FR3exFc and FR1exFc showed that both compete efficiently for FGF9 binding and stimulating FGFR3, thus, demonstrating their potential as inhibitors of FGFRs function and as a target (FR3exFc) for selecting FGFR3 inhibitory molecules.

Neutralizing effect of soluble receptors

The ability of hFR3-TDhis and FR3exFc to inhibit FGF-dependent FDCP-R3 cell proliferation was compared. Both soluble receptors inhibited FDCP-R3 cell proliferation, however, FR3exFc was about 60 times more potent than hFR3TDhis (Fig. 4; legend: ◇- FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + FGF9, □-FR3exFc on FDCP-FR3 cells + FGF9, Δ- FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + IL, X- FR3exFc on FDCP-FR3 cells + IL3). Neither had an effect on FDCP cells stimulated with IL3. The fact that FR3exFc is entirely in dimeric form whereas only a small proportion (1/10) of hFR3²³⁻³⁷⁴TDhis is in a dimeric form might explain, at least in part, this difference.

Example 3: Screening for Antibodies

Panning and first screening of Ab Binding Characterization

The screening strategies to identify Fabs from the Human Combinatorial Antibody Library (HuCAL®), developed at MorphoSys, Munich, Germany and disclosed in WO 97/08320, U.S. patent 6,300,064, and Knappik et al., (2000), the entire contents of which are incorporated herein by reference, using soluble dimeric forms of the extracellular domain of the FGFR3 receptor are shown in Table 2.

TABLE 2

Panning Strategies

	Panning Round 1	Panning Round 2	Panning Round 3
Screen 1	FR3-TDhis	HEK293	FR3-TDhis
Screen 2	FR3exFc captured with mouse anti-human IgG	RCJ-FR3ach	FR3exFc captured with mouse anti-human IgG
Screen 3	FR3-TDhis (Round 1 of panning 1)	RCJ-FR3ach & RCJ-FR3wt	FR3exFc Captured with mouse anti-human IgG

The screening was carried out, for example in Screen 1, by coating the wells of a 96 well plate with hFR3²³⁻³⁷⁴TDhis (FR3-TDhis), panning with the bacteriophage library and selecting the positive clones. The positive clones were then tested on HEK293 (293, human embryonic kidney) cells, expressing endogenous FGFR3. The positive clones were selected and rescreened on FR3-TDhis. Two additional similar screenings were carried out as shown in Table 2. In screen 2 the first and third pannings were carried out with the FR3exFc antigen and the second panning carried out with RCJ cells expressing a mutant (achondroplasia) form of FGFR3.

An overview of the number of initial hits and of the candidate clones is shown in Table 3.

Table 3
Overview of Screenings 1, 2 and 3 on FGFR3

	screened clones	primary hits	sequenced clones	consolidated candidate clones (ELISA & FACS)
Screen 1	1076	208	69	15 MSPRO 1-15
Screen 2	864	300	32	22 MSPRO 20-33 and 52-59
Screen 3	768	487	52	11 MSPRO 40-50

5 Example 4: Analysis of Fabs identified by first screening.

Specificity of Antibody recognition

The first screening done at Morphosys yielded 15 different Fabs that specifically recognize FGFR3 *in vitro* and on the cell surface. Fourteen of these were produced and sent to ProChon for further analysis. LY6.3, an anti-lysosyme antibody, was isolated from the same library and serves as a control. ELISA analysis, according to the following protocol

10 was carried out to determine the specificity of the isolated Fabs for FGFR3 or FGFR1.

Fab-FR3/Fc Binding Assay

MaxiSorp ELISA plates were coated with 100 µl anti-human Fc (10 µg/ml) in bicarbonate overnight at 4°C. Wells were washed five consecutive times with a PBS solution

15 containing 0.1% Tween 20 (PBST). The well surface was blocked with 250 µl PBST+3%BSA (blocking solution) for 1 hour at 37°C. This was followed by capturing 1 µg of FGFR/Fc for 1 hour at room temperature. To assess the antibody binding to the captured FGFR/Fc, 1 µg each of the tested Fabs was incubated in 100 µl blocking solution per well 1 hour at room temperature. Wells were washed 5 times with PBST. Reaction was initiated with

20 the addition of 100 µl of 0.8µg/ml goat anti-human Fab-HRP diluted in blocking solution, subsequently washed and detected with TMB substrate (Pierce). The absorbance was

measured at 450 nm. A comparison of ELISA analyses done in the laboratory of the present inventors is presented in the following Table 4.

Table 4

	<u>ProChon</u>		<u>Morphosys</u>	
	FR1/Fc	FR3/Fc	FR1/Fc	FR3/Fc
MS-PRO1	++	++	+/-	+
MS-PRO2	-	++	-	++
MS-PRO3	+	++	-	++
MS-PRO4	-	+	-	++
MS-PRO5	-	++	+/-	+
MS-PRO6	-	++	-	+
MS-PRO7	-	++	-	+
MS-PRO8	+	++	-	+
MS-PRO9	-	+/-	+/-	+
MS-PRO10	+	++	-	++
MS-PRO11	-	+/-	+	++
MS-PRO12	-	+/-	-	++
MS-PRO13	-	+/-	+/-	+
MS-PRO14	-	-	-	+
LY6.3 (control)	-	-		

5

In most cases, the data generated in the labs of the present inventors are in agreement. However, some Fabs behave differently. For example, MS-PRO3 and 10 were found to be completely FGFR3 specific at Morphosys' lab. In Prochon's lab, the data show that both MS-PRO3 and 10 considerably cross-react with FGFR1. The FACS analysis, done at Morphosys, supports the Prochon results for MS-PRO3, but not for MS-PRO10. Taking into account the potency and specificity of the Fabs, MS-PRO2 has the highest score according to these preliminary data.

10

Example 5: Affinity of Fab to FGFR3

The affinity measurements were performed by BIAcore according to the standard procedure recommended by the supplier (Pharmacia). The anti-Fc antibody was coupled via

15

the EDC/NHS chemistry to the chip and subsequently FGFR3 was captured. The Fabs of the invention were then bound to this surface.

Table 5 shows a comparison of affinities of Fabs candidates to FGFR3 as determined by BIAcore and by FACS-scatchard.

5

Table 5

**Comparison of Antibody Affinities to FGFR3
determined by BIAcore and FACS-Scatchard**

Fab clone	BIAcore [nM]	Indirect FACS-Scatchard [nM]
MS-Pro-2	37 ± 10	43
MS-Pro-11	4 ± 2	4
MS-Pro-12	14 ± 2	6.5
MS-Pro-21	9 ± 2	0.6
MS-Pro-24	10 ± 2	0.3
MS-Pro-26	4 ± 1	1.4
MS-Pro-28	9 ± 0.4	0.3
MS-Pro-29	6 ± 4	0.4

10

Table 6 shows the affinity as determined by BIAcore for the Fab candidates shown in Table 5 converted into the Fab mini-antibody format, Fab-dHLX-MH, where a dimer of the Fab monomer is produced after insertion into an expression vector as a fusion protein.

Table 6

Fab-miniantibody format for candidate clones

Clone	BIAcore K_D [nM]
MSPro2-dHLX-MH	4.3
MSPro11-dHLX-MH	0.7
MSPro12-dHLX-MH	1.2
MSPro22-dHLX-MH	2.2
MSPro24-dHLX-MH	2
MSPro26-dHLX-MH	2
MSPro28-dHLX-MH	1.6

Table 7 shows the results of a competition assay wherein each MSPRO Fab was bound to the FGFR3 at a concentration of 500nM or 1, 000 nM and coinjected in pairs with the other MSPRO Fabs. The (-) indicates binding to the same or nearby epitope while (+) indicates binding to different epitope. The results show that MSPRO2 and 12 bind to the same or nearby epitope while MSPRO11, 21, 24, 26, 28 and 29 bind to an epitope different from that of MSPRO2 or 12.

Example 6: Specific Neutralizing Activity of the Antibodies**A: FDCP Cell Proliferation Assay**

The FDCP cell line is a murine immortalized, interleukin 3 (IL3) dependent cell line of myelocytic bone marrow origin, which does not express endogenous FGF Receptors (FGFR). Upon transfection with FGFR cDNA, the FDCP cell line exhibits an FGF dose dependent proliferative response that can replace the dependence on IL3. FDCP cell lines, transfected with FGFR cDNAs can therefore be used to screen for specific inhibitors or activators of FGFR, as well as for analyzing FGFR signaling. The FDCP cell response to various ligands was quantitated by a cell proliferation assay with XTT reagent (Cell Proliferation Kit, Biological Industries Co.). The method is based on the capability of mitochondrial enzymes to reduce tetrazolium salts into soluble colored formazan compounds which can be quantitated and is indicative of cell viability. Specifically, FDCP cells expressing

FGFR3IIIb, FGFR3IIIc or FGFR1 were grown in “full medium” (Iscove’s Medium containing 2ml glutamine, 10% FCS, 100ug/ml penicillin, 100ug/ml streptomycin) supplemented with 5ug/ml heparin and 10ng/ml FGF9. Cells were split every 3 days and kept in culture no more than one month. One day prior to the experiment, the cells were split.

5 Before the experiment, the cells were washed 3 times (1000 rpm, 6 min) with full medium. Later, the cells were resuspended and counted with Trypan Blue. Twenty thousand (20,000) cells /well were added to wells in a 96-well plate in 50ul in full medium containing 5 ug/ml heparin. Conditioned medium was added in an additional volume of 50ul full medium containing FGF9 at varying concentrations to a final volume of 100ul. A primary stock

10 solution (usually 2x the higher concentration) of the antibody (or Fabs) was prepared in Iscove’s+++ containing 5µg/ml heparin and 2.5ng/ml FGF9 or IL-3 (final concentration 1.25 ng/ml). Dilutions were filtered in a 0.2 µm syringe nitrocellulose filter blocked first with 1mg/ml BSA and washed then with Iscove’s+++ . Aliquots of requested serial dilutions were prepared. Dilutions were kept on ice until use. 50 µl of the corresponding 2x final

15 concentration was added to each well and the plate was incubated at 37⁰C for either 40 hours or either 64 hours. After incubation, the reaction was developed as follows: 100 µl of activator solution was added to 5 ml XTT reagent and mixed gently. 50 µl of mixture was added to each well. Optical density (OD) at 490 nm at this point gave the zero time reading.

Cells were then incubated at 37⁰C for 4 hours (in the case of a 40 hour incubation) or 2

20 hours (in the case of a 64 hour incubation) and proliferation was measured by O.D. at 490 nm (A490).

It is noted that the assay is successful when the O.D. of untreated control growing with saturated amounts of FGF (10 and 20 ng/ml) is at least 1.3 O.D. units. Furthermore, it is noted that the background of wells with no cells should be 0.2-0.35 O.D. units and that the

25 O.D. absorbance of 1.25 ng/ml FGF9 should not be less than 40% of the O.D. absorbance achieved with saturated FGF 9 concentration (10 and 20 ng/ml). Specific inhibition of FGF and FGF receptor mediated proliferation should always be accompanied with lack of any inhibition of the same antibody concentration on IL-3 dependent cell proliferation.

The following FDCP cell lines were used:

- 30 *FDCP-C10: FDCP cells transfected with the human wild-type FGF receptor 3IIIc.
- *FDCP-R3: FDCP cells transfected with the human wild-type FGF receptor 3IIIb.
- *FDCP-R1: FDCP cells transfected with the human wild-type FGFR1.

*FDCP-F3Ach: FDCP cells infected with human FGFR3 mutated at amino acid Glycine 380 to Arginine (G380R), analogous to the most common human achondroplasia mutation.

B: Neutralizing activity

The neutralizing activity of the antibodies was measured by the aforementioned cell proliferation analysis in FDCP-FR3 and FDCP-FR1 cell lines. Increasing amounts of the indicated Fabs were added to FDCP-FR3 (diamond, *, and X) or FDCP-FR1 (triangle, square and circle) grown in the presence of FGF9 (Fig. 5 for MSPRO 2, 3 and 4). Two days later, an XTT proliferation assay was performed. While none of the Fabs inhibited FDCP-FR1 cell proliferation, MSPRO2 and 3 inhibited FDCP-FR3 proliferation with a similar IC50 of about 1.0µg/ml (Fig. 5). In contrast, MS-PRO4 had no inhibitory effect on FDCP-FR3 proliferation. These data are in agreement with those generated at Morphosys. The rest of the Fabs were similarly analyzed on FDCP-FR3 expressing cells. Increasing amounts of the indicated Fabs were added to FDCP-FR3 grown in the presence of FGF9 (Fig. 6). The results of the proliferation assay done at Morphosys and at Prochon are compared in Table 7. (NA- data not available)

Table 7

	<u>Prochon</u>		<u>Morphosys</u>	
	FDCP-FR1	FDCP-FR3	FDCP-FR1	FDCP-FR3
MSPRO1	-	++	NA	NA
MSPRO2	-	++	NA	++
MSPRO3	-	++	NA	++
MSPRO4	-	-	NA	-
MSPRO5	-	+	NA	+
MSPRO6	-	-	NA	+/-
MSPRO7	-	++	NA	+
MSPRO8	-	+/-	NA	+/-
MSPRO9	-	+	NA	+
MSPRO10	-	+	NA	NA
MSPRO11	-	+++	NA	++
MSPRO12	-	+++	NA	+++
MSPRO13	-	-	NA	NA
MSPRO14	-	-	NA	NA
LY6.3	-	-	NA	NA

As shown in Table 7, there is an excellent agreement between the Prochon and
5 Morphosys data. About half of the Fabs show considerable neutralizing activity, MSPRO12
being the most potent. Most of the inhibitory Fabs performed well in the binding assay (Table
4), with MSPRO11 and MSPRO12 being the exception to the rule, however, clearly remain
good candidates to pursue. None of the Fabs (including those that crossreact with FGFR1)
inhibited FGF-dependent FDCP-FR1 proliferation. In addition, FDCP-FR3 grown in the
10 presence of IL3 were not affected by any of the Fabs.

An additional 20 new Fabs were selected from the second panning done at Morphosys.
Three of these new Fabs (MSPRO52, MSPRO54 and MSPRO55) were subjected to the
FDCP cell proliferation test and all were found to neutralize the receptor (Fig. 7A).
Interestingly (and in accord with MorphoSys affinity data), one Fab (MSPRO54) showed
15 strong neutralizing activity against FGFR1 (Fig. 7B).

Example 7: Receptor Expression and Activation in RCJ Cells**RCJ cell assay**

RCJ cells (fetal rat calvaria-derived mesenchymal cells, RCJ 3.1C5.18; Grigoriadis, 1988) were generated to express various FGF Receptors an inducible manner, in the absence of tetracycline. The RCJ-M14 line (RCJ-FR3ach) expresses FGFR3-ach380 mutant upon induction by the removal of tetracycline. The cells were then incubated in low serum after which FGF was added to stimulate receptor function and signaling. The cells were lysed and the receptor level, receptor activation and signaling are assessed by Western with anti-FGFR3 (Santa Cruz), anti-phospho-tyrosine (Promega), and anti-active ERK (or JNK) (Promega) respectively.

RCJ-M14 cells were grown in α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 μ g/ml neomycin, 2 μ g/ml tetracycline, 50 μ g/ml hygromycin B to subconfluence. The medium was aspirated off and the cells washed with trypsin, 1 ml/10 cm dish, then trypsinized with 0.5 ml/10 cm dish. The cells were resuspended in 10 ml α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 μ g/ml neomycin, and 2 μ g/ml tetracycline.

6x10⁵ cells/well were seeded in a 6-well dish. Alternatively, twice that number may be seeded. The cells were washed thrice 24 hours later (or 8 hours later if twice the amount of cells are seeded) with 1 ml α -MEM, and then incubated with α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, and 1x glutamine (induction medium) for 16 hours. Cells were washed thrice with 1 ml α -MEM and allowed to grow for 4 additional hours in 1 ml of 0.5% exhausted serum (prepared by diluting the induction medium X30 with α -MEM).

1 ng/ml FGF9 was added for 5 minutes and cells are then placed on ice. The cells were washed twice with ice-cold PBS and then lysed with 0.5 ml lysis buffer. The cells are scraped into an eppendorf tube, vortexed once and placed on ice for 10 minutes. The lysate was microcentrifuged 10 minutes at 4⁰C and the cleared lysate transferred into a fresh Eppendorf tube.

The protein content was determined by Bradford or DC protein assay (Bio-Rad, cat# 500-0116 - see manufacture instructions). Total protein aliquots, supplemented with 1/5 volume of 5x sample buffer, were boiled for 5 minutes and stored at -20⁰C until ready to load

on gel. In parallel an immunoprecipitation (IP) assay was performed, 10 µl anti- FGFR3 antibodies were added to the rest of the lysates and incubated for 4 hours at 4⁰C. 40 µl protein A-Sepharose was added and incubated for 1 hour at 4⁰C with continuous shaking. Afterwards, the mixture was microcentrifuged 15 seconds, and the fluid was aspirated,
5 carefully leaving a volume of ~30 µl above the beads. The beads were washed 3 times with 1 ml lysis buffer. At this step, the protease inhibitor mix is omitted from the buffer.

After the final wash, 15 µl of 5x sample buffer was added, samples were boiled 5 minutes and stored at -20⁰C until ready to load onto gel. Samples were loaded on 7.5% SDS-PAGE, cast on a Mini-PROTEAN II electrophoresis cell, and run at 100 V through the upper
10 gel and at 150 V through the lower gel. Proteins were transferred onto nitrocellulose sheet using the Mini trans-blot electrophoretic transfer cell at 100 V for 75 minutes or at 15 V overnight. The lower part of the total lysate Western blots was probed with anti-active MAPK (ERK) and the upper part is probed with anti-phosphotyrosine, both diluted 5x10³. The IP lysate Western blots were probed with anti-anti-phosphotyrosine (R&D Systems).
15 Hybridization was detected by ECL following the manufacturer's instructions.

BIAcore and proliferation analyses done at MorphoSys showed that among the new Fabs, MS-PRO54 is highly cross reactive with FGFR1. To further test the cross reactivity of the new Fabs, RCJ cells expressing either FGFR3ach (RCJ-M14; M14 on figure 9A) FGFR3 wild type (W11 on figure 9B), FGFR1 (R1-1 on figure 9C) or FGFR2 (R2-2 on figure 9D)
20 were incubated with increasing amount of MS-PRO54 and MS-PRO59 for 1 hour. FGF9 was added for 5 minutes and cell lysates were analyzed by Western for pERK activation (Figs. 8A-B, 9A-9D). Figure 8A shows that MSPRO2 and MSPRO12 block FGFR3 receptor activation in W11 and RCJ-FR3ach expressing cells. Furthermore MSPRO13 was able to block FGFR1 activation while none of the Fabs blocked FGFR2 activation. Figures 8B and
25 9A-9D show the results of several Fabs on RCJ expressing wildtype FGFR3 (8B) or the different FGFR types. MS-PRO29 appeared as the best FGFR3 blocker and was also effective in blocking FGFR1 (Fig. 9c); however, MS-PRO54 was the most effective Fab against FGFR1. None of the Fabs significantly inhibited FGFR2 activity. There are only a few amino acid residues, within the third Ig domain, that are shared by FGFR3 and FR1 but
30 not by FR2. Making mutants at these sites should clarify their role in Fab-receptor binding. Figure 8B depicts the dose effect of MS-PRO12, 29 and 13, stimulated with FGF9 and analyzed by Western blot using anti-ERK antibodies.

Example 8: Epitope mapping of selected Fabs

Constructs containing cDNAs that code for segments of the extracellular domain of FGFR3 were generated (Fig. 10). These include pChFR3^{D2}Fc that codes for Ig-like domain 2 of FGFR3 and pChFR3^{D2,3}Fc that codes for domain 2 and 3, both as human Fc fusions. The corresponding chimeric proteins, as well as the control hFR3exFc (containing domains 1, 2 and 3) were anchored to an ELISA plate coated with α human Fc antibody. A panel of 8 best Fabs, MSPRO2, 11, 12, 21, 24, 26, 28 and 29, were added, and bound Fab was determined with HRP- α human Fab (Fig. 11). The results in Fig. 11 demonstrate that MSPRO2 and 12 differ from the other tested Fabs. Both bind to the Ig like domain 2 while the others require domain 3 for binding. It was then tested whether or not Fabs that belong to the second group would distinguish the FGFR3IIIc isoform from the FGFR3IIIb from. FDCP-FR3IIIb or FDCP-FR3IIIc cells were incubated in the presence of 1.25 ng/ml FGF9 with increasing doses of either MSPRO12 or MSPRO29. Ly6.3 was included as control. After 2 days in culture, cell proliferation was measured with the XTT reagent. Clearly, MSPRO29 was completely ineffective against the IIIb isoform (Fig. 12). In contrast, MSPRO12 (square on hatched or solid lines) was equally effective against both isoforms. These data suggest that residues that differ between the two isoform are critical for MSPRO29 (and probably also for the other Fabs in the same group) FGFR3 binding.

Domains in FGFR3 recognized by the new Fabs.

In agreement with data generated at Morphosys, MSPROs can be divided into 2 groups, one that includes Fabs that bind the FGFR3 Ig II domain (MS-PRO2 and 12) and a second with members that require the Ig III domain for binding (MSPRO11, 21, 24, 26, 28, and 29). To classify the new Fabs obtained from the last screen performed at Morphosys, as well as some previously obtained Fabs, a proliferation assay of FDCP cells expressing either FR3IIIb or FR3IIIc was performed. The cells were incubated in the presence of 10 (IIIb) or 5 (IIIc) ng/ml FGF9 with increasing doses of the indicated Fabs. After 2 days in culture, cell proliferation was measured with the XTT reagent.

In agreement with Morphosys data, MSPRO59 efficiently inhibited both FDCP-FR3IIIb and FDCP-FR3IIIc cells while MSPRO21, 24, 26, 28, 29 and 54 inhibited FDCP-FR3IIIc proliferation only (Fig. 13).

Example 9: Bone culture

Radiolabeled MSPRO29 was used to determine whether or not MSPRO Fabs can enter the bone growth plate.

To determine the effect of iodination on Fab activity, 50 µg of MSPRO29 was first labeled with cold iodine using Pierce IodoGen coated tubes. The process was carried out either without iodine, with 0.04 mM or with 1 mM NaI. MSPRO29 was then purified through a sephadex G-50 column. The ability of the modified Fab to bind FGFR3 was determined by ELISA. MaxiSorp wells were coated with anti-human Fc. FGFR3/Fc was then anchored to the wells. In parallel, a similar set of wells was left in blocking buffer only (no FR3/Fc). The unmodified (no I) or the modified MSPRO29 (low for that labeled with 0.04 mM NaI and high for that labeled at 1 mM NaI; 2 G-50 fractions each) were added at approximately 5 µg/well and binding was measured with anti-human Fab. Fresh MSPRO29 and buffer alone were included as controls.

MSPRO29 labeled in the presence of 0.04 mM NaI showed equal binding to the receptor as compared to the control unmodified Fab (Fig. 14). MSPRO29 labeled in the presence of 1 mM NaI also bound the receptor, however, the noise level of this sample was as high as the signal itself suggesting that at the high Iodide concentration the Fab was inactivated.

The neutralizing activity of the modified Fab was tested in a proliferation assay using FDCP-FR3 (C10) (Fig. 15). FDCP-FR3 (C10) cells were treated with the indicated amount of labeled or unlabeled (without I) MSPRO29. The proliferation rate of the cells was determined by XTT analysis. The Fab was labeled at either 0.04 mM (Low) or 1 mM NaI (High). Two G-50 fraction (I and II) were analyzed. Fresh MSPRO29 and buffer alone (mock) were included as controls.

This showed that MSPRO29, labeled at 0.04 mM NaI, kept its activity almost entirely while that labeled at 1 mM NaI lost its activity completely. MS-PRO29 was labeled with 1 mCi ¹²⁵I. The specific activity of the Fab was 17 µCi/µg.

Ex vivo distribution of ¹²⁵I MSPRO29 in bone culture

Femora prepared from newborn mice were incubated with 2 µg ¹²⁵I-MSPRO29 (17 µCi/µg) or ¹²⁵I-Ly6.3 (20 µCi/µg) for 1, 3 or 5 days in culture. Then, sections were processed for radiomicroscopy. After 3 days in culture, MSPRO29 was predominantly visualized at the higher hypertrophic zone and to a lesser extent at the secondary ossification region (Figs. 16A-16F). Hematoxylin-eosin staining of growth plate treated with radiolabelled MS-PRO29

or Ly6.3 (Figs. 16A and 16D, respectively) x100 magnification. Radiomicroscopic sections of growth plate treated with radiolabelled MS-PRO29 or Ly6.3 (Figs. 16B and 16E) at X100 magnification. Figs. 16C and 16F are the same as Figs. 16B and 16E but at x400 magnification. The arrow in figure 16C indicates the location of the specific binding of the radiolabelled MS-PRO29 to the higher hypertrophic zone of the growth plate.

As compared to MSPRO29, the control Ly6.3 Fab was weakly and evenly distributed throughout the whole growth plate. At day 1 in culture, the signal was weaker but with similar distribution pattern. This distribution also holds at 5 days in culture with a less favorable signal to noise ratio (data not shown). This clearly demonstrates that MSPRO29 binds FGFR3 in our target organ.

Example 10: Neutralizing Activity on Constitutively Activating Receptors

The inhibitory activity of MSPRO antibodies on ligand-dependent and ligand-independent FDCP proliferation expressing FGFR3 Achondroplasia mutation was tested.

A proliferation assay was carried out using FDCP-FR3wt (C10) or FDCP-FR3ach cells incubated with 1.25 or 5 ng/ml FGF9 respectively and with increasing amounts of MSPRO54 or MSPRO59. As shown in Fig. 17, both MSPRO54 and 59 antibodies neutralize the mutant receptor. Few of the FDCP-FR3ach acquired ligand independent cell proliferation due to the high expression of the FGFR3ach mutation.

FDCP cells that express the achondroplasia FGFR3 (FDCP-FR3ach) and proliferate independently of ligand were incubated with the indicated amount of MSPRO12, 29, 59 or the control Ly6.3. Two days later, cell proliferation was determined by an XTT analysis. When inhibition of cell proliferation by the MS-PRO 12, 29, 54 and 59 were tested, only the antibodies 12 and 59 (the only Ab which recognized D2 domain) inhibited the ligand-independent cell proliferation (Figs. 18A and 18B). Previously, the activity of MSPRO Fabs generated in the first and second screens (MSPRO1-15 and MSPRO21-31, respectively) by XTT analysis of FDCP-FR3ach cells were tested. These cells, when generated, show ligand-dependent proliferation. With time, however, they acquired a ligand-independent ability to proliferate. Accordingly, neutralizing Fabs were able to block the ligand-dependent, but not the ligand-independent, proliferation of these cells. To show whether this is also true for the new batch of Fabs, FDCP-FR3ach cells, which is the FDCP-derived cell line that expresses a constitutive FGFR3-G380R (Ach), were subjected to XTT analysis in the presence of MSPRO59 and MSPRO29. Surprisingly, and in contrast to the ineffective MSPRO29,

MSPRO59 completely blocked cell proliferation (Fig. 18B). Whether other Fabs that, like MSPRO59, bind to the second Ig like domain would also inhibit FDCP-FR3ach cell proliferation was tested next. Indeed, it was found that MSPRO12 strongly inhibits the constitutive cell proliferation. However, the third member in this family, MSPRO2, had no effect on either the constitutive or the ligand-dependent cell growth, suggesting that the Fab may have lost its neutralizing activity (not shown).

Example 11: RCS Chondrocyte Culture

Effect of Fabs on growth arrest of RCS Chondrocytes

RCS is a rat chondrosarcoma derived cell line expressing preferentially high levels of FGFR2 and FGFR3 and low levels of FGFR1 (Sahni, 1999). In this cell line FGFR functions as an inhibitor of cell proliferation similar to its expected role in the achondroplasia phenotype. Analysis of RCS cell proliferation mediated by the addition of different molecules of the invention, showed that MSPRO54 and MSPRO59 were able to restore cell proliferation.

The screening was performed on RCS parental cells in 96 wells plates. Cells were seeded at a concentration of 2,000 cells/well. The following day 10ng/ml FGF-9 and 5µg/ml heparin were added to the cells. 50ug/ml of the antibodies were added. Positive and negative controls for cell proliferation are included in this assay at the same concentrations as the tested molecules. On the fourth day of incubation, plates were observed under the microscope. If all cells were viable, no quantitative assay to measure the effect of the variants was performed. If cell death was observed, the Cy-Quant assay kit is used to measure the amount of the cells. The results are measured in a fluoro ELISA reader. Figure 19 shows the ELISA results in bar graph form. Untreated cells are shown speckled, ligand treated cells are shown in gray, control antibody (LY6.3)treated cells are in black while MSPRO54 and MSPRO59 treated cells are shown in hatched or checkered bars, respectively.

Example 12: *Ex vivo* Bone Culture

The femoral bone cultures were performed by excising the hind limbs of 369-mice, heterozygous or homozygous mice for the achondroplasia G369C mutation (age P0). The limbs were carefully cleaned up from the surrounding tissue (skin and muscles) and the femora exposed. The femora were removed and further cleared from tissue remains and ligaments. The femora were measured for their initial length, using a binocular with an eyepiece micrometer ruler. The bones were grown in 1 ml of medium in a 24 well tissue

culture dish. The growing medium is α -MEM supplemented with penicillin (100 units/ml), streptomycin (0.1 mg/ml) and nystatin (12.5 units/ml). In addition, the medium contains BSA (0.2%), α -glycerophosphate (1 mM) and freshly prepared ascorbic acid (50 μ g/ml). The bones were cultured for 15 days. Measurements of bone length and medium replacement
5 were performed every three days.

At the end of the experiment, the growth rate of the bones was determined. The growth rate of bones is calculated from the slope of a linear regression fit on the length measurements obtained from day 3 to 9.

The results shown in Fig. 20 demonstrate a dose dependent increase in the growth rate
10 of bones treated with MS-PRO 59 in comparison to non-relevant control LY6.3 Fab. The LY6.3-treated control femurs, marked with a circle, grew at the slowest rate. The MSPRO59 treated femurs exhibited a higher growth rate, with the optimal rate achieved at MSPRO59 concentration of 100ug/ml (square) while the higher cocentration (400ug/ml, triangle) showed inhibition. Moreover, the growth rates achieved by 400 microgram/ml of MSPRO59
15 doubled in comparison to the control Ab (3.55 U/day as compared to 1.88 U/day, respectively). This experiment shows the neutralizing effect of the MSPRO59 antibody on constitutively active FGFR3, in an *ex vivo* model.

Example 13: *In-vivo* trials

FDCP-FR3ach cells, but not FDCP (control) cells, were found to be tumorigenic when
20 injected into nude mice. Each of 9 mice received two sub-cutaneous injections with different amount of transfected cells. Fourteen days after injection, progressively growing tumors started to appear at the site of FDCP-FR3ach injection but not at the FDCP site of injection. External examination of the tumors showed a high vascular capsule. ¹²⁵I-labeled MSPRO59 and LY6.3 were injected I.P. into nude mice carrying the FDCP-FR3ach derived tumor. The
25 tumors were dissected 4 and 24 hrs later and radioactivity was measured. Concentration of MSPRO59 Abs in FDCP-FR3ach derived tumors is shown in Fig. 22.

Example 14: Animal Model for Bladder Carcinoma

Recent studies have shown that the IIIb isoform of FGFR3 is the only form expressed in bladder
30 carcinoma, in particular an FGFR3 with an amino acid substitution wherein Serine 249 is replaced by Cysteine (S249C). The progression of the cancer is believed to be a result of the constitutive activation resulting from this amino acid substitution. In order to create the FGFR3 IIIb form, we isolated the IIIb region of FGFR3 from HeLa cells and generated a full length FGFR3IIIb isoform in

pLXSN. Retroviruses, expressing either normal FGFR3 (FR3wt) or mutant FGFR3 (FR3-S249C) were produced and used to infect FDCP cells. Stable pools were generated and further used for *in-vitro* and *in-vivo* experiments.

A. MSPRO-59 reduces tumor size in mice

5 Twelve nude mice were injected with 2×10^6 FDCP-S249C cells subcutaneous at 2 locations, one on each flank. A week later MSPRO59 was administered i.p. at 400ug per mouse (3 mice in total), followed by 3 injections of 275 ug each, in 2 to 3 days intervals. Following 24 and 26 days the tumor size was measures. Figure 23 shows the inhibitory effect of MSPRO59 on tumor size.

B. Treating FDCP-S249C-derived tumors with MSPRO59

10 Nude mice (3 in each group), were injected subcutaneous at 2 locations, one on each flank, with 2×10^6 FDCP-S249C cells each. A week later, 400 or 80 μ g MSPRO59 were injected IP. Three days later, mice were injected with 400 μ g followed by 5 additional injections with 275 μ g MSPRO59, each, every 3 or 4 days. Mice initially treated with 80 μ g MSPRO59 were similarly given an additional 80 μ g MSPRO59 followed by 5 injections with
15 50 μ g MSPRO59 at the same schedule. Mice injected with PBS were used as control. Tumors typically appeared three weeks post injection of the cells. Tumor volume was estimated from measurements in 3 dimensions at 16,20, 23 or 32 days post cell injection.

The data indicate that there is both a delay in tumor appearance and an inhibitory effect on tumor progression in the treated mice (data not shown). This indicates that these FGFR3
20 inhibitors are potent *in-vivo*.

These data may also help us understand the mechanism by which the S249C-derived tumors were developed. Since we are using pools of cells, treatment with MSPRO59 inhibited the susceptible cells, leading to delay in tumor appearance. However, over time, the resistant cells survived and proliferated, giving rise to a solid tumor.

25 C. MSPRO59 inhibits FDCP-FR3ach380 derived tumor growth.

Nude mice were injected subcutaneously in the flank with 2×10^6 FDCP-FR3ach380 cells, each. Treatment with MSPRO59 began at the day of tumor appearance. Three mice were treated with a known tyrosine kinase inhibitor (TKI -50 mg/Kg/injection) and three with 400 μ g followed by 3 additional injections with 300 μ g MSPRO59, every 3 or 4 days. Three
30 mice were treated with PBS alone as control. The tumor size was estimated as before at the indicated days after cell injection. The dose schedule is shown in Table 8 below.

Table 8

	Days After FDCP-FR3 ^{ach380} Cell Injection			
	21	25	28	31
MSPRO59 (μg)	400μg	300μg	300μg	300μg
PBS (μl)	50	50	50	50

D. MSPRO59 inhibits FDCP-S249C induced tumor growth

To overcome the instability of the FDCP-derived pools, clones from each pool FDCP-S249C clone #2) were isolated and characterized. These clones were tested in an XTT proliferation assay and were shown to be inhibited by MSPRO59. 2x10⁶ cells from each clone were injected into nude mice. Tumors appeared 18-30 after injection.

FDCP-S249C clone #2 was injected subcutaneously on the flank. A week later mice were injected with 280 μg MSPRO59 single chain (SC) I.P. every day. Mice injected with PBS were used as control. Tumor volume was estimated from measurements in 3 dimensions at 18 or 24 days post cell injection. An apparent inhibition of tumor growth by MSPRO59(SC) was observed in tumors derived from clone 2 (data not shown).

Having now fully described this invention, it will be appreciated by those skilled in the art that the same can be performed within a wide range of equivalent parameters, concentrations, and conditions without departing from the spirit and scope of the invention and without undue experimentation.

While this invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications. This application is intended to cover any variations, uses, or adaptations of the inventions following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth as follows in the scope of the appended claims.

All references cited herein, including journal articles or abstracts, published or corresponding U.S. or foreign patent applications, issued U.S. or foreign patents, or any other references, are entirely incorporated by reference herein, including all data, tables, figures, and text presented in the cited references. Additionally, the entire contents of the references cited within the references cited herein are also entirely incorporated by references.

Reference to known method steps, conventional methods steps, known methods or conventional methods is not in any way an admission that any aspect, description or embodiment of the present invention is disclosed, taught or suggested in the relevant art.

5 The foregoing description of the specific embodiments will so fully reveal the general nature of the invention that others can, by applying knowledge within the skill of the art (including the contents of the references cited herein), readily modify and/or adapt for various applications such specific embodiments, without undue experimentation, without departing from the general concept of the present invention. Therefore, such adaptations and modifications are intended to be within the meaning and range of equivalents of the disclosed
10 embodiments, based on the teaching and guidance presented herein. It is to be understood that the phraseology or terminology herein is for the purpose of description and not of limitation, such that the terminology or phraseology of the present specification is to be interpreted by the skilled artisan in light of the teachings and guidance presented herein, in combination with the knowledge of one of ordinary skill in the art.

15

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WHAT IS CLAIMED IS:

1. A molecule comprising the antigen binding portion of an isolated antibody which has an increased affinity for a receptor protein tyrosine kinase and which blocks constitutive activation of said receptor protein tyrosine kinase.
- 5 2. The molecule according to claim 1, wherein said molecule binds to the extracellular domain of the receptor protein tyrosine kinase.
3. The molecule according to claim 1 wherein the antibody binds the dimeric form of the receptor.
- 10 4. The molecule according to claim 1, wherein the receptor protein tyrosine kinase is selected from the group consisting of EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR- α , PDGFR- β , CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROS, and Alk, and heterodimeric combinations thereof.
- 15 5. The molecule according to claim 4, wherein said receptor protein tyrosine kinase is a fibroblast growth factor receptor (FGFR).
6. The molecule according to claim 5, wherein said FGFR is FGFR3.
7. The molecule according to claim 1, comprising a VL region and a VH region, respectively, selected from the group consisting of SEQ ID NO: 43 and 51; SEQ ID
20 NO: 44 and 58; and SEQ ID NO: 42 and 60.
8. The molecule according to claim 1, comprising V_L-CDR3 and V_H-CDR3 regions, respectively, selected from the group consisting of SEQ ID NO: 9 and 8; SEQ ID NO: 13 and 12; and SEQ ID NO: 25 and 24.
- 25 9. A pharmaceutical composition, comprising, as an active ingredient, the molecule according to claim 1 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.
10. An isolated nucleic acid molecule, comprising SEQ ID NO: 74, 70, 75, 67, 64, 71, 62, 65, 73, 69, 76, or a nucleotide sequence hybridizing under high stringency conditions thereto.

11. An isolated nucleic acid molecule, comprising SEQ ID NO: 84, 85, 89, 78, 79, 86, 80, 87, 82, 83, 91, or a nucleotide sequence hybridizing under high stringency conditions thereto.
- 5 12. An isolated nucleic acid molecule, comprising nucleotides encoding a VL region and a VH region, respectively, selected from the group consisting of SEQ ID NO: 74 and 84; SEQ ID NO: 70 and 85; SEQ ID NO: 75 and 89; SEQ ID NO: 67 and 78; SEQ ID NO: 64 and 79; SEQ ID NO: 71 and 86; SEQ ID NO: 62 and 80; SEQ ID NO: 65 and 87; SEQ ID NO: 73 and 82; SEQ ID NO: 69 and 83; and SEQ ID NO: 76 and 91.
- 10 13. A vector comprising a nucleic acid molecule according to any one of claims 10-12.
14. A host cell transformed with the vector according claim 13.
15. A molecule comprising the antigen-binding portion of an antibody which binds to a fibroblast growth factor receptor (FGFR) and which blocks ligand-dependent activation of said FGFR.
- 15 16. The molecule according to claim 15, wherein said molecule binds to the extracellular domain of the FGFR.
17. The molecule according to claim 16, wherein the FGFR is FGFR3.
18. The molecule according to claim 17, comprising a VL region and a VH region, respectively, selected from the group consisting of respectively, selected from the
20 group consisting of SEQ ID NO: 38 and 52; SEQ ID NO: 36 and 47; SEQ ID NO: 33 and 48; SEQ ID NO: 39 and 53; SEQ ID NO: 34 and 54; and SEQ ID NO: 45 and 55.
19. The molecule according to claim 17, comprising V_L-CDR3 and V_H-CDR3 regions, respectively, selected from the group consisting of SEQ ID NO: 11 and 10; SEQ ID
25 NO: 15 and 14; SEQ ID NO: 17 and 16; SEQ ID NO: 19 and 18; SEQ ID NO: 21 and 20; and SEQ ID NO: 23 and 22.
20. A pharmaceutical composition, comprising the molecule according to claim 15 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.
21. A kit comprising the molecule of claim 1 or 15 and at least one reagent suitable for
30 detecting the presence of said molecule when bound to said receptor protein tyrosine kinase and instructions for use.

22. A method for treating or inhibiting a skeletal dysplasia or a craniosynostosis disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 9 or 20 to a subject in need thereof.
23. The method according to claim 22, wherein the skeletal dysplasia is selected from the group consisting of achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia.
24. The method according to claim 23 wherein the skeletal dysplasia is achondroplasia.
25. The method according to claim 22 wherein the craniosynostosis disorder is Muenke coronal craniosynostosis or Crouzon syndrome with acanthosis nigricans.
26. The method according to claim 22, wherein the disorder is associated with constitutive activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 9.
27. The method according to claim 22, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 20.
28. A method for treating or inhibiting a cell proliferative disease or disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 9 or 20 to a subject in need thereof.
29. The method according to claim 28, wherein the cell proliferative disease or disorder is tumor progression.
30. The method according to claim 29, wherein the tumor progression is the progression of transitional cell carcinoma.
31. The method according to claim 29, wherein the tumor progression is the progression of osteo or chondrosarcoma.
32. The method according to claim 29, wherein the tumor progression is the progression of multiple myeloma.
33. The method according to claim 29, wherein the receptor protein tyrosine kinase is FGFR3 and the tumor progression is the progression of mammary carcinoma.

34. The method according to claim 28, wherein the disorder is associated with the action of a constitutively activated receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 9.
- 5 35. The method according to claim 28, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 20.
- 10 36. An immunoglobulin molecule that has an increased affinity for a receptor protein kinase and comprising a V_L and V_H sequence, respectively, selected from the group consisting of SEQ ID NO: 43 and 51; SEQ ID NO: 44 and 58; SEQ ID NO: 42 and 60; SEQ ID NO: 38 and 52; SEQ ID NO: 36 and 47; SEQ ID NO: 33 and 48; SEQ ID NO: 39 and 53; SEQ ID NO: 34 and 54; and SEQ ID NO: 45 and 55.

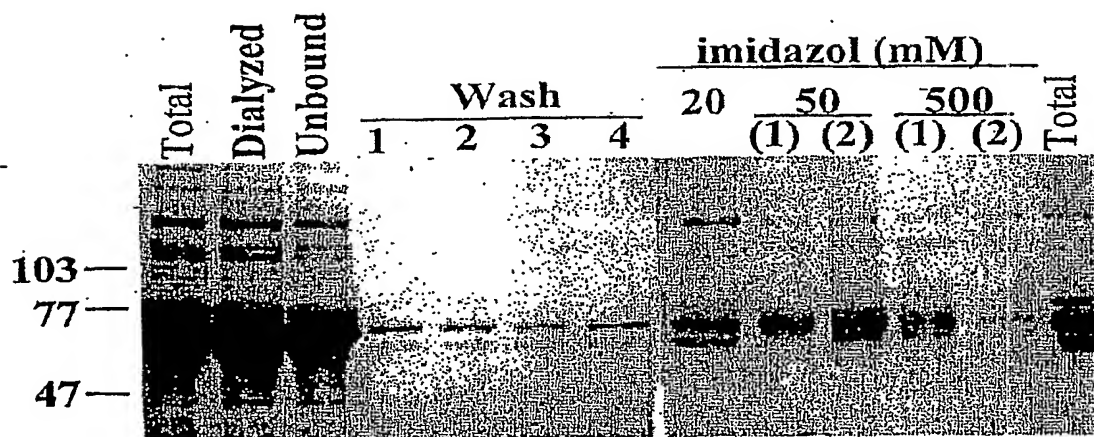


FIG. 1

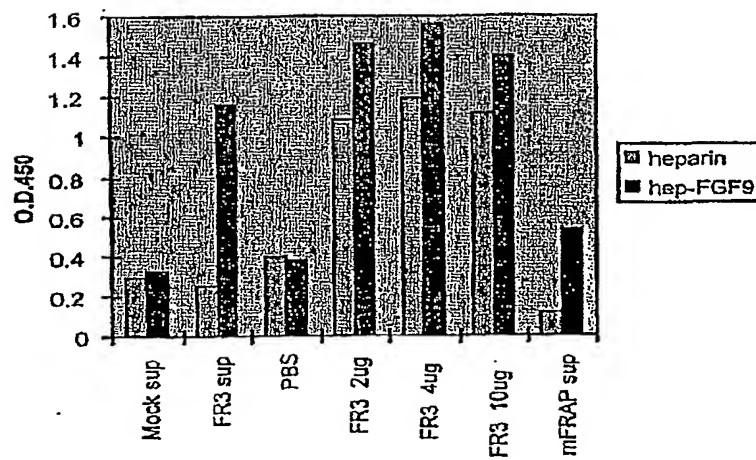


FIG. 2

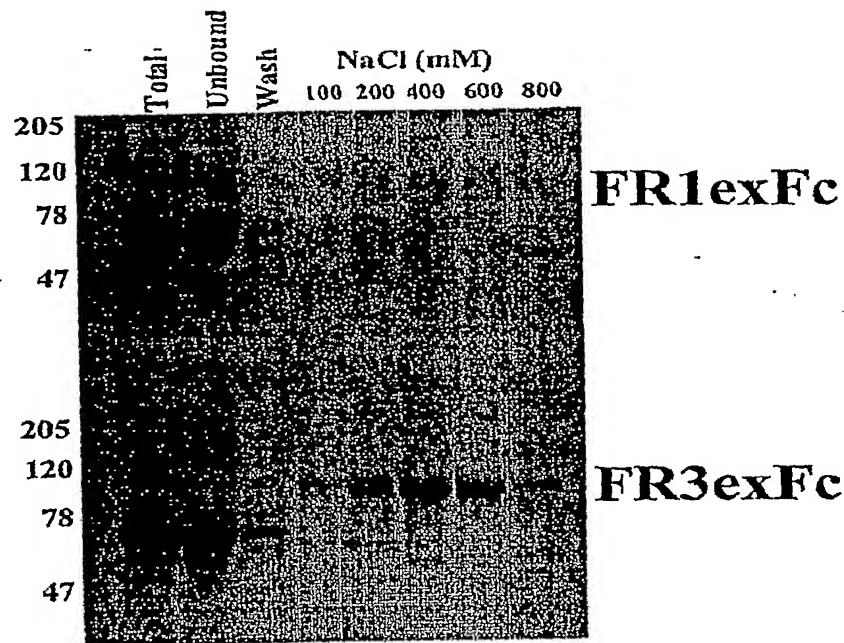


FIG. 3

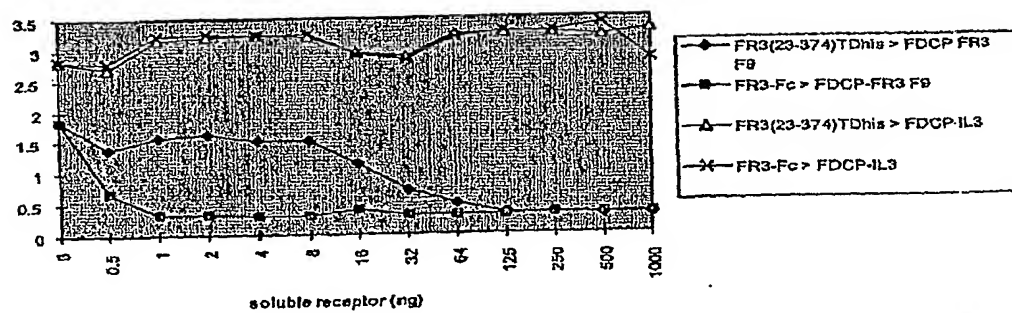


FIG. 4

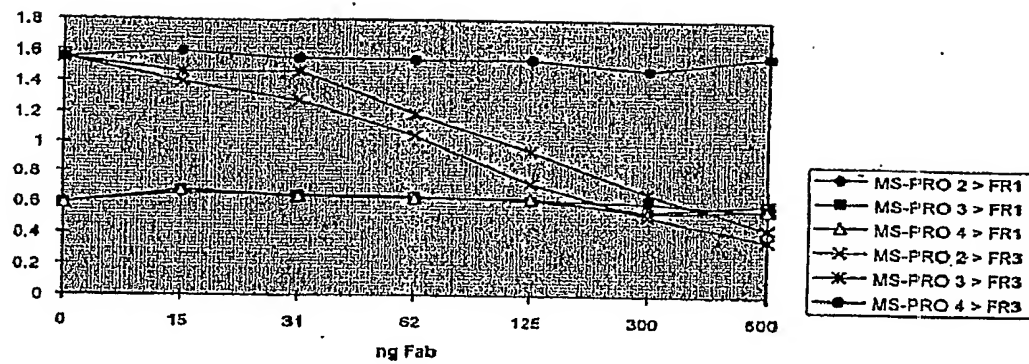


FIG. 5

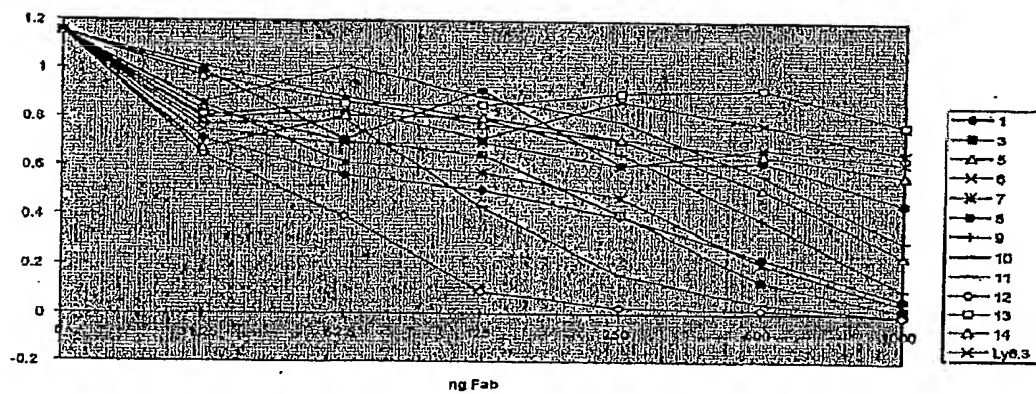


FIG. 6

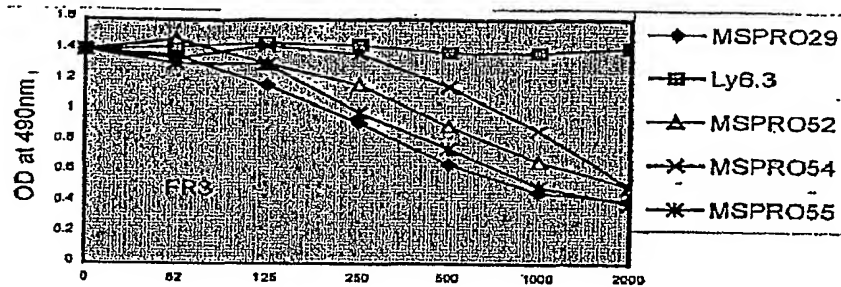


FIG. 7A

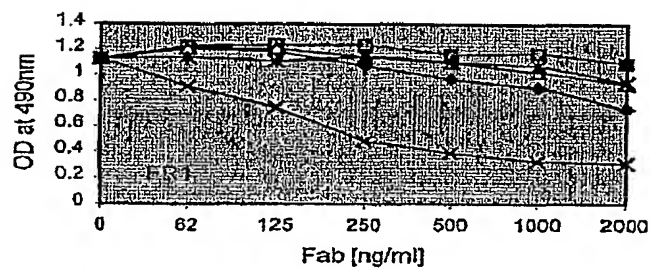


FIG. 7B

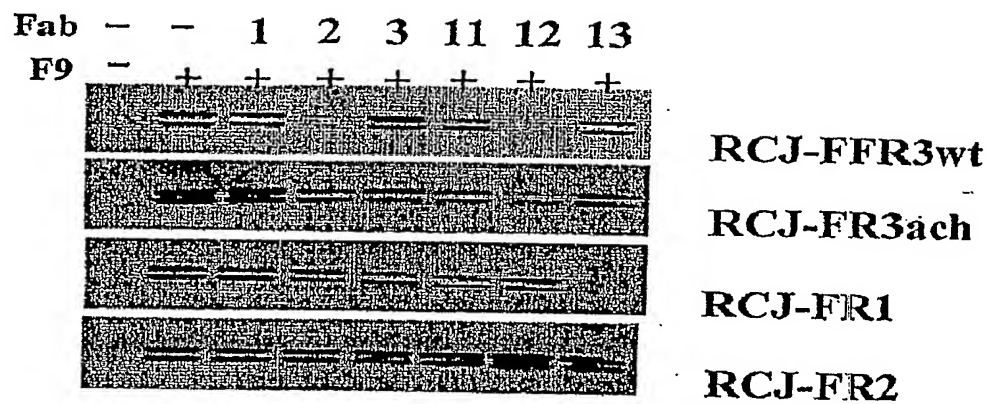


FIG. 8A

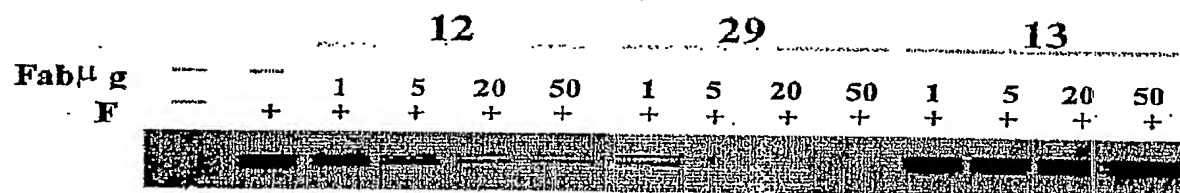
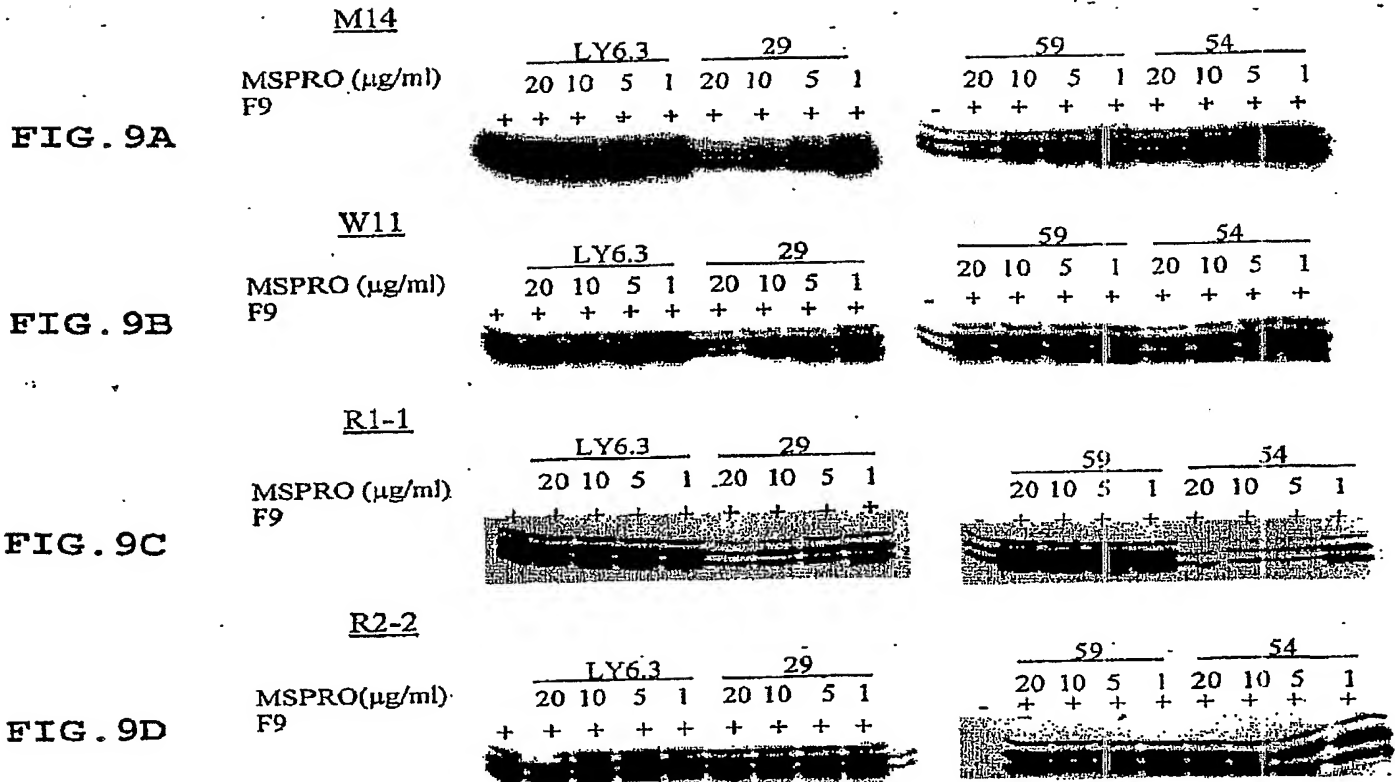


FIG. 8B



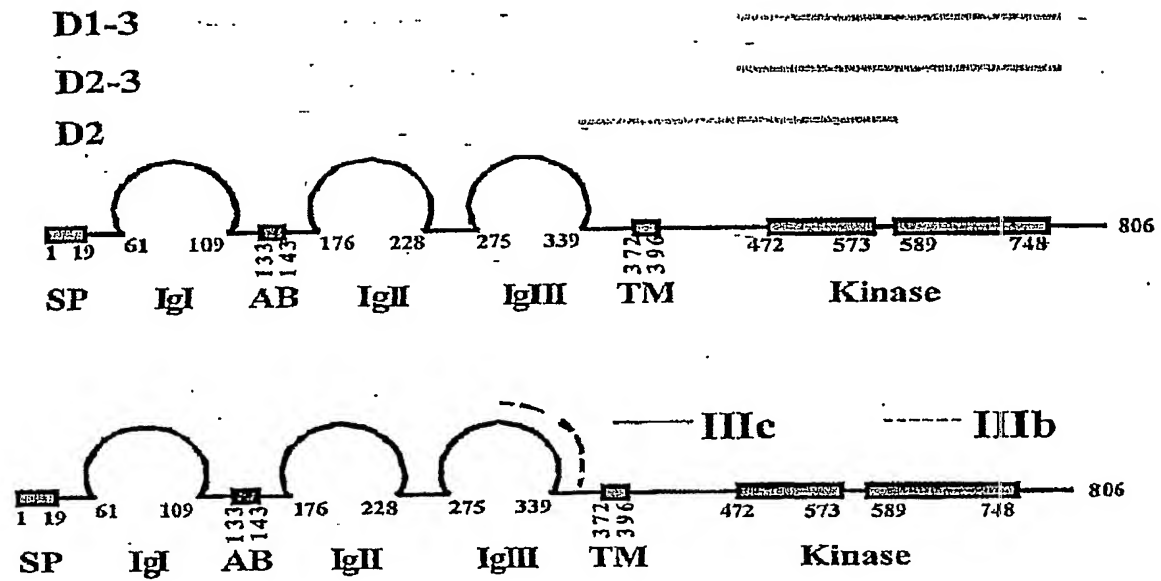


FIG. 10

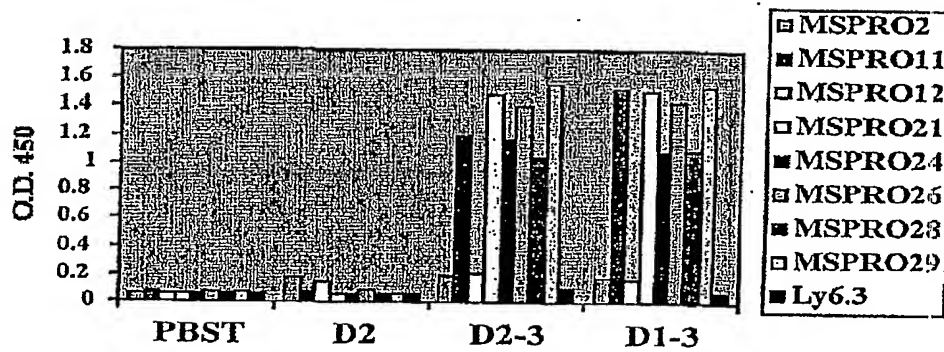


FIG. 11

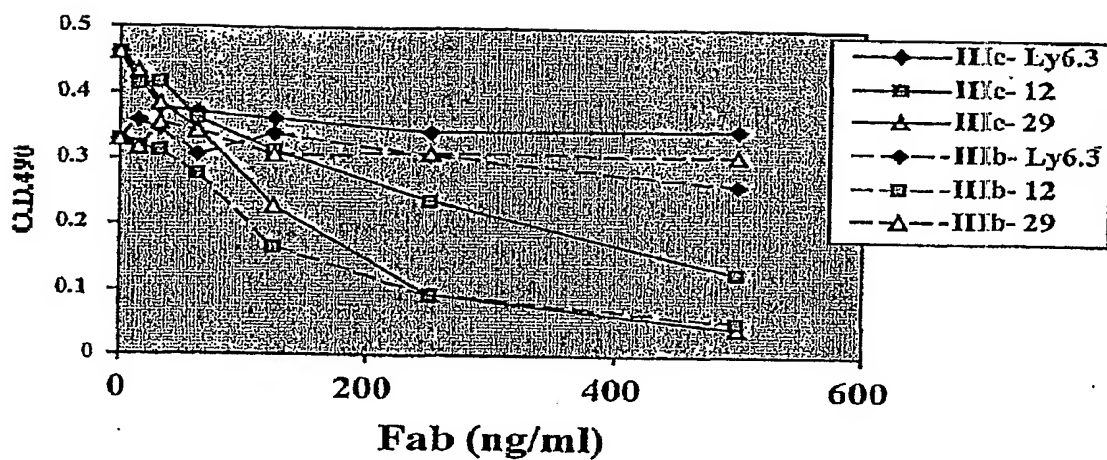


FIG. 12

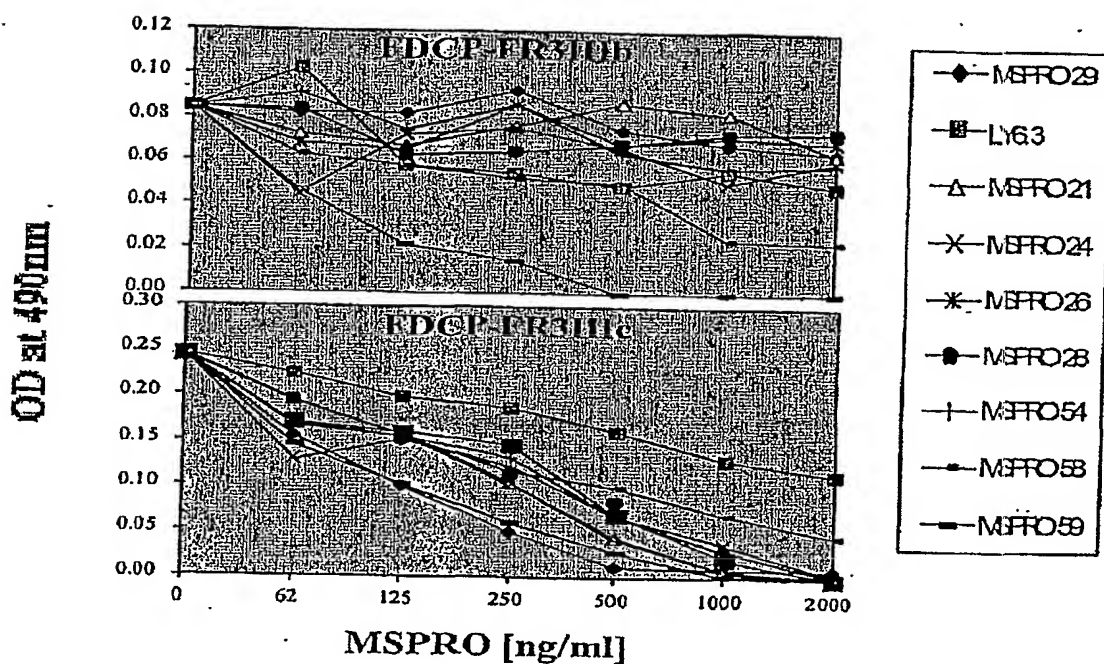


FIG. 13

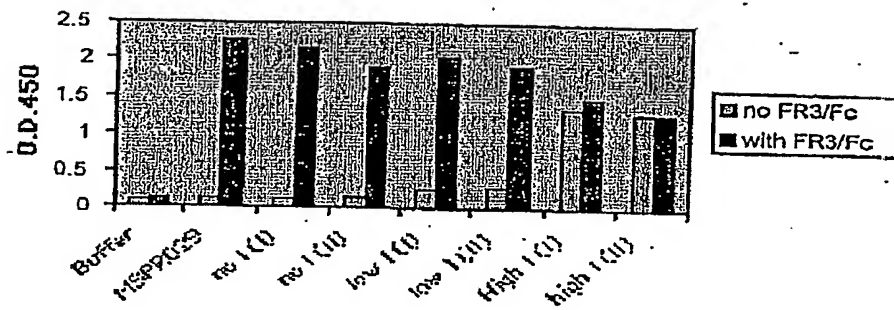


FIG. 14

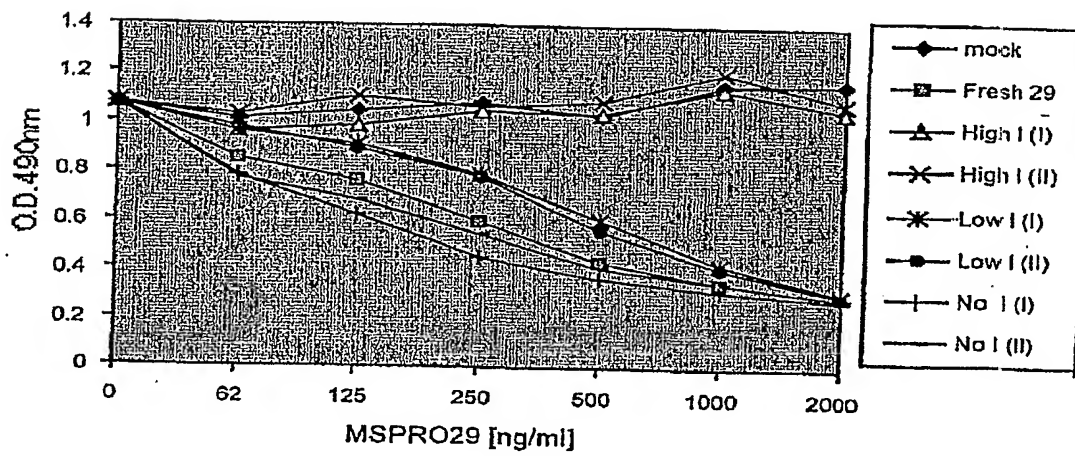


FIG. 15

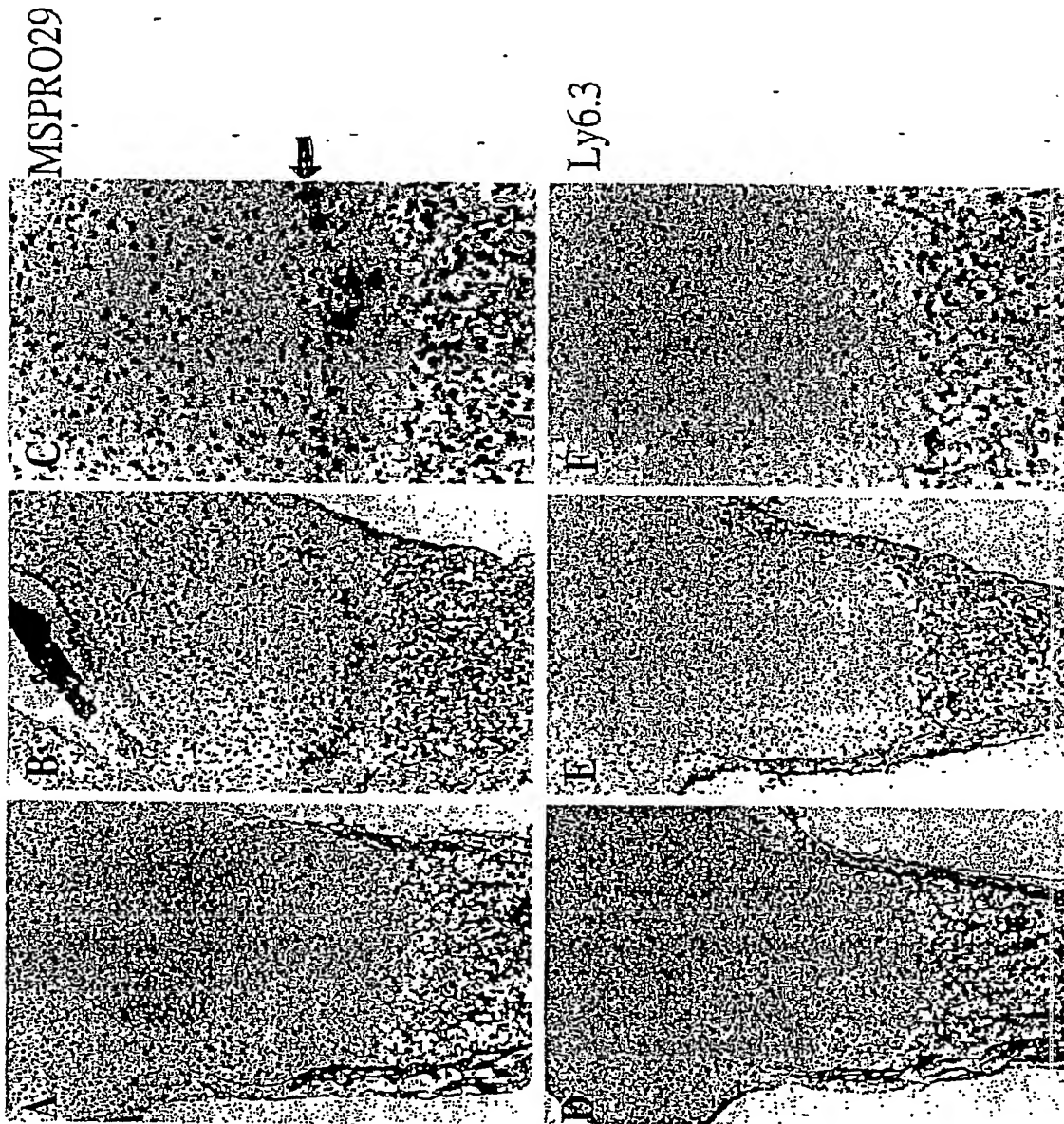


FIG.16

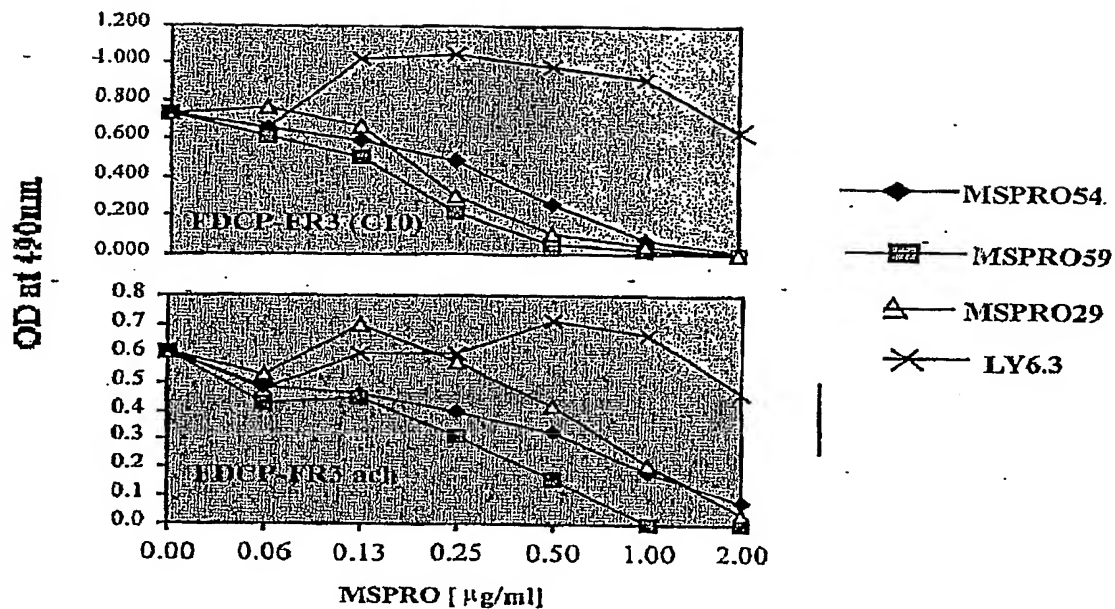


FIG. 17

FIG. 18A

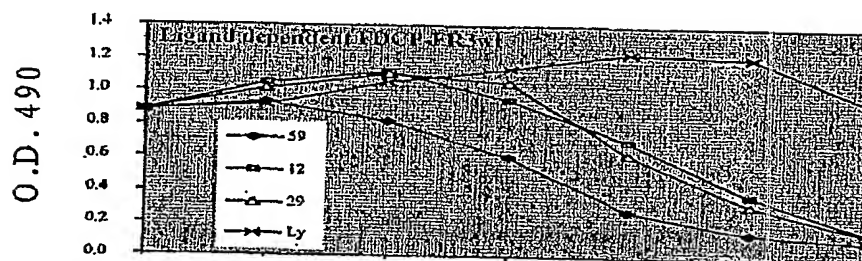
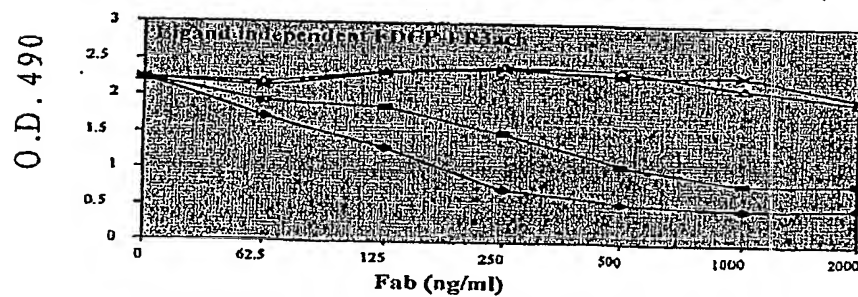


FIG. 18B



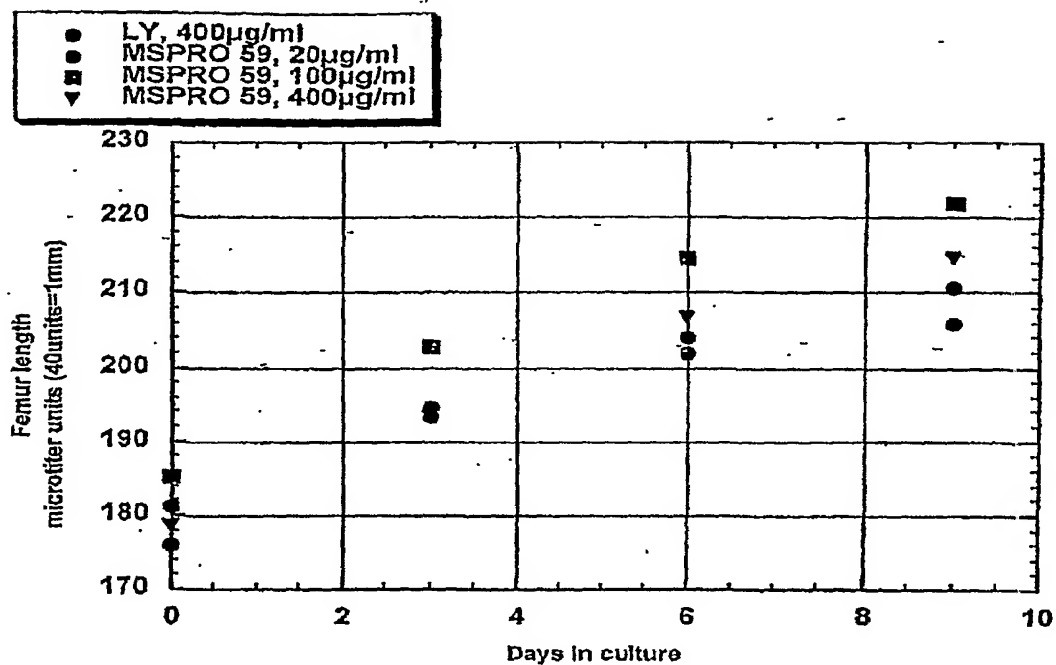


FIG. 19

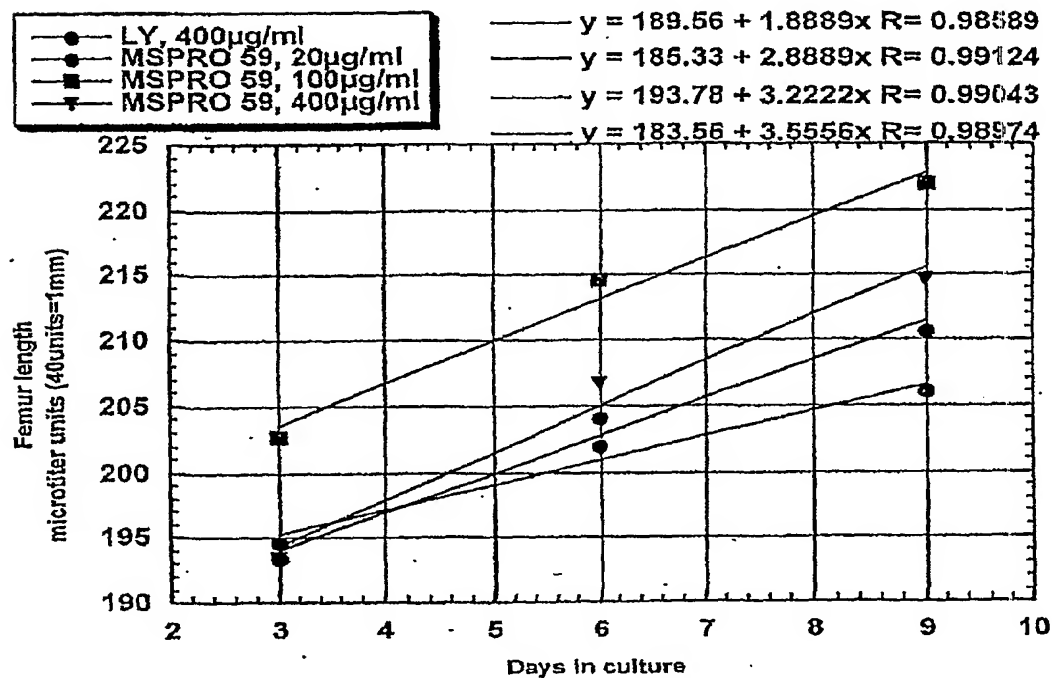


FIG. 20

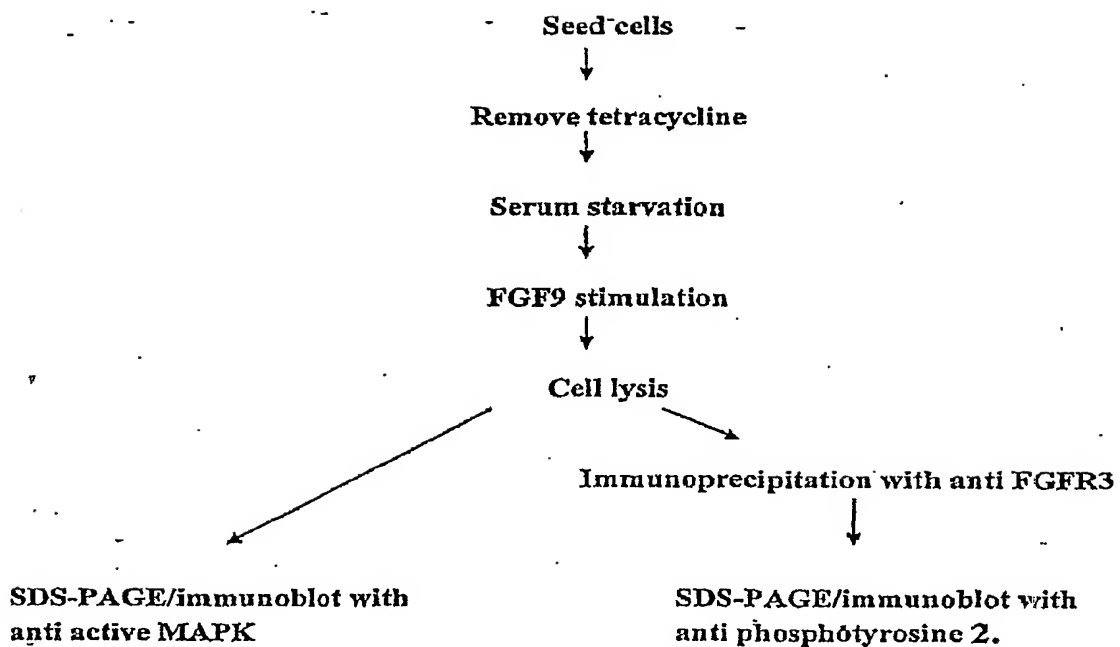


FIG. 21

FIG. 22A

O.D. 490

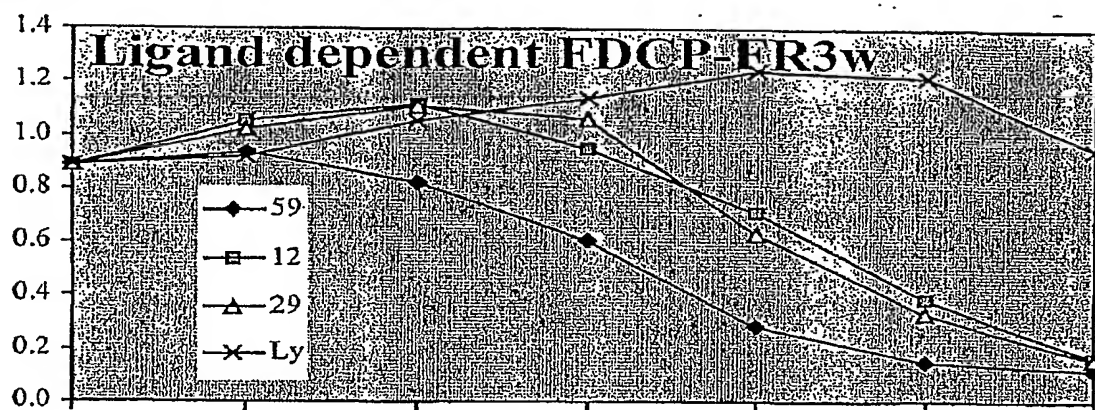
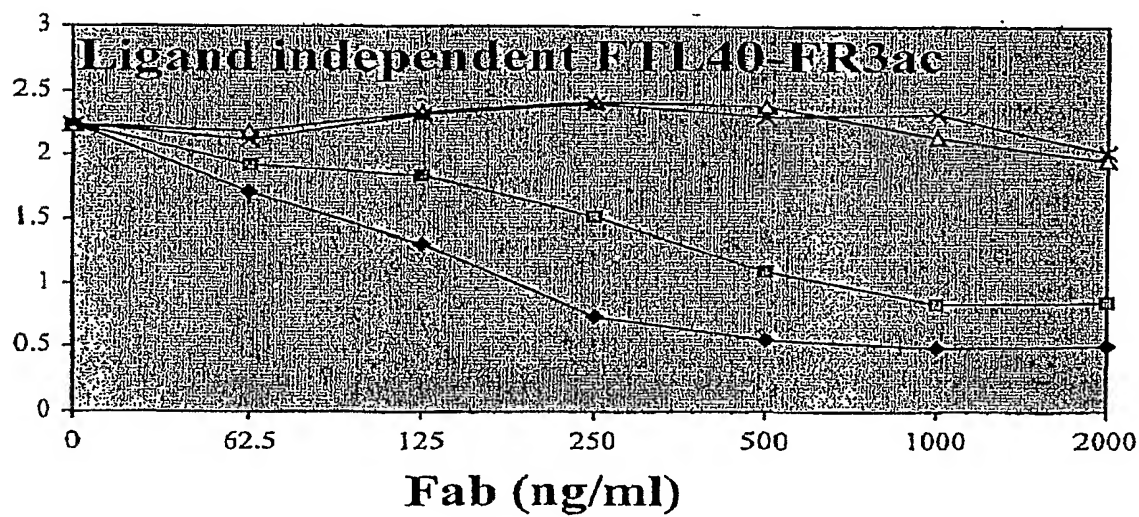


FIG. 22B



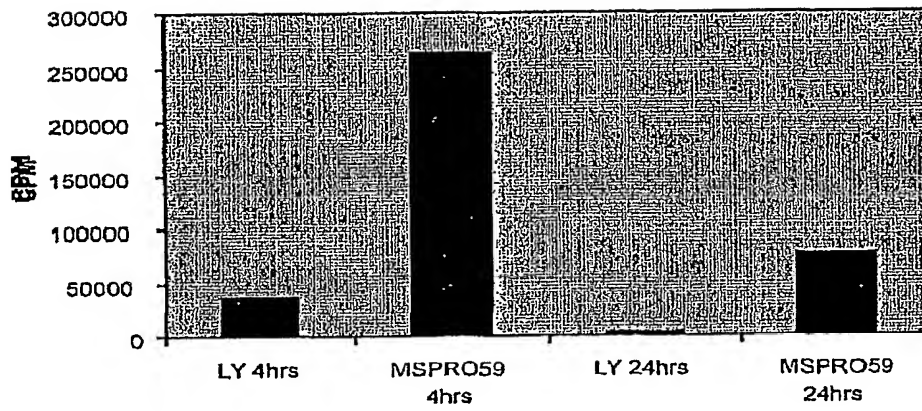


FIG. 23

Figure 24

Binding of Fab Miniantibodies to FGFR3-Fc and FGFR1-Fc (ELISA)

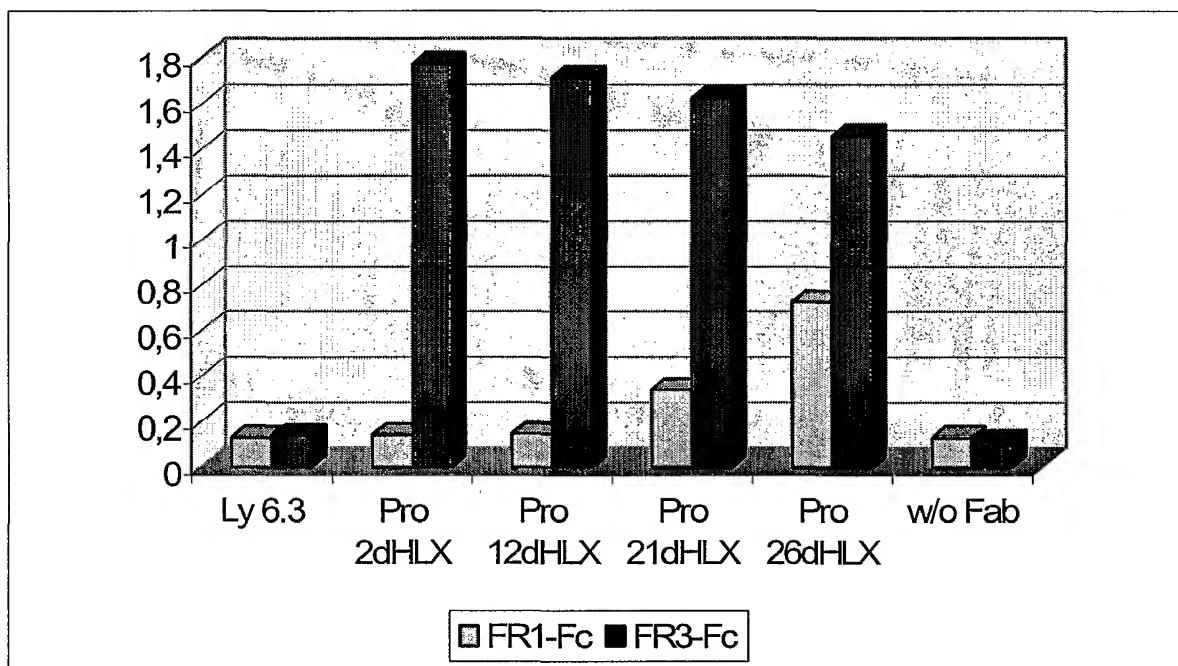


Figure 25A

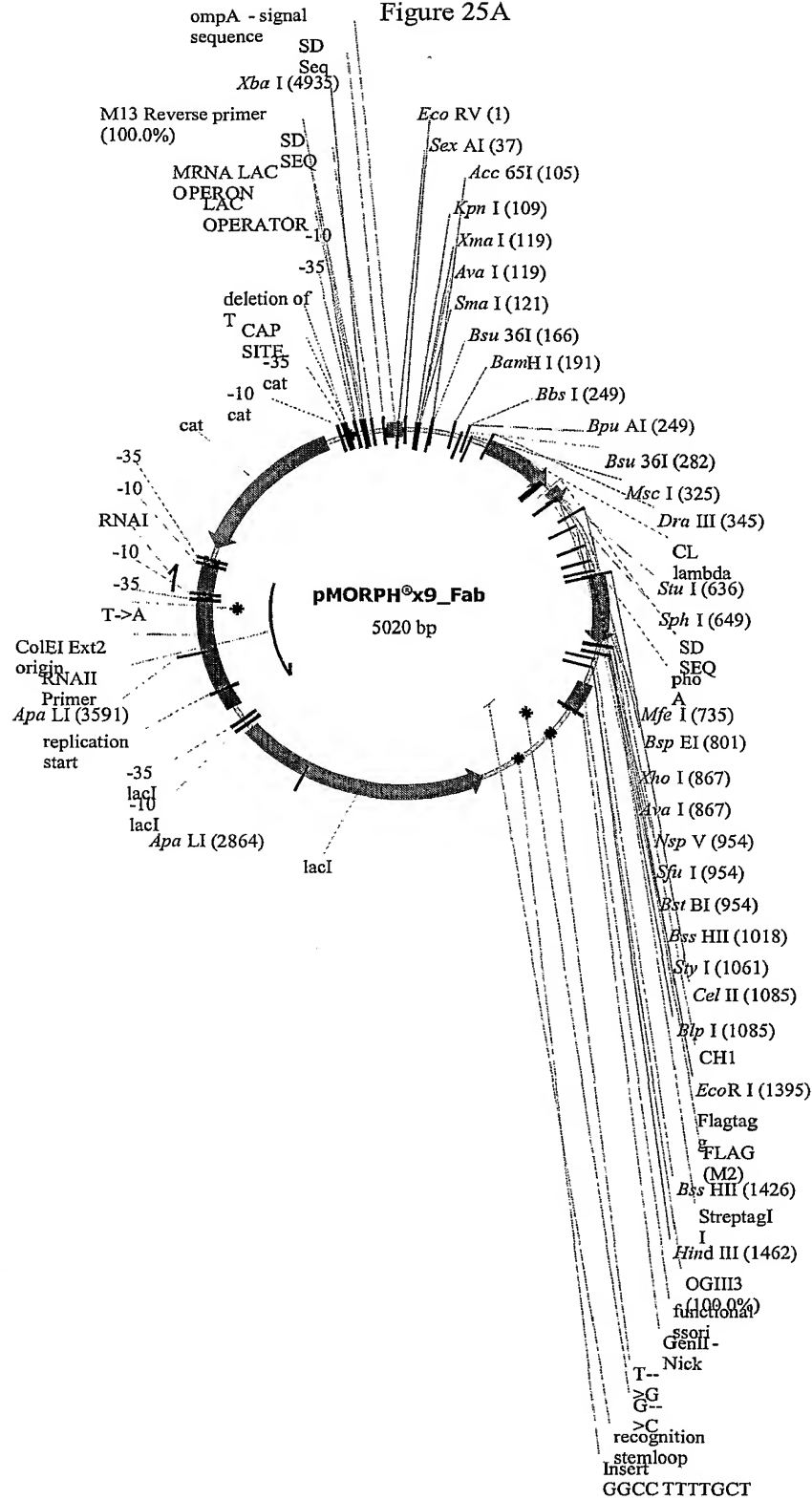


Figure 25B

EcoRV SexAI
 ~~~~~  
 SEQ ID NO: 92 1 ATCGTGCTGA CCCAGCCGCC TTCAGTGAGT GGCGCACCAG GTCAGCGTGT  
 SEQ ID NO: 93 TAGCACGACT GGGTCGGCGG AAGTCACTCA CCGCGTGGTC CAGTCGCACA  
  
 51 GACCATCTCG TGTAGCGGCA GCAGCAGCAA CATTGGCAGC AACTATGTGA  
 CTGGTAGAGC ACATCGCCGT CGTCGTCGTT GTAACCGTCG TTGATACT  
  
 ~~~~~  
 XmaI

 KpnI SmaI
 ~~~~~  
 Acc65I          Aval  
 ~~~~~  
 101 GCTGGTACCA GCAGTTGCCC GGGACGGCGC CGAAACTGCT GATTATATGAT
 CGACCATGGT CGTCAACGGG CCCTGCCGCG GCTTTGACGA CTAAATACTA

 Bsu36I BamHI
 ~~~~~  
 151 AACAAACCAGC GTCCCTCAGG CGTGCCGGAT CGTTTTAGCG GATCCAAAAG  
 TTGTTGGTCG CAGGGAGTCC GCACGGCCTA GCAAATCGC CTAGGTTTTC  
  
 BpuAI  
 ~~~~~  
 BbsI
 ~~~~~  
 201 CGGCACCAGC GCGAGCCTTG CGATTACGGG CCTGCAAAGC GAAGACGAAG  
 GCCGTGGTCG CGCTCGGAAC GCTAATGCCC GGACGTTTCG CTTCTGCTTC  
  
 Bsu36I  
 ~~~~~  
 251 CGGATTATTA TTGCCAGAGC TATGACATGC CTCAGGCTGT GTTTGGCGGC
 GCCTAATAAT AACGGTCTCG ATACTGTACG GAGTCCGACA CAAACCGCCG

 MscI DraIII
 ~~~~~  
 301 GGCACGAAGT TTAACCGTTC TTGGCCAGCC GAAAGCCGCA CCGAGTGTGA  
 CCGTGCTTCA AATTGGCAAG AACCGGTCGG CTTTCGGCGT GGCTCACACT  
  
 351 CGCTGTTTCC GCCGAGCAGC GAAGAATTGC AGGCGAACAA AGCGACCCTG  
 GCGACAAAGG CGGCTCGTCG CTTCTTAACG TCCGCTTGTT TCGCTGGGAC  
  
 401 GTGTGCCTGA TTAGCGACTT TTATCCGGGA GCCGTGACAG TGGCCTGGAA  
 CACACGGA CT AATCGCTGAA AATAGGCCCT CGGCACTGTC ACCGGACCTT  
  
 451 GGCAGATAGC AGCCCCGTCA AGGCGGGAGT GGAGACCACC ACACCCTCCA  
 CCGTCTATCG TCGGGGCAGT TCCGCCCTCA CCTCTGGTGG TGTGGGAGGT  
  
 501 AACAAAGCAA CAACAAGTAC GCGGCCAGCA GCTATCTGAG CCTGACGCCT  
 TTGTTTCGTT GTTGTTCATG CGCCGGTCGT CGATAGACTC GGA CTGCGGA  
  
 551 GAGCAGTGGA AGTCCCACAG AAGCTACAGC TGCCAGGTCA CGCATGAGGG  
 CTCGTCACTT TCAGGGTGTC TTCGATGTCG ACGGTCCAGT GCGTACTCCC  
  
 StuI              SphI  
 ~~~~~  
 601 GAGCACCGTG GAAAAAACCG TTGCGCCGAC TGAGGCCTGA TAAGCATGCG
 CTCGTGGCAC CTTTTTTGGC AACGCGGCTG ACTCCGGA CT ATTCGTACGC

651 TAGGAGAAAA TAAATGAAA CAAAGCACTA TTGCACTGGC ACTCTTACCG
ATCCTCTTTT ATTTTACTTT GTTTCGTGAT AACGTGACCG TGAGAATGGC

MfeI
~~~~~

701 TTGCTCTTCA CCCCTGTTAC CAAAGCCCAG GTGCAATTGA AAGAAAGCGG  
AACGAGAAGT GGGGACAATG GTTTCGGGTC CACGTAACT TTCTTTCGCC

BspEI  
~

751 CCCGGCCCTG GTGAAACCGA CCCAAACCCCT GACCCTGACC TGTACCTTTT  
GGGCCGGGAC CACTTTGGCT GGGTTTGGA CTGGGACTGG ACATGGAAAA

BspEI  
~~~~~

801 CCGGATTTAG CCTGTCCACG TCTGGCGTTG GCGTGGGCTG GATTGCGCAG
GGCCTAAATC GGACAGGTGC AGACCGCAAC CGCACCCGAC CTAAGCGGTC

XhoI
~~~~~

AvaI  
~~~~~

851 CCGCCTGGGA AAGCCCTCGA GTGGCTGGCT CTGATTGATT GGGATGATGA
GGCGGACCCT TTCGGGAGCT CACCGACCGA GACTAACTAA CCCTACTACT

901 TAAGTATTAT AGCACCAGCC TGAAAACGCG TCTGACCATT AGCAAAGATA
ATTCATAATA TCGTGGTCGG ACTTTTGCGC AGACTGGTAA TCGTTTCTAT

BstBI
~~~~~

SfuI  
~~~~~

NspV
~~~~~

951 CTTCGAAAAA TCAGGTGGTG CTGACTATGA CCAACATGGA CCCGGTGGAT  
GAAGCTTTT AGTCCACCAC GACTGATACT GGTTGTACCT GGGCCACCTA

BssHII  
~~~~~

1001 ACGGCCACCT ATTATTGCGC GCGTTCTCCT CGTTATCGTG GTGCTTTTGA
TGCCGGTGGA TAATAACGCG CGCAAGAGGA GCAATAGCAC CACGAAAACCT

BlpI
~~~~~

StyI  
~~~~~

CellI
~~~~~

1051 TTATTGGGGC CAAGGCACCC TGGTGACGGT TAGCTCAGCG TCGACCAAAG  
AATAACCCCG GTTCCGTGGG ACCACTGCCA ATCGAGTCGC AGCTGGTTTC

1101 GTCCAAGCGT GTTTCCGCTG GCTCCGAGCA GCAAAAGCAC CAGCGGCGGC  
CAGGTTTCGA CAAAGGCGAC CGAGGCTCGT CGTTTTCTGT GTCGCCGCCG

1151 ACGGCTGCCC TGGGCTGCCT GGTTAAAGAT TATTTCCCGG AACCAGTCAC  
TGCCGACGGG ACCCGACGGA CCAATTCTA ATAAAGGGCC TTGGTCAGTG

1201 CGTGAGCTGG AACAGCGGGG CGCTGACCAG CGGCGTGCAT ACCTTTCGGG  
GCACTCGACC TTGTCGCCCC GCGACTGGTC GCCGCACGTA TGGAAGGCC

1251 CGGTGCTGCA AAGCAGCGGC CTGTATAGCC TGAGCAGCGT TGTGACCGTG  
GCCACGACGT TTCGTCGCCG GACATATCGG ACTCGTCGCA AACTGGCAC

1301 CCGAGCAGCA GCTTAGGCAC TCAGACCTAT ATTTGCAACG TGAACCATAA  
GGCTCGTCGT CGAATCCGTG AGTCTGGATA TAAACGTTGC ACTTGGTATT

EcoRI

1351 ACCGAGCAAC ACCAAAGTGG ATAAAAAAGT GGAACCGAAA AGCGAATTCTG  
TGGCTCGTTG TGGTTTCACC TATTTTTTCA CCTTGGCTTT TCGCTTAAGC

BssHII

1401 ACTATAAAGA TGACGATGAC AAAGGCGCGC CGTGGAGCCA CCCGCAGTTT  
TGATATTTCT ACTGCTACTG TTTCCGCGCG GCACCTCGGT GGGCGTCAAA

HindIII

1451 GAAAAATGAT AAGCTTGACC TGTGAAAGTGA AAAATGGCGC AGATTGTGCG  
CTTTTTACTA TTCGAACTGG ACACTTCACT TTTTACCGCG TCTAACACGC  
OGIII3 100.0%

1501 ACATTTTTTT TGTCTGCCGT TTAATTAAAG GGGGGGGGGG GCCGGCCTGG  
TGTAACAAAA ACAGACGGCA AATTAATTTT CCCCCCCCCC CGGCCGGACC

1551 GGGGGGGTGT ACATGAAATT GTAAACGTGA ATATTTTGTT AAAATTTCGC  
CCCCCCCCA TGTACTTTAA CATTTGCAAT TATAAACAA TTTAAGCGC

1601 TTAAATTTTT GTTAAATCAG CTCATTTTTT AACCAATAGG CCGAAATCGG  
AATTTAAAA CAATTTAGTC GAGTAAAAAA TTGGTTATCC GGCTTTAGCC

1651 CAAAATCCCT TATAATCAA AAGAATAGAC CGAGATAGGG TTGAGTGTTG  
GTTTTAGGGA ATATTTAGTT TTCTTATCTG GCTCTATCCC AACTCAAAAC

1701 TTCCAGTTTG GAACAAGAGT CCACTATTAA AGAACGTGGA CTCCAACGTC  
AAGGTCAAAC CTTGTTCTCA GGTGATAATT TCTTGACCT GAGGTTGCAG

1751 AAAGGGCGAA AAACCGTCTA TCAGGGCGAT GGCCCACTAC GAGAACCATC  
TTCCCGCTT TTTGGCAGAT AGTCCCGCTA CCGGGTGATG CTCTTGTTAG

1801 ACCCTAATCA AGTTTTTTGG GGTGAGGTG CCGTAAAGCA CTAAATCGGA  
TGGGATTAGT TCAAAAAACC CCAGCTCCAC GGCATTTCTG GATTTAGCCT

1851 ACCCTAAAGG GAGCCCCCGA TTTAGAGCTT GACGGGGAAA GCCGGCGAAC  
TGGGATTTCC CTCGGGGGCT AAATCTCGAA CTGCCCTTT CGGCCGCTTG

1901 GTGGCGAGAA AGGAAGGGAA GAAAGCGAAA GGAGCGGGCG CTAGGGCGCT  
CACCGCTCTT TCCTTCCCTT CTTTCGCTTT CCTCGCCCGC GATCCCGCGA

1951 GGCAAGTGTA GCGGTCACGC TGCGCGTAAC CACCACACCC GCCGCGCTTA  
CCGTTACAT CGCCAGTGCG ACGCGCATTG GTGGTGTGGG CGGCGCGAAT

2001 ATGCGCCGCT ACAGGGCGCG TGCTAGACTA GTGTTTAAAC CGGACCGGGG  
TACGCGGCGA TGTCCCGCGC ACGATCTGAT CACAAATTTG GCCTGGCCCC

2051 GGGGGCTTAA GTGGGCTGCA AAACAAAACG GCCTCCTGTC AGGAAGCCGC  
CCCCGAATT CACCCGACGT TTTGTTTTGC CGGAGGACAG TCCTTCGGCG



2101 TTTTATCGGG TAGCCTCACT GCCCGCTTTC CAGTCGGGAA ACCTGTCTGT  
AAAATAGCCC ATCGGAGTGA CGGGCGAAAG GTCAGCCCTT TGGACAGCAC

2151 CCAGCTGCAT CAGTGAATCG GCCAACGCGC GGGGAGAGGC GGTTCGCGTA  
GGTCGACGTA GTCAGTAGC CGGTTGCGCG CCCCTCTCCG CCAAACGCAT

2201 TTGGGAGCCA GGGTGGTTTT TCTTTTCACC AGTGAGACGG GCAACAGCTG  
AACCCTCGGT CCCACCAAAA AGAAAAGTGG TCACTCTGCC CGTTGTGCGAC

2251 ATTGCCCTTC ACCGCCTGGC CCTGAGAGAG TTGCAGCAAG CGGTCCACGC  
TAACGGGAAG TGGCGGACCG GGACTCTCTC AACGTCGTTC GCCAGGTGCG

2301 TGGTTTGCCC CAGCAGGCGA AAATCCTGTT TGATGGTGGT CAGCGGCGGG  
ACCAAACGGG GTCGTCCGCT TTTAGGACAA ACTACCACCA GTCGCCGCC

2351 ATATAACATG AGCTGTCCTC GGTATCGTCG TATCCCACTA CCGAGATGTC  
TATATTGTAC TCGACAGGAG CCATAGCAGC ATAGGGTGAT GGCTCTACAG

2401 CGCACCAACG CGCAGCCCGG ACTCGGTAAT GGCACGCATT GCGCCACGG  
GCGTGGTTGC GCGTCGGGCC TGAGCCATTA CCGTGCGTAA CGCGGGTCGC

2451 CCATCTGATC GTTGGCAACC AGCATCGCAG TGGGAACGAT GCCCTCATTC  
GGTAGACTAG CAACCGTTGG TCGTAGCGTC ACCCTTGCTA CGGGAGTAAG

2501 AGCATTTGCA TGGTTTGTG AAAACCGGAC ATGGCACTCC AGTCGCCTTC  
TCGTAAACGT ACCAAACAAC TTTTGGCCTG TACCGTGAGG TCAGCGGAAG

2551 CCGTTCGCT ATCGGCTGAA TTTGATTGCG AGTGAGATAT TTATGCCAGC  
GGCAAGGCGA TAGCCGACTT AACTAACGC TCACTCTATA AATACGGTCG

2601 CAGCCAGACG CAGACGCGCC GAGACAGAAC TTAATGGGCC AGCTAACAGC  
GTCGGTCTGC GTCTGCGCGG CTCTGTCTTG AATTACCCCG TCGATTGTCG

2651 GCGATTTGCT GGTGGCCCAA TGCAGCCAGA TGCTCCACGC CCAGTCGCGT  
CGCTAAACGA CCACCGGGT ACGCTGGTCT ACGAGGTGCG GGTCAGCGCA

2701 ACCGTCTCA TGGGAGAAAA TAATACTGTT GATGGGTGTC TGGTCAGAGA  
TGGCAGGAGT ACCCTCTTTT ATTATGACAA CTACCCACAG ACCAGTCTCT

2751 CATCAAGAAA TAACGCCGGA ACATTAGTGC AGGCAGCTTC CACAGCAATA  
GTAGTTCTTT ATTGCGGCCT TGTAATCACG TCCGTCGAAG GTGTCGTTAT

2801 GCATCCTGGT CATCCAGCGG ATAGTTAATA ATCAGCCAC TGACACGTTG  
CGTAGGACCA GTAGGTCGCC TATCAATTAT TAGTCGGGTG ACTGTGCAAC

ApaLI

2851 CGCGAGAAGA TTGTGCACCG CCGCTTTACA GGCTTCGACG CCGCTTCGTT  
GCGCTCTTCT AACACGTGGC GCGGAAATGT CCGAAGCTGC GGCGAAGCAA

2901 CTACCATCGA CACGACCACG CTGGCACCCA GTTGATCGGC GCGAGATTTA  
GATGGTAGCT GTGCTGGTGC GACCGTGGGT CAACTAGCCG CGCTCTAAAT

2951 ATCGCCGCGA CAATTTGCGA CGGCGCGTGC AGGGCCAGAC TGGAGGTGGC  
TAGCGGCGCT GTTAAACGCT GCCGCGCACG TCCCGGTCTG ACCTCCACCG

3001 AACGCCAATC AGCAACGACT GTTTGCCCGC CAGTTGTTGT GCCACGCGGT  
TTGCGGTTAG TCGTTGCTGA CAAACGGGCG GTCAACAACA CGGTGCGCCA

3051 TAGGAATGTA ATTCAGCTCC GCCATCGCCG CTTCCAATTT TTCCCGCGTT  
ATCCTTACAT TAAGTCGAGG CGGTAGCGGC GAAGGTGAAA AAGGGCGCAA

3101 TTCGCAGAAA CGTGGCTGGC CTGGTTCACC ACGCGGGAAA CGGTCTGATA  
AAGCGTCTTT GCACCGACCG GACCAAGTGG TGCGCCCTTT GCCAGACTAT

3151 AGAGACACCG GCATACTCTG CGACATCGTA TAACGTTACT GGTTTCACAT  
TCTCTGTGGC CGTATGAGAC GCTGTAGCAT ATTGCAATGA CCAAAGTGTA

3201 TCACCACCCT GAATTGACTC TCTTCCGGGC GCTATCATGC CATACCGCGA  
AGTGGTGGGA CTTAACTGAG AGAAGGCCCG CGATAGTACG GTATGGCGCT

3251 AAGGTTTTGC GCCATTCGAT GCTAGCCATG TGAGCAAAAG GCCAGCAAAA  
TTCCAAAACG CGGTAAGCTA CGATCGGTAC ACTCGTTTTC CGGTCGTTTT

3301 GGCCAGGAAC CGTAAAAAGG CCGCGTTGCT GCGTTTTTTC CATAGGCTCC  
CCGGTCCTTG GCATTTTTCC GCGCAACGA CCGCAAAAAG GTATCCGAGG

3351 GCCCCCTGA CGAGCATCAC AAAAATCGAC GCTCAAGTCA GAGGTGGCGA  
CGGGGGGACT GCTCGTAGTG TTTTLAGCTG CGAGTTCAGT CTCCACCGCT

3401 AACCCGACAG GACTATAAAG ATACCAGGCG TTTCCCCTG GAAGCTCCCT  
TTGGGCTGTC CTGATATTTC TATGGTCCGC AAAGGGGGAC CTTCGAGGGA

3451 CGTGCGCTCT CCTGTTCCGA CCCTGCCGCT TACCGGATAC CTGTCCGCCT  
GCACGCGAGA GGACAAGGCT GGGACGGCGA ATGGCCTATG GACAGGCGGA

3501 TTCTCCCTTC GGAAGCGTG GCGCTTTCTC ATAGCTCACG CTGTAGGTAT  
AAGAGGGAAG CCCTTCGCAC CGCGAAAGAG TATCGAGTGC GACATCCATA

## ApaLI

3551 CTCAGTTCGG TGTAGGTCGT TCGCTCCAAG CTGGGCTGTG TGCACGAACC  
GAGTCAAGCC ACATCCAGCA AGCGAGGTTT GACCCGACAC ACGTGCTTGG

3601 CCCCCTTCAG CCCGACCGCT GCGCCTTATC CGGTAAGTAT CGTCTTGAGT  
GGGGCAAGTC GGGCTGGCGA CGCGGAATAG GCCATTGATA GCAGAACTCA

3651 CCAACCCGGT AAGACACGAC TTATCGCCAC TGGCAGCAGC CACTGGTAAC  
GGTTGGGCCA TTCTGTGCTG AATAGCGGTG ACCGTCGTCG GTGACCATTG

3701 AGGATTAGCA GAGCGAGGTA TGTAGGCGGT GCTACAGAGT TCTTGAAGTG  
TCCTAATCGT CTCGCTCCAT ACATCCGCCA CGATGTCTCA AGAACTTCAC

3751 GTGGCCTAAC TACGGCTACA CTAGAAGAAG AGTATTTGGT ATCTGCGCTC  
CACCGGATTG ATGCCGATGT GATCTTCTTG TCATAAACCA TAGACGCGAG

3801 TGCTGTAGCC AGTTACCTTC GGAAAAAGAG TTGGTAGCTC TTGATCCGGC  
ACGACATCGG TCAATGGAAG CCTTTTTCTC AACCATCGAG AACTAGGCCG

3851 AAACAAACCA CCGCTGGTAG CGGTGGTTTT TTTGTTTGCA AGCAGCAGAT  
TTTGTTTGGT GCGGACCATC GCCACCAAAA AAACAAACGT TCGTCGTCTA

3901 TACGCGCAGA AAAAAAGGAT CTCAAGAAGA TCCTTTGATC TTTTCTACGG  
ATGCGCGTCT TTTTTCCTA GAGTTCCTCT AGGAACTAG AAAAGATGCC

3951 GGTCTGACGC TCAGTGGAAC GAAAACTCAC GTTAAGGGAT TTTGGTCAGA  
CCAGACTGCG AGTCACCTTG CTTTTGAGTG CAATTCCTA AAACCACTCT

4001 TCTAGCACCA GCGTTTAAG GGCACCAATA ACTGCCTTAA AAAAATTACG  
AGATCGTGGT CCGCAAATTC CCGTGTTAT TGACGGAATT TTTTAAATGC

4051 CCCCCGCCCTG CCACTCATCG CAGTACTGTT GTAATTCATT AAGCATTCTG  
GGGGCGGGAC GGTGAGTAGC GTCATGACAA CATTAAAGTAA TTCGTAAGAC

4101 CCGACATGGA AGCCATCACA AACGGCATGA TGAACCTGAA TCGCCAGCGG  
GGCTGTACCT TCGGTAGTGT TTGCCGTACT ACTTGGACTT AGCGGTCGCC

4151 CATCAGCACC TTGTCGCCCTT GCGTATAATA TTTGCCCATTA GTGAAAACGG  
GTAGTCGTGG AACAGCGGAA CGCATATTAT AAACGGGTAT CACTTTTGCC

4201 GGGCGAAGAA GTTGTCCATA TTGGCTACGT TTAAATCAAA ACTGGTGAAA  
CCCCTTCTT CAACAGGTAT AACCGATGCA AATTTAGTTT TGACCACTTT

4251 CTCACCCAGG GATTGGCTGA GACGAAAAAC ATATTCTCAA TAAACCCCTT  
GAGTGGGTCC CTAACCGACT CTGCTTTTTG TATAAGAGTT ATTTGGGAAA

4301 AGGGAAATAG GCCAGGTTTT CACCGTAACA CGCCACATCT TCGCAATATA  
TCCCTTTATC CGGTCAAAA GTGGCATTGT GCGGTGTAGA ACGTTATAT

4351 TGTGTAGAAA CTGCCGAAA TCGTCGTGGT ATTCACTCCA GAGCGATGAA  
ACACATCTTT GACGGCCTTT AGCAGCACCA TAAGTGAGGT CTCGCTACTT

4401 AACGTTTCAG TTTGCTCATG GAAAACGGTG TAACAAGGGT GAACACTATC  
TTGCAAAGTC AAACGAGTAC CTTTTGCCAC ATTGTTCCCA CTTGTGATAG

4451 CCATATCACC AGCTCACCGT CTTTCATTGC CATACGGAAC TCCGGGTGAG  
GGTATAGTGG TCGAGTGGCA GAAAGTAACG GTATGCCTTG AGGCCCACTC

4501 CATTCATCAG GCGGGCAAGA ATGTGAATAA AGGCCGGATA AAACCTTGTC  
GTAAGTAGTC CGCCGTTCT TACACTTATT TCCGGCCTAT TTTGAACACG

4551 TTATTTTTCT TTACGGTCTT TAAAAAGGCC GTAATATCCA GCTGAACGGT  
AATAAAAAGA AATGCCAGAA ATTTTCCGG CATTATAGGT CGACTTGCCA

4601 CTGGTTATAG GTACATTGAG CAACTGACTG AAATGCCTCA AAATGTTCTT  
GACCAATATC CATGTAACTC GTTGACTGAC TTTACGGAGT TTTACAAGAA

4651 TACGATGCCA TTGGGATATA TCAACGGTGG TATATCCAGT GATTTTTTTC  
ATGCTACGGT AACCTTATAT AGTTGCCACC ATATAGGTCA CTAAAAAAG

4701 TCCATTTTAG CTTCCCTTAGC TCCTGAAAAT CTCGATAACT CAAAAAATAC  
AGGTAAAATC GAAGGAATCG AGGACTTTTA GAGCTATTGA GTTTTTTATG

4751 GCGCGGTAGT GATCTTATTT CATTATGGTG AAAGTTGGAA CCTCACCCGA  
CGGGCCATCA CTAGAATAAA GTAATACCAC TTTCAACCTT GGAGTGGGCT

4801 CGTCTAATGT GAGTTAGCTC ACTCATTAGG CACCCAGGC TTTACACTTT  
GCAGATTACA CTCAATCGAG TGAGTAATCC GTGGGGTCCG AAATGTGAAA

4851 ATGCTTCCGG CTCGTATGTT GTGTGGAATT GTGAGCGGAT AACAATTTCA  
TACGAAGGCC GAGCATACAA CACACCTTAA CACTCGCCTA TTGTTAAAGT

M13 Reverse primer 100.0% XbaI

4901 CACAGGAAAC AGCTATGACC ATGATTACGA ATTTCTAGAT AACGAGGGCA  
GTGTCCTTTG TCGATACTGG TACTAATGCT TAAAGATCTA TTGCTCCCGT

4951 AAAAATGAAA AAGACAGCTA TCGCGATTGC AGTGGCACTG GCTGGTTTCG  
TTTTTACTTT TTCTGTCGAT AGCGCTAACG TCACCGTGAC CGACCAAAGC

EcoRV

~~~~~  
5001 CTACCGTAGC GCAGGCCGAT
GATGGCATCG CGTCCGGCTA

Figure 26A

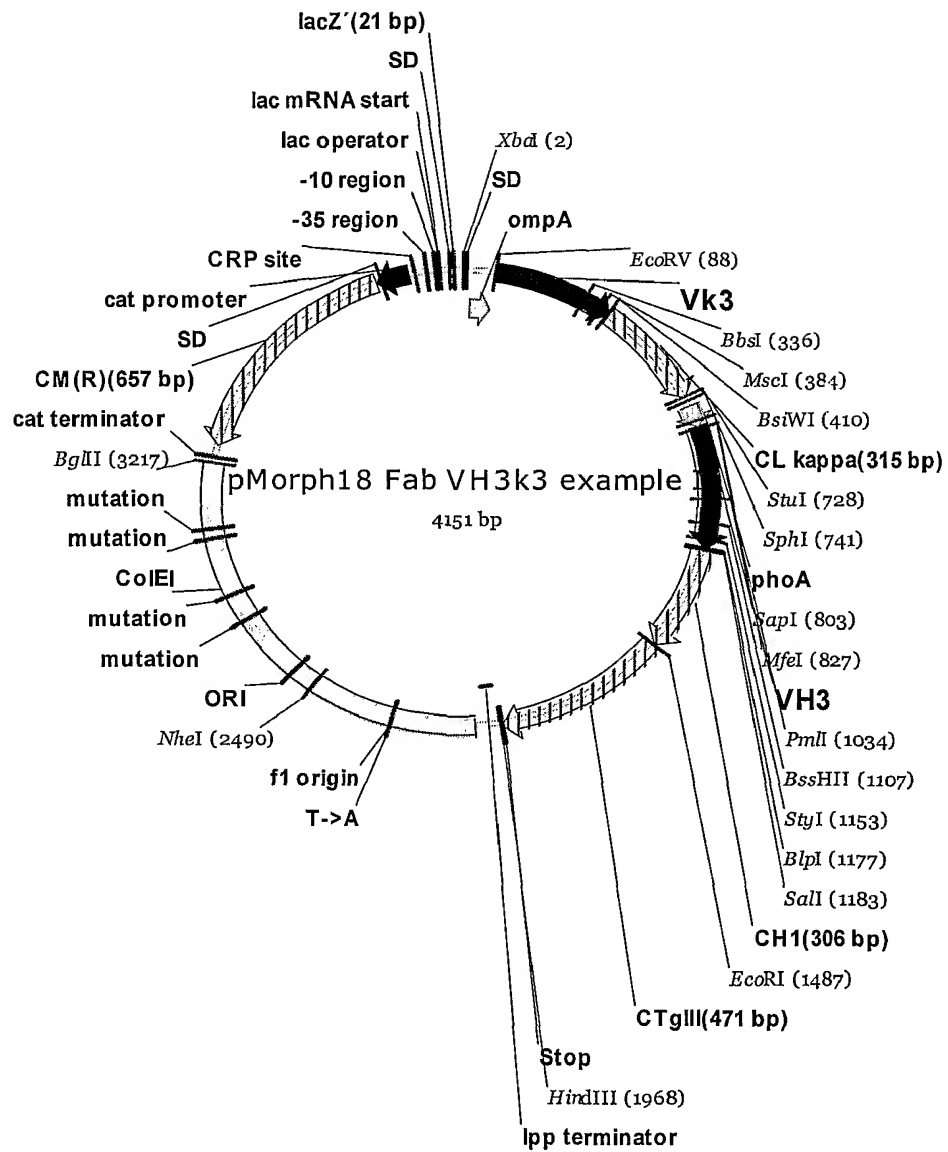


Figure 26B

lacZ' SD ompA
 ~~~~~  
 XbaI  
 ~~~~~  
 M K K T A I A I A V ·
 SEQ ID NO: 94 1 TCTAGATAAC GAGGGCAAAA AATGAAAAAG ACAGCTATCG
 CGATTGCAGT
 SEQ ID NO: 95 AGATCTATTG CTCCCGTTTT TTA CTTTTC TGTCGATAGC GCTAACGTCA
 Vk3
 ~~~~~  
 ompA  
 ~~~~~  
 EcoRV
 ~~~~~  
 SEQ ID NO: 96 · A L A G F A T V A Q A D I V L T Q ·  
 51 GGCAC TGGCT GGTTCGCTA CCGTAGCGCA GGCCGATATC GTGCTGACCC  
 CCGTGACCGA CCAAAGCGAT GGCATCGCGT CCGGCTATAG CACGACTGGG  
 Vk3  
 ~~~~~  
 101 · S P A T L S L S P G E R A T L S
 AGAGCCCGGC GACCCTGAGC CTGTCTCCGG GCGAACGTGC GACCCTGAGC
 TCTCGGGCCG CTGGGACTCG GACAGAGGCC CGCTTGACACG CTGGGACTCG
 Vk3
 ~~~~~  
 151 C R A S Q S V S S S Y L A W Y Q Q ·  
 TGCAGAGCGA GCCAGAGCGT GAGCAGCAGC TATCTGGCGT GGTACCAGCA  
 ACGTCTCGCT CGGTCTCGCA CTCGTCGTCG ATAGACCGCA CCATGGTCGT  
 Vk3  
 ~~~~~  
 201 · K P G Q A P R L L I Y G A S S R A ·
 GAAACCAGGT CAAGCACCGC GTCTATTAAT TTATGGCGCG AGCAGCCGTG
 CTTTGGTCCA GTTCGTGGCG CAGATAATTA AATACCGCGC TCGTCGGCAC
 Vk3
 ~~~~~  
 251 · T G V P A R F S G S G S G T D F  
 CAACTGGGGT CCCGGCGCGT TTAGCGGGCT CTGGATCCGG CACGGATTTT  
 GTTGACCCCA GGGCCGCGCA AAATCGCCGA GACCTAGGCC GTGCCTAAAA  
 Vk3  
 ~~~~~  
 BbsI
 ~~~~~  
 301 T L T I S S L E P E D F A V Y Y C ·  
 ACCCTGACCA TTAGCAGCCT GGAACCTGAA GACTTTGCGG TGTATTATTG  
 TGGGACTGGT AATCGTCGGA CCTTGGACTT CTGAAACGCC ACATAATAAC  
 Vk3  
 ~~~~~  
 MscI
 ~~~~~  
 351 · Q Q H Y T T P P T F G Q G T K V E ·  
 CCAGCAGCAT TATACCACCC CGCCGACCTT TGGCCAGGGT ACGAAAGTTG  
 GGTCGTCGTA ATATGGTGGG GCGGCTGGAA ACCGGTCCCA TGCTTTCAAC  
 CL kappa  
 ~~~~~  
 Vk3
 ~~~~~  
 BsiWI  
 ~~~~~  
 401 · I K R T V A A P S V F I F P P S
 AAATTAAACG TACGGTGGCT GCTCCGAGCG TGTTTATTTT TCCGCCGAGC
 TTTAATTTGC ATGCCACCGA CGAGGCTCGC ACAAATAAAA AGGCGGCTCG

CL kappa

451 DEQLKSGTASVVCLLNN·
GATGAACAAC TGAAAAGCGG CACGGCGAGC GTGGTGTGCC TGCTGAACAA
CTACTTGTG ACTTTTCGCC GTGCCGCTCG CACCACACGG ACGACTTGT

CL kappa

501 ·FYPREAKVQWKVDNALQ·
CTTTTATCCG CGTGAAGCGA AAGTTCAGTG GAAAGTAGAC AACGCGCTGC
GAAAATAGGC GCACTTCGCT TTCAAGTCAC CTTTCATCTG TTGCGCGACG

CL kappa

551 ·SGNSQESVTEQDSKDS
AAAGCGGCAA CAGCCAGGAA AGCGTGACCG AACAGGATAG CAAAGATAGC
TTTCGCCGTT GTCGGTCCTT TCGCACTGGC TTGTCCTATC GTTCTATCG

CL kappa

601 TYSLSSTLTLSKADYEK·
ACCTATTCTC TGAGCAGCAC CCTGACCCTG AGCAAAGCGG ATTATGAAAA
TGGATAAGAG ACTCGTCGTG GGACTGGGAC TCGTTTCGCC TAATACTTTT

CL kappa

651 ·HKVYACEVTHQGLSSPV·
ACATAAAGTG TATGCGTGCG AAGTGACCCA TCAAGGTCTG AGCAGCCCGG
TGTATTTAC ATACGCACGC TTCCTGGGT AGTTCCAGAC TCGTCGGGCC

CL kappa

701 StuI SphI
·TKSFNRGEA
TGACTAAATC TTTTAATCGT GCGAGGCCT GATAAGCATG CGTAGGAGAA
ACTGATTTAG AAAATTAGCA CCGCTCCGGA CTATTCGTAC GCATCCTCTT

phoA

751 SapI
MKQSTIALALLPLLF·
AATAAAATGA AACAAAGCAC TATTGCACTG GCACTCTTAC CGTTGCTCTT
TTATTTTACT TTGTTTCGTG ATAACGTGAC CGTGAGAATG GCAACGAGAA

VH3

phoA

801 SapI MfeI
·TPVTKAQVQLVESGGGL·
CACCCCTGTT ACCAAAGCCG AAGTGCAATT GGTGGAAAGC GGCGGCGGCC
GTGGGGACAA TGGTTTCGGC TTCACGTAA CCACCTTTCG CCGCCGCCGG

VH3

851 ·VQPGGSLRLSCAASGF
TGGTGCAACC GGGCGGCAGC CTGCGTCTGA GCTGCGCGGC CTCCGGATTT
ACCACGTTGG CCCGCCGTCG GACGCAGACT CGACGCGCCG GAGGCCTAAA

VH3

901 TFSSYAMSWVRQAPGKG·
ACCTTTAGCA GCTATGCGAT GAGCTGGGTG CGCCAAGCCC CTGGGAAGGG
TGGAATCGT CGATACGCTA CTCGACCCAC GCGGTTCTGGG GACCCTTCCC

VH3

951 ·LEWVS AISGS GGS TYA·
TCTCGAGTGG GTGAGCGCGA TTAGCGGTAG CGGCGGCAGC ACCTATTATG

AGAGCTCACC CACTCGCGCT AATCGCCATC GCCGCCGTCG TGGATAATAC
 VH3

PmlI

1001 · D S V K G R F T I S R D N S K N
 CGGATAGCGT GAAAGGCCGT TTTACCATTT CACGTGATAA TTCGAAAAAC
 GCCTATCGCA CTTTCCGGCA AAATGGTAAA GTGCACTATT AAGCTTTTTG
 VH3

1051 T L Y L Q M N S L R A E D T A V Y ·
 ACCCTGTATC TGCAAATGAA CAGCCTGCGT GCGGAAGATA CGGCCGTGTA
 TGGGACATAG ACGTTTACTT GTCGGACGCA CGCCTTCTAT GCCGGCACAT
 VH3

BssHII

1101 · Y C A R W G G D G F Y A M D Y W G ·
 TTATTGCGCG CGTTGGGGCG GCGATGGCTT TTATGCGATG GATTATTGGG
 AATAACGCGC GCAACCCCGC CGCTACCGAA AATACGCTAC CTAATAACCC
 CH1

VH3

Sall

StyI BlnI

1151 · Q G T L V T V S S A S T K G P S
 GCCAAGGCAC CCTGGTGACG GTTAGCTCAG CGTCGACCAA AGGTCCAAGC
 CGGTTCCGTG GGACCACTGC CAATCGAGTC GCAGCTGGTT TCCAGGTTCC
 CH1

1201 V F P L A P S S K S T S G G T A A ·
 GTGTTTCCGC TGGCTCCGAG CAGCAAAAGC ACCAGCGGCG GCACGGCTGC
 CACAAAGGCG ACCGAGGCTC GTCGTTTTCG TGGTCGCCGC CGTGCCGACG
 CH1

1251 · L G C L V K D Y F P E P V T V S W ·
 CCTGGGCTGC CTGGTTAAAG ATTATTTCCC GGAACCAGTC ACCGTGAGCT
 GGACCCGACG GACCAATTTC TAATAAAGGG CCTTGGTCAG TGGCACTCGA
 CH1

1301 · N S G A L T S G V H T F P A V L
 GGAACAGCGG GGCGCTGACC AGCGGCGTGC ATACCTTTCC GGCGGTGCTG
 CCTTGTCGCC CCGCGACTGG TCGCCGCACG TATGGAAAGG CCGCCACGAC
 CH1

1351 Q S S G L Y S L S S V V T V P S S ·
 CAAAGCAGCG GCCTGTATAG CCTGAGCAGC GTTGTGACCG TGCCGAGCAG
 GTTTCGTCGC CGGACATATC GGAATCGTCG CAACACTGGC ACGGCTCGTC
 CH1

1401 · S L G T Q T Y I C N V N H K P S N ·
 CAGCTTAGGC ACTCAGACCT ATATTTGCAA CGTGAACCAT AAACCGAGCA
 GTCGAATCCG TGAGTCTGGA TATAAACGTT GCACTTGGTA TTGGGCTCGT
 CH1 CTgIII

EcoRI

1451 · T K V D K K V E P K S E F G G G
 ACACCAAAGT GGATAAAAAA GTGGAACCGA AAAGCGAATT CGGGGGAGGG

TGTGGTTTCA CCTATTTTTT CACCTTGGCT TTTCGCTTAA GCCCCCTCCC
 CTgIII
 ~~~~~  
 S G S G D F D Y E K M A N A N K G ·  
 1501 AGCGGGAGCG GTGATTTTGA TTATGAAAAG ATGGCAAACG CTAATAAGGG  
 TCGCCCTCGC CACTAAAAC TAACTTTTC TACCGTTTGC GATTATTCCC  
 CTgIII  
 ~~~~~  
 · A M T E N A D E N A L Q S D A K G ·
 1551 GGCTATGACC GAAAATGCCG ATGAAAACGC GCTACAGTCT GACGCTAAAG
 CCGATACTGG CTTTACGGC TACTTTTGC GATGTCAGA CTGCGATTTC
 CTgIII
 ~~~~~  
 · K L D S V A T D Y G A A I D G F  
 1601 GCAAACCTTGA TTCTGTCGCT ACTGATTACG GTGCTGCTAT CGATGGTTTC  
 CGTTTGAAC TAAACAGCGA TGAATAATGC CACGACGATA GCTACCAAAG  
 CTgIII  
 ~~~~~  
 I G D V S G L A N G N G A T G D F ·
 1651 ATTGGTGACG TTTCCGGCCT TGCTAATGGT AATGGTGCTA CTGGTGATTT
 TAACCACTGC AAAGGCCGGA ACGATTACCA TTACCACGAT GACCACTAAA
 CTgIII
 ~~~~~  
 · A G S N S Q M A Q V G D G D N S P ·  
 1701 TGCTGGCTCT AATTCCCAA TGGCTCAAGT CGGTGACGGT GATAATTCAC  
 ACGACCGAGA TTAAGGGTTT ACCGAGTTCA GCCACTGCCA CTATTAAGTG  
 CTgIII  
 ~~~~~  
 · L M N N F R Q Y L P S L P Q S V
 1751 CTTAATGAA TAATTTCCGT CAATATTTAC CTTCCCTCCC TCAATCGGTT
 GAAATTACTT ATTAAGGCA GTTATAAATG GAAGGGAGGG AGTTAGCCAA
 CTgIII
 ~~~~~  
 E C R P F V F G A G K P Y E F S I ·  
 1801 GAATGTCGCC CTTTGTCTT TGGCGCTGGT AAACCATATG AATTTTCTAT  
 CTTACAGCGG GAAAACAGAA ACCGCGACCA TTTGGTATAC TAAAAGATA  
 CTgIII  
 ~~~~~  
 · D C D K I N L F R G V F A F L L Y ·
 1851 TGATTGTGAC AAAATAAACT TATTCGTGG TGTCTTTGCG TTTCTTTTAT
 ACTAACACTG TTTTATTTGA ATAAGGCACC ACAGAAACGC AAAGAAAATA
 CTgIII
 ~~~~~  
 · V A T F M Y V F S T F A N I L R  
 1901 ATGTTGCCAC CTTTATGTAT GTATTTTCTA CGTTTGCTAA CATACTGCGT  
 TACAACGGTG GAAATACATA CATAAAAGAT GCAAACGATT GTATGACGCA  
 CTgIII  
 ~~~~~  
 Stop lpp terminator
 ~~~~~  
 HindIII  
 ~~~~~  
 N K E S
 1951 AATAAGGAGT CTTGATAAGC TTGACCTGTG AAGTGAAAAA TGGCGCAGAT
 TTATTCCTCA GAACATTCG AACTGGACAC TTCACTTTTT ACCGCGTCTA
 lpp terminator
 ~~~~~  
 2001 TGTGCGACAT TTTTTTTGTC TGCCGTTTAA TGAAATTGTA AACGTTAATA  
 ACACGCTGTA AAAAAACAG ACGGCAAATT ACTTTAACAT TTGCAATTAT  
 ~~~~~  
 fl origin
 2051 TTTTGTAAAA ATTGCGGTTA AATTTTGTGTT AAATCAGCTC ATTTTTTAAC

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AAAACAATTT TAAGCGCAAT TTA AAAACAA TTTAGTCGAG TAAAAAATTG
~~~~~
fl origin
2101 CAATAGGCCG AAATCGGCAA AATCCCTTAT AAATCAAAAG AATAGACCGA
GTTATCCGGC TTTAGCCGTT TTAGGGAATA TTTAGTTTTC TTATCTGGCT
~~~~~
fl origin
2151 GATAGGGTTG AGTGTGTTC CAGTTTGGA CAAGAGTCCA CTATTAAAGA
CTATCCCAAC TCACAACAAG GTCAAACCTT GTTCTCAGGT GATAATTTCT
~~~~~
fl origin
2201 ACGTGGACTC CAACGTCAA GGGCGAAAA CCGTCTATCA GGGCGATGGC
TGCACCTGAG GTTGCAGTTT CCCGCTTTT GGCAGATAGT CCCGCTACCG
~~~~~
fl origin
T->A
~
2251 CCACTACGAG AACCATCACC CTAATCAAGT TTTTGGGGT CGAGGTGCCG
GGTGATGCTC TTGGTAGTGG GATTAGTTCA AAAAACCCCA GCTCCACGGC
~~~~~
fl origin
2301 TAAAGCACTA AATCGGAACC CTAAAGGGAG CCCCGATTT AGAGCTTGAC
ATTCGTGAT TTAGCCTTGG GATTCCCTC GGGGGCTAAA TCTCGAACTG
~~~~~
fl origin
2351 GGGGAAAGCC GCGAACGTG GCGAGAAAGG AAGGGAAGAA AGCGAAAGGA
CCCTTTCGG CCGCTTGAC CGCTTTTCC TTCCCTTCT TCGCTTTCCT
~~~~~
fl origin
2401 GCGGGCGCTA GGGCGCTGGC AAGTGTAGCG GTCACGCTGC GCGTAACCAC
CGCCGCGAT CCCGCGACCG TTCACATCGC CAGTGCGACG CGCATTGGTG
~~~~~
fl origin
NheI
~
2451 CACACCCGCC GCGCTTAATG CGCCGCTACA GGGCGCGTGC TAGCCATGTG
GTGTGGGCGG CGCGAATTAC GCGGCGATGT CCCGCGCACG ATCGGTACAC
~~~~~
fl origin ColEI
2501 AGCAAAAGGC CAGCAAAAGG CCAGGAACCG TAAAAAGGCC GCGTTGCTGG
TCGTTTTCCG GTCGTTTTCC GGTCTTGGC ATTTTCCCG CGCAACGACC
~~~~~
ColEI
ORI
~
2551 CGTTTTTCCA TAGGCTCCGC CCCCTGACG AGCATCACAA AAATCGACGC
GCAAAAAGGT ATCCGAGGCG GGGGACTGC TCGTAGTGTT TTTAGCTGCG
~~~~~
ColEI
2601 TCAAGTCAGA GGTGGCGAAA CCCGACAGGA CTATAAAGAT ACCAGGCGTT
AGTTCAGTCT CCACCGCTT GGGCTGTCCT GATATTTCTA TGGTCCGCAA
~~~~~
ColEI
2651 TCCCCCTGGA AGCTCCCTCG TCGCTCTCC TGTTCGACC CTGCCGCTTA
AGGGGACCT TCGAGGGAGC ACGCGAGAGG ACAAGGCTGG GACGGCGAAT
~~~~~
ColEI
mutation
~
2701 CCGGATACCT GTCCGCCTT CTCCCTTCGG GAAGCGTGGC GCTTTCTCAT
GGCCTATGGA CAGGCGGAAA GAGGGAAGCC CTTGCGACCG CGAAAGAGTA
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ColEI
mutation
~
2751 AGCTCACGCT GTAGGTATCT CAGTTCGGTG TAGGTCGTTT GCTCCAAGCT
TCGAGTGC GA CATCCATAGA GTCAAGCCAC ATCCAGCAAG CGAGGTTT CGA
~~~~~
ColEI
mutation
~
2801 GGGCTGTGTG CACGAACCCC CCGTTCAGTC CGACCGCTGC GCCTTATCCG
CCCACACAC GTGCTTGGGG GGCAAGTCAG GCTGGCGACG CGGAATAGGC
~~~~~
ColEI
2851 GTAAGTATCG TCTTGAGTCC AACCCGGTAA GACACGACTT ATCGCCACTG
CATTGATAGC AGAAGTCAGG TTGGGCCATT CTGTGCTGAA TAGCGGTGAC
~~~~~
ColEI
2901 GCAGCAGCCA CTGGTAACAG GATTAGCAGA GCGAGGTATG TAGGCGGTGC
CGTCGTGGT GACCATTTGC CTAATCGTCT CGCTCCATAC ATCCGCCACG
~~~~~
ColEI
mutation
~
2951 TACAGAGTTC TTGAAGTGGT GGCCTAACTA CGGCTACACT AGAAGAACAG
ATGTCTCAAG AACTTCACCA CCGGATTGAT GCCGATGTGA TCTTCTTGTC
~~~~~
ColEI
mutation
~
3001 TATTTGGTAT CTGCGCTCTG CTGTAGCCAG TTACCTTCGG AAAAAGAGTT
ATAAACATA GACGCGAGAC GACATCGGTC AATGGAAGCC TTTTCTCAA
~~~~~
ColEI
3051 GGTAGCTCTT GATCCGGCAA ACAAACCACC GCTGGTAGCG GTGGTTTTTT
CCATCGAGAA CTAGGCCGTT TGTGTTGGTG CGACCATCGC CACCAAAAAA
~~~~~
ColEI
3101 TGTTTGCAAG CAGCAGATTA CGCGCAGAAA AAAAGGATCT CAAGAAGATC
ACAAACGTTT GTCGTCTAAT GCGCGTCTTT TTTTCCTAGA GTTCTTCTAG
~~~~~
ColEI
3151 CTTTGATCTT TTCTACGGGG TCTGACGCTC AGTGGAACGA AAAGTCACGT
GAAAGTAGAA AAGATGCCCC AGACTGCGAG TCACCTTGCT TTTGAGTGCA
~~~~~
ColEI
cat terminator
~~~~~
BglII
~~~~~
3201 TAAGGGATTT TGGTCAGATC TAGCACCAGG CGTTTAAGGG CACCAATAAC
ATCCCTAAA ACCAGTCTAG ATCGTGGTCC GCAAAATCCC GTGGTTATTG
~~~~~
ColEI
cat terminator
~~~~~
3251 TGCCTTAAAA AAATTACGCC CCGCCCTGCC ACTCATCGCA GTACTGTTGT
ACGGAATTTT TTTAATGCGG GCGGGGACGG TGAGTAGCGT CATGACAACA
~~~~~
CM(R)
3301 AATTCATTAA GCATTCTGCC GACATGGAAG CCATCACAAA CGGCATGATG
TTAAGTAATT CGTAAGACGG CTGTACCTTC GGTAAGTGTG GCCGTACTAC
~~~~~

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CM(R)
 3351 AACCTGAATC GCCAGCGGCA TCAGCACCTT GTCGCCTTGC GTATAATATT
 TTGGACTTAG CGGTCGCCGT AGTCGTGGAA CAGCGGAACG CATATTATAA
 ~~~~~  
 CM(R)  
 3401 TGCCCATAGT GAAAACGGGG GCGAAGAAGT TGTCCATATT GGCTACGTTT  
 ACGGGTATCA CTTTGGCCCC CGCTTCTTCA ACAGGTATAA CCGATGCAAA  
 ~~~~~  
 CM(R)
 3451 AAATCAAAAC TGGTGAAACT CACCCAGGGA TTGGCTGAGA CGAAAAACAT
 TTAGTTTTG ACCACTTTGA GTGGGTCCCT AACCGACTCT GCTTTTTGTA
 ~~~~~  
 CM(R)  
 3501 ATTCTCAATA AACCTTTAG GGAAATAGGC CAGGTTTTCA CCGTAACACG  
 TAAGAGTTAT TTGGGAAATC CCTTATCCG GTCCAAAAGT GGCATTGTGC  
 ~~~~~  
 CM(R)
 3551 CCACATCTTG CGAATATATG TGTAAGAACT GCCGGAAATC GTGCTGGTAT
 GGTGTAGAAC GCTTATATAC ACATCTTTGA CGGCCTTTAG CAGCACCATA
 ~~~~~  
 CM(R)  
 3601 TCACTCCAGA GCGATGAAAA CGTTTCAGTT TGCTCATGGA AAACGGTGTA  
 AGTGAGGTCT CGCTACTTTT GCAAAGTCAA ACGAGTACCT TTTGCCACAT  
 ~~~~~  
 CM(R)
 3651 ACAAGGGTGA AACTATCCC ATATCACCAG CTCACCGTCT TTCATTGCCA
 TGTTCCTACT TGTGATAGGG TATAGTGGTC GAGTGGCAGA AAGTAACGGT
 ~~~~~  
 CM(R)  
 3701 TACGGAATC CGGGTGAGCA TTCATCAGGC GGGCAAGAAT GTGAATAAAG  
 ATGCCTTGAG GCCCACTCGT AAGTAGTCCG CCCGTTCTTA CACTTATTC  
 ~~~~~  
 CM(R)
 3751 GCCGATAAAA ACTTGTGCTT ATTTTCTTT ACGGTCTTTA AAAAGGCCGT
 CGGCCTATTT TGAACACGAA TAAAAAGAAA TGCCAGAAAT TTTCCGGCA
 ~~~~~  
 CM(R)  
 3801 AATATCCAGC TGAACGGTCT GGTATAGGT ACATTGAGCA ACTGACTGAA  
 TTATAGGTCG ACTTGCCAGA CCAATATCCA TGTAACCTCGT TGAAGTACTT  
 ~~~~~  
 CM(R)
 3851 ATGCCTCAAA ATGTTCTTTA CGATGCCATT GGGATATATC AACGGTGGTA
 TACGGAGTTT TACAAGAAAT GCTACGGTAA CCCTATATAG TTGCCACCAT
 ~~~~~  
 CM(R)  
 3901 TATCCAGTGA TTTTTTCTC CATTTTAGCT TCCTTAGCTC CTGAAAACT  
 ATAGGTCACT AAAAAAGAG GTAAAATCGA AGGAATCGAG GACTTTTAGA  
 ~~~~~  
 CM(R) SD
 ~~~~~  
 cat promoter  
 3951 CGATAACTCA AAAAATACGC CCGGTAGTGA TCTTATTCA TTATGGTGAA  
 GCTATTGAGT TTTTATGCG GGCCATCACT AGAATAAAGT AATACCACTT  
 ~~~~~  
 cat promoter
 CRP site
 ~~~~~  
 4001 AGTTGGAACC TCACCCGACG TCTAATGTGA GTTAGCTCAC TCATTAGGCA  
 TCAACCTTGG AGTGGGCTGC AGATTACACT CAATCGAGTG AGTAATCCGT  
 ~~~~~  
 cat promoter
 lac mRNA start

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                                lac operator
                                ~~~~~
                        -35 region      -10 region
                        ~~~~~          ~~~~~
4051  CCCCAGGCTT TACACTTTAT GCTTCCGGCT CGTATGTTGT GTGGAATTGT
      GGGGTCCGAA ATGTGAAATA CGAAGGCCGA GCATACAACA CACCTTAACA
      lac operator      SD      lacZ'
      ~~~~~          ~~~~~          ~~~~~
4101  GAGCGGATAA CAATTTTACA CAGGAAACAG CTATGACCAT GATTACGAAT
      CTCGCCTATT GTTAAAGTGT GTCCTTTGTC GATACTGGTA CTAATGCTTA
      lacZ'
      ~
4151  T
      A

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Position34/39

35/39

36/39

NSPV

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